

Fig. S1

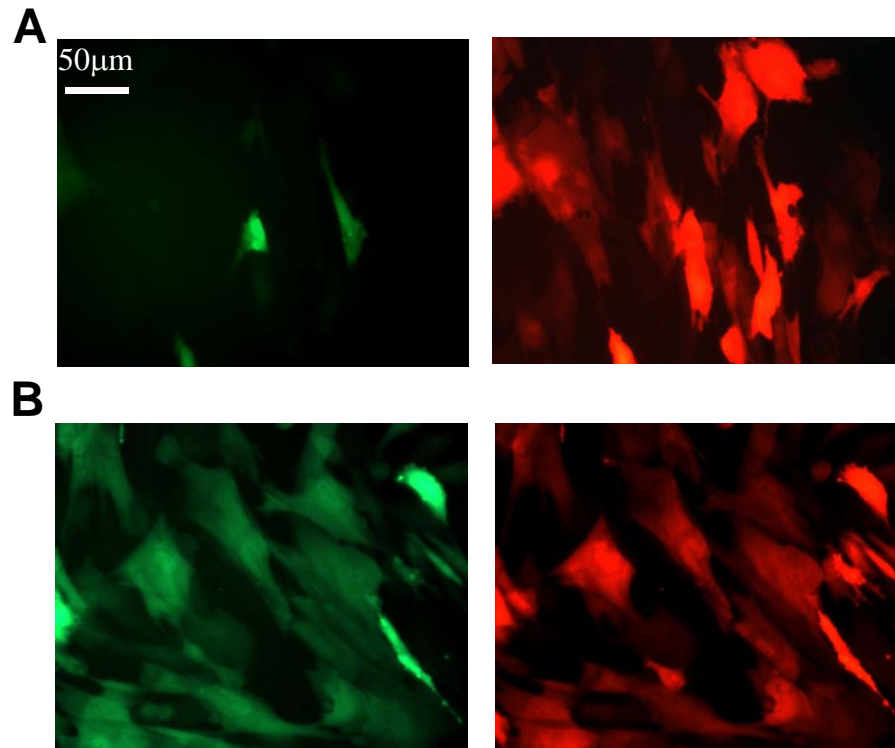


Fig.S2

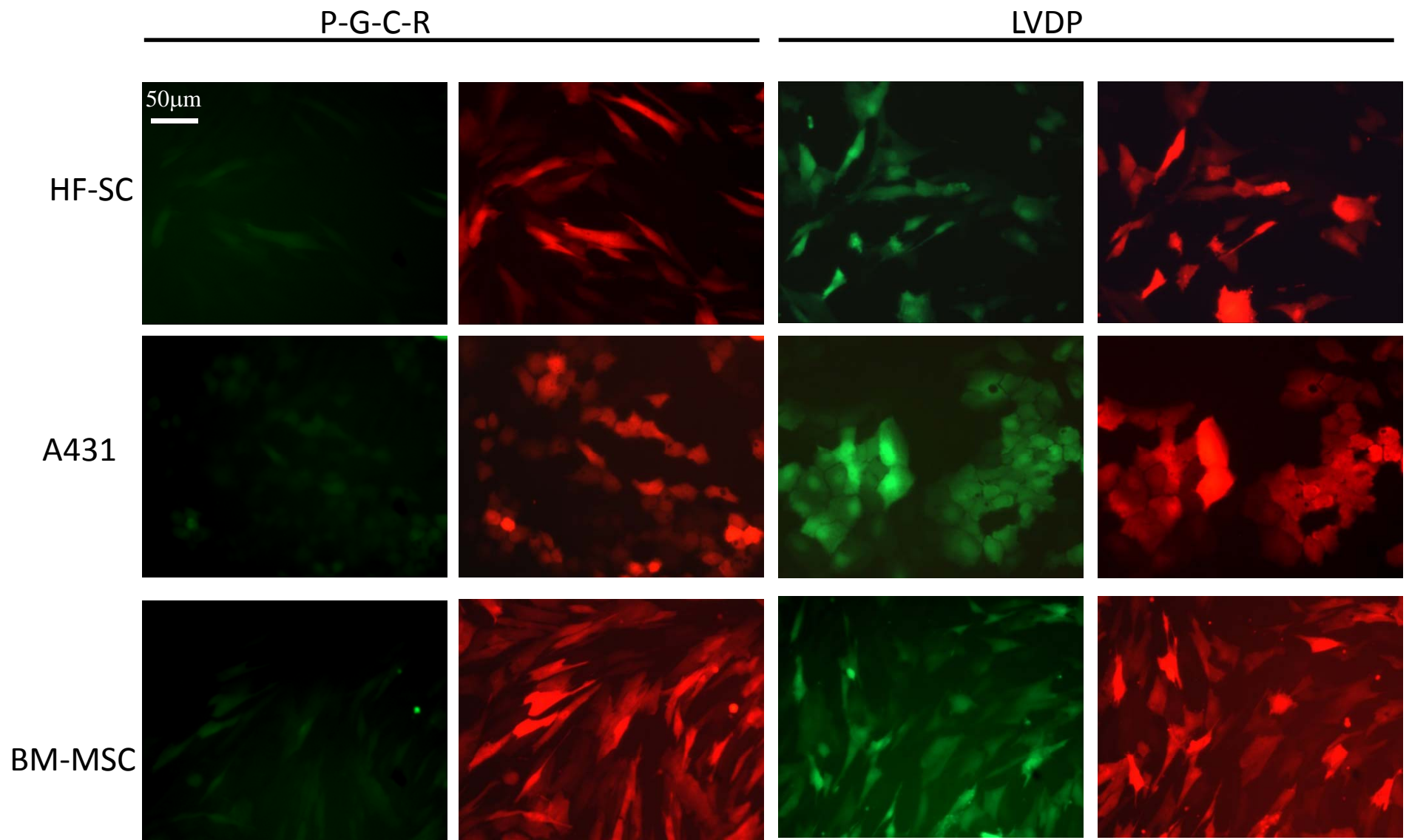


Fig.S3

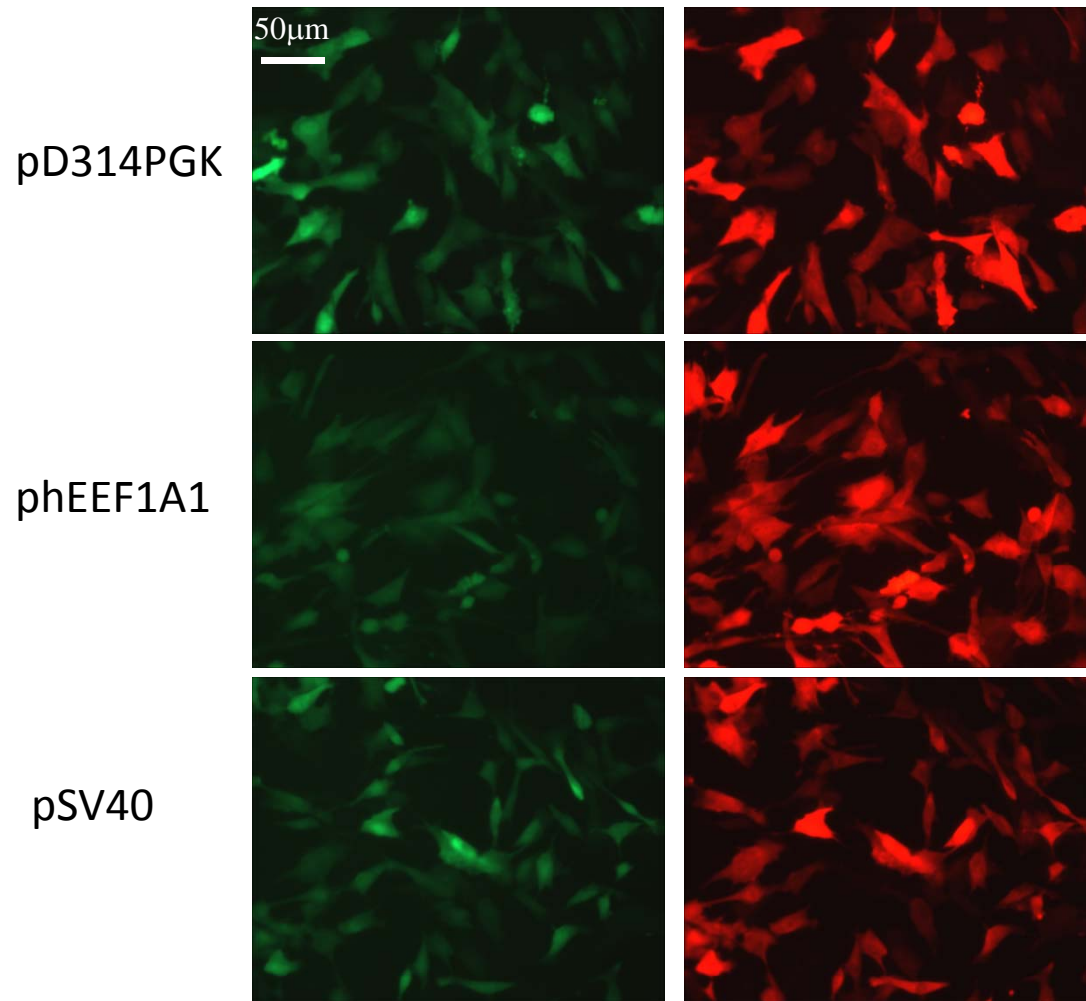


Fig.S4

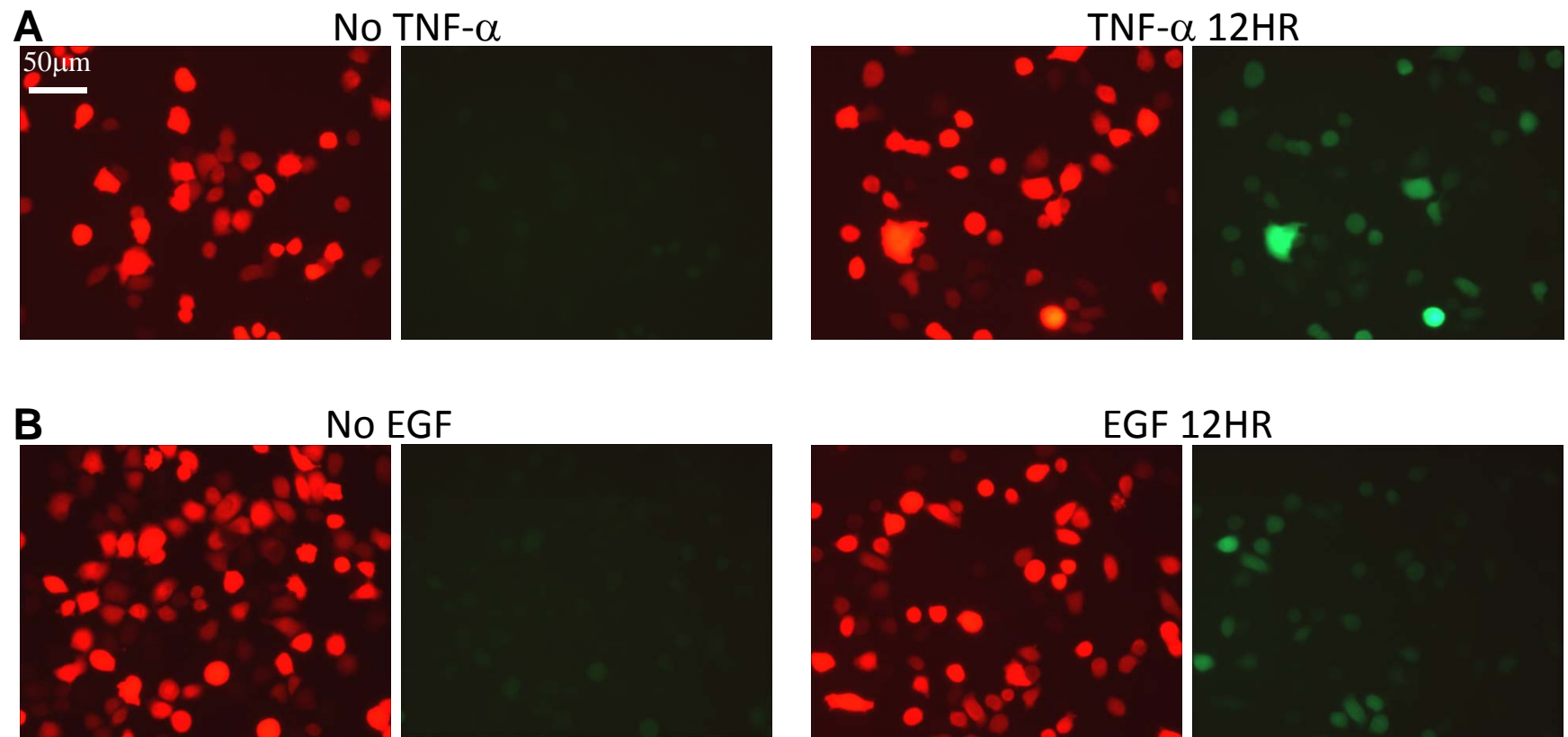
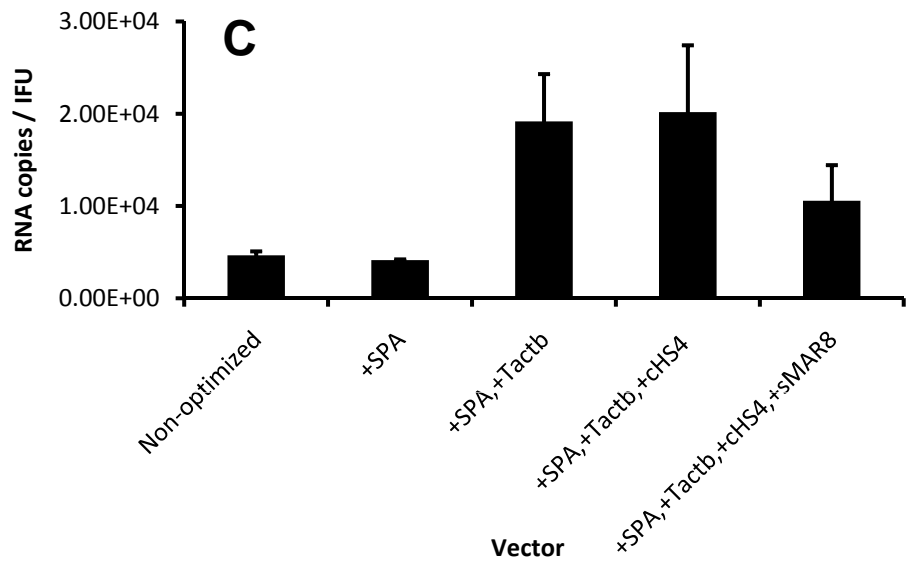
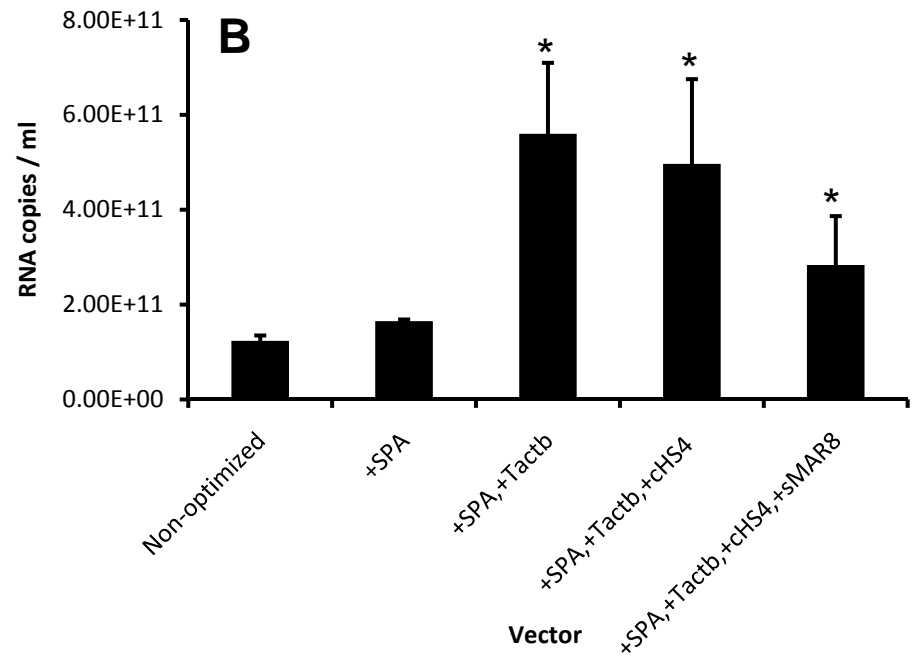
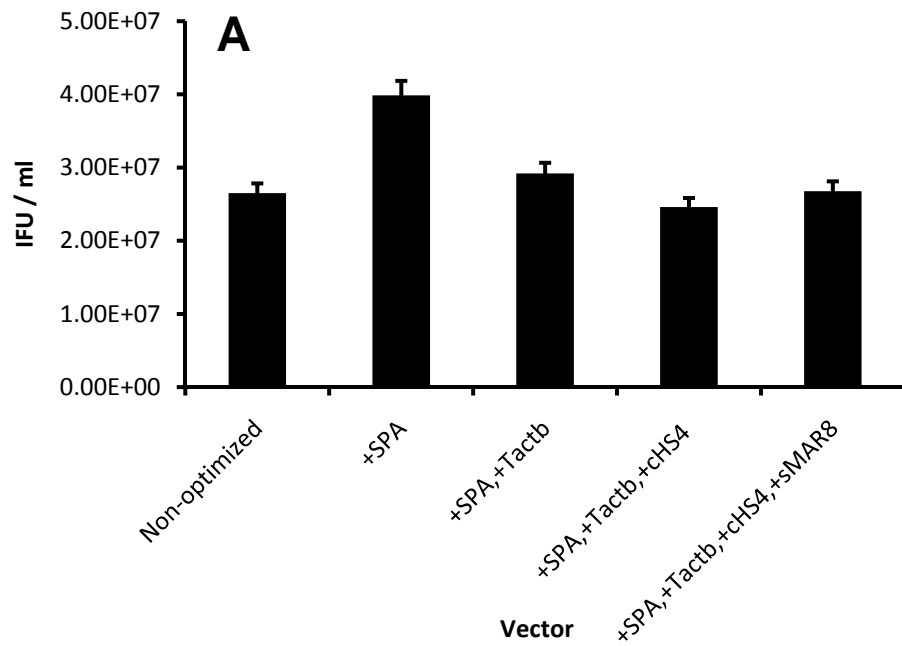


Fig.S5



Purpose	Vector #	Parental vector	Insert	Cloning sites	Primers/Oligos	Source of inserts	Remarks
