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SUPPORTING INFORMATION

<u>Title:</u> Regioselective Domino Metathesis of 7-Oxanorbornenes and Its Application to the Synthesis of Biologically Active Glutamate Analogues <u>Author(s):</u> Minoru Ikoma, Masato Oikawa,* Martin B. Gill, Geoffrey T. Swanson, Ryuichi Sakai, Keiko Shimamoto, Makoto Sasaki <u>Ref. No.:</u> O200800704

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Bn⁻⁻ŅH PMB Bn ŅӉ <mark>Ӊ</mark> рмв 0 → AcO //////_ 0 =0 ЧΗ Ò H' Ĥ 3 **8** (87%, *E*/*Z* = 13:1)

General Methods. All reactions sensitive to air or moisture were carried out under argon atmosphere in oven-dried glassware unless otherwise noted. Anhydrous dichloromethane (CH₂Cl₂), acetonitrile (MeCN) and tetrahydrofuran (THF) were purchased from Kanto Chemical Co., Inc., Wako Pure Chemical Industries, Ltd. or Aldrich Chemical Co., and used without further drying. All other reagents were used as supplied unless otherwise stated. Analytical thin-layer chromatography (TLC) was performed using E. Merck silica gel 60 F₂₅₄ pre-coated plates (0.25-mm thickness). For column chromatography, Fuji Silysia silica gel BW-300 (200-400 mesh) was used. For reversed-phase column chromatography, Wako gel [®]100C18 (63-212 μm) was used. IR spectra were recorded on a JASCO FT/IR-4100 spectrometer. ¹H and ¹³C NMR spectra were recorded on Varian Gemini2000, Unity INOVA