Parental vectors	1	pCSCG	pA and multiple cloning site (MCS)	BamHI, XhoI	pAMCSF :TCGAAGAATTACCGGGAATCGATAACCGTAGGCGGCCAGGATCCACTCGAGAATGACGGCAATAAAAAGACAGATAAAAACCCACGGGTGTTGGGTCGTTTGTTCATAAACCGCGGA; pAMCSR :GATCTCCGGTTTATGAACAACACGCCAACACCCGTGGGTTTTATTCTGTCTTTTATTGCCGTCATTCTCGAGTGGATCTGGCGCGCTACC GGTTATCGATTCCGCGGTGAATCT	oligos	the oligoes were annealed and ligated into pCSCG to replace CMV-GFP segment in the original vector. After ligation, multiple cloning sites (EcoRI, SacI, ClaI, AgeI, AscI, BamHI, XhoI) and the core sequence of Herplex Simplex Virus (HSV) thymidine kinase polyA were introduced.
	2	1	CMV_GFP	BamHI, XhoI		subclone from pCSCG	subclone
	3	2	DsRed2	NheI, XhoI	NheI_DsRed2F:AACAACGCTAGCTCGCCACCATGGCCCTCCTC; XhoI_DsRed2R:TGTTGTCTCGAGGGGGGAGGTGGGAGGTTT	PCR from pDsRed2-1	DsRed2 was amplified with PCR and cloned into vector 2 to replace GFP
	4	3	ZsG	BamHI, AgeI	BamHI_ZsGR:TGGAGGGATCCTCTAGAAGTTAGGGCAAGCGGGA; AgeI_ZsGF:ACAACAACCGGTATCCGCCACCATGGCCACGT	PCR from pSIREN-RetroQ-ZsG	ZsG was amplified with PCR from pSIREN-RetroQ-ZsG vector and cloned into vector 3. Note that a cloning site XbaI was created upstream of BamHI
	5	4	phPGK	ClaI, AgeI	ClaI_hPGKF :ACAACAATCGATAATTCACGGGTTGGGGTT; AgeI_hPGKR :TGTTGTACCGGTTACAGCTGGGGAGAGAGTTC	PCR from hgDNA	phPGK was amplified from hgDNA
Clone polyAs downstream of vector #5	6	5	hGHPA	XbaI, BamHI	XbaI_hGHPAF :ACAACATCTAGACAGTCCGCTCTGTGGAGG; BamHI_hGHPAR :TGTTGTGGATCCGGCATGGCCAGGTAGCCTAT	PCR from hgDNA	hGHPA was amplified from hgDNA
	7	5	rGbpA	XbaI, BamHI	XbaI_rGbpAF :ACATGTTCTAGAGGGCAACGTGCTGTTGTTG; BamHI_rGbpAR :TGTTGTGGATCCAGAGAAGAGGGACAGCTATG	PCR from pC4S1-FM4-FCS-hGH	rGbpA was amplified from vector pC4S1-FM4-FCS-hGH
	8	5	SPA	XbaI, BamHI	XbaI_SPAF :CTAGCAATAAAAAGATCTTTATTTTCATTAGATCTGTGTGGTTTTTTGTGTCTAGAG; BamHI_SPAR :GATCCTCTAGACACAAAAACCAACACACAGATCTAATGAAAAATAAGATCTTTTATTG	oligos	the oligoes of SPA were annealed and ligated into vector 5. Note that after ligation the original XbaI was lost and a new XbaI was created downstream of SPA for cloning other sequences.
Clone terminators/pause sites downstream of SPA using vector #8	9	8	S	XbaI, BamHI	BamHI_SF :ACAACAGGATCCAGGCGTAAATGTAAGCGT; XbaI_SR :TGTTGGGATCCCCCTGAGCGGCGATTAAG	PCR from pEGFP-1	the spacer sequence was amplified by PCR from vector pEGFP-1
	10	8	CoTC	XbaI, BamHI	XbaI_CoTCF :ACAACATCTAGAATGTCCCTTATGGTGCTTCT; BamHI_CoTCR :ACAACAGGATCCGTGTTAATTAATTTGGCCTTTGCTCTCAT	PCR from hgDNA	the CoTC sequence was amplified by PCR from hgDNA. Note a PacI site was created downstream of CoTC for cloning other sequences.
	11	8	T _{actb}	XbaI, BamHI	XbaI_T_{actb}F :ACAACATCTAGAGTGGGGCAGTGGGGCCAA; BamHI_T_{actb}R :ACAACAGGATCCACATTAATTAAGTACAGCCAGTATCCATA	PCR from hgDNA	the CoTC sequence was amplified by PCR from hgDNA. Note a PacI site was created downstream of T _{actb} for cloning other sequences.
	12	8	MAZ4	XbaI, BamHI	XbaI_MAZ4F :CTAGAGCCTTGGGGAGGGGAGGCCAGAAGCCCTGGGGAGGGGAGGCCAGAAATTAATGTTG; BamHI_MAZ4R :GATCCACATTAATTAATTTGGCCTCCCTCCCTCCCAAGGCCTTGGCCTCCCTCCCTCCCAAGGGCT	oligos	the oligoes for MAZ4 were annealed and ligated into vector 8. Note a PacI site was created downstream of MAZ4 for cloning other sequences.
	13	8	T _{msa}	XbaI, BamHI	XbaI_T_{msa}F :ACAACATCTAGACTCCCTCTGTCGCCACACT; BamHI_T_{msa}R :ACAACAGGATCCACATTAATTAACCTCTCAAAAACAGAGCTCACT	PCR from mouse gDNA	the CoTC sequence was amplified by PCR from mouse gDNA. Note a PacI site was created downstream of T _{msa} for cloning other sequences.
Clone insulators downstream of Tactb using vector #11	14	11	sMAR4	PacI, BamHI	PacI_sMAR4F :GATCCACATTAATTAATTTCTAAATATATTAGAAATTAAGAATCTAAATATATTAGAAATTAAGATTCTAAATATATTAGAAATTAAGATTCTAAATATATTAGAAATTAAGAAAT; BamHI_sMAR4R :TCTTTAATTTCTAATATATTAGAATCTTTAATTTCTAATATATTAGAATCTTTAATTTCTAATATATTAGAATCTTTAATTTCTAATATATTAGAATTAATGTTG	oligos	the oligos of sMAR4 were annealed and ligated into vector 11. Note that after ligation the original PacI was lost and a new PacI was created downstream of sMAR4 for cloning other sequences.
	15	14	sMAR8	PacI, BamHI	PacI_sMAR4F :GATCCACATTAATTAATTTCTAAATATATTAGAAATTAAGAATCTAAATATATTAGAAATTAAGATTCTAAATATATTAGAAATTAAGAAAT; BamHI_sMAR4R :TCTTTAATTTCTAATATATTAGAATCTTTAATTTCTAATATATTAGAATCTTTAATTTCTAATATATTAGAATTAATGTTG	oligos	similar to cloning vector 14, another copy of sMAR4 was cloned.
	16	11	cHS4	PacI, BamHI	PacI_cHS4F :ACAACATTAATTAACCTTCCCGGGAGCTCACGG; BamHI_cHS4R :TGTTGGGATCCGTGACGCACCTGAACAGGTTG	PCR from chicken gDNA	cHS4 sequence was amplified from chicken gDNA
	17	15	cHS4	PacI, BamHI	PacI_cHS4F :ACAACATTAATTAACCTTCCCGGGAGCTCACGG; BamHI_cHS4R :TGTTGGGATCCGTGACGCACCTGAACAGGTTG	PCR from chicken gDNA	cHS4 sequence was amplified from chicken gDNA
	18	pCSCG	New MCS	BamHI, XhoI	NpAMCSF :TCGAAGAATTACCTCGAGAATGACGGCAATAAAAAGACAGAAATAAACCCACGGGTGTTGGGTCGTTTGTTCATAAACCGGGAAGCGCTACGGTCCGAGTTAACA; NpAMCSR :GATCTGTTAACTCGGACCGTAGCGCTCCCGCTTTATGAACAAACGACCAACACCCGTGGGTTTATCTGTCTTTTATTGCCGTCATTCTGAGTGAATCT	oligos	After ligation, original BamHI and XhoI were. Note new MCS (AfeI, RsrII, XhoI, EcoRI, HpaI) and the same TKpA as in vector 1 were introduced.
	19	18	M9	AfeI, RsrII	AfeI_M9F :ATTTCATATAGAACAATGTAATTTCTTCTAGAGTTAAGCAGGGCTTTTATAAGCTGAAGCGCTACG; RsrII_M9R :GACCGTAGCGCTTACGTTATGAAAAAGCCCTGCTTAACCTTAGGAAAGAAITTACATITGTTCTATATGGAAT	oligos	oligos for M9 were annealed and ligated into vector 18

Clone insulators downstream of TKpA	20	18	m3'HS1	AfeI, RsrII	AfeI_m3'HS1F: TCATTTCTCTAATGATCCTGTTGCATACCAGTAGGGGGCAGAAGTGTCC ACTGATTTCCGCCCTCCTCCAGCGCTACG; RsrII_m3'HS1R: GACCGTAGCGCTGGAGAGGAGGGCGGAAATCAGTGGAA CACTTCTGCCCCCTACTGGTATGCAACAGGATCATTAGAGAAATGA	oligos	oligos for m3'HS1 were annealed and ligated into vector 18
	21	18	H19HS2	AfeI, RsrII	RsrII_H19HS2F: ACAACACCGTCCGCCAGCAGTGTGGCTCACTA; AfeI_H19HS2R: ACAACAAGCGTGTCTGCCGAGCAATATGTAG	PCR from mouse gDNA	insulator H19HS2 was amplified from mouse gDNA
	22	19	DsRed2_CMV_cHS4_Tactb_SPA_ZsG_phPGK	EcoRI, XhoI		subclone from vector 16	The insert DsRed2_CMV_cHS4_Tactb_SPA_ZsG_phPGK was cut from vector 16 and subcloned into vector 19
	23	20	DsRed2_CMV_cHS4_Tactb_SPA_ZsG_phPGK	EcoRI, XhoI		subclone from vector 16	The insert DsRed2_CMV_cHS4_Tactb_SPA_ZsG_phPGK was cut from vector 16 and subcloned into vector 20
	24	21	DsRed2_CMV_cHS4_Tactb_SPA_ZsG_phPGK	EcoRI, XhoI		subclone from vector 16	The insert DsRed2_CMV_cHS4_Tactb_SPA_ZsG_phPGK was cut from vector 16 and subcloned into vector 21
	25	pCSCG	sMAR4	BamHI, XhoI	XhoI_sMAR4F: TCGAGTGTGGATCCTTCTAAATATATTAGAATTAAGAT TCTAAATATATTAGAAATTAAGATCTAAATATATTAGAAATTAAGAT CTAAATATATTAGAAATTAAGAT; BamHI_sMAR4R: GATCATCTTTAATTCTAATATATTAGAATCTTTAATT CTAATATATTAGAAATCTTTAATTCTAATATATTAGAAATCTTTAATTCT AATATATTAGAAAGATCCACAC	oligos	The oligos for sMAR4 were annealed and ligated into pCSCG to replace CMV-GFP segment in the original vector. After ligation, the original BamHI was lost but a new BamHI was created upstream of sMAR4
	26	25	sMAR4	BamHI, XhoI		same oligos as used in vector 25	similar to cloning vector 25, another copy of sMAR4 was cloned.
	27	26	New MCS	BamHI, XhoI		same oligos as used in vector 1	similar to cloning vector 1, HSV TKpA together with multiple cloning sites was cloned.
	28	27	DsRed2_CMV_cHS4_Tactb_SPA_ZsG_phPGK	EcoRI, XhoI		subclone from vector 16	The insert DsRed2_CMV_cHS4_Tactb_SPA_ZsG_phPGK was cut from vector 16 and subcloned into vector 27
Use EGFP to replace ZsGreen	29	28	SPA-EGFP	AgeI, XbaI	AgeI_EGFP_SPAF: ACAACACTCTAGACACACAAAAACACACAGAT CTAATGAAAATAAAGATCTTTATTGCTACCGCTTACTTGTACAGCTC; XbaI_EGFP_SPAR: ACAACACTCTAGACCGCTTACTTGTACAGCTC	PCR from pEGFP-1	EGFP was amplified from pEGFP-1 with primer containing SPA and cloned into vector 29 to replace ZsG-SPA
Clone promoters in place of PGK	30	27	DsRed2_CMV_cHS4_Tactb_SPA_EGFP	XhoI, AgeI		subclone from vector 29	The insert DsRed2_CMV_cHS4_Tactb_SPA_EGFP was cut from vector 29 and subcloned into vector 27
	31	30	phEEF1A1	EcoRI, ClaI	EcoRI_hEEF1A1F: ACAACAGAATTCGGAGAAGAGCATCGGTGAGG ClaI_hEEF1A1R: ACAACAATCGATAGGGGTAGTTTTACAGACAC	PCR from hgDNA	phEEF1A1 was amplified from hgDNA
	32	30	phD314PGK	ClaI, AgeI	ClaI_hd314PGKF: ACAACAATCGATTGGCTGTGGCCAATAGCGG; AgeI_hpGKR: TGTTGTACCGGTTACAGCTGGGAGAGAGGTC	PCR from hgDNA	phD314PGK was amplified from hgDNA
	33	30	pSV40	EcoRI, AgeI	EcoRI_pSV40F: ACAACAGAATTCCTGAGGCGGAAAGAACCCAGC; AgeI_pSV40R: TGTTGTACCGGTCGATCCTCATCTGTCTCTT	PCR from pd2EGFP-1	pSV40 was amplified from pEGFP-1
	34	30	pACTA2	ClaI, AgeI	ClaI_ACTA2F: ACAACAATCGATAACAGCTGGTCAATGGTGTGA; AgeI_ACTA2R: TGTTGTACCGGTCGATGAACCCAGCCAAATCC	PCR from hgDNA	pACTA2 was amplified from hgDNA
use d2EGFP to replace EGFP	35	30	SPA-d2EGFP	AgeI, XbaI	AgeI_d2EGFP: ACAACAACCGGTGGTCCACCATTGGTGAGCA; XbaI_SPA_d2EGFP: ACAACATCTAGACACACAAAAACACACAGAT CTAATGAAAATAAAGATCTTTATTGCTAGAGCCATCTACACATTGATC	PCR from pd2EGFP-1	SPA-d2EGFP was amplified from pd2EGFP-1 with primer containing SPA and cloned into vector 30 to replace EGFP-SPA
Clone response elements	36	35	RE-NFKB	ClaI, AgeI		subclone from pRE-NFKB_CMVmini_d2EGFP	RE-NFKB together with CMV _{mini} was cut from vector pRE-NFKB_CMV _{mini} _d2EGFP and cloned into vector 35
	37	35	RE-API	ClaI, AgeI		subclone from pRE-API_CMVmini_d2EGFP	RE-NFKB together with CMV _{mini} was cut from vector pRE-API_CMV _{mini} _d2EGFP and cloned into vector 35

Note: all genomic DNA used was obtained from EMD (Gibbstown, NJ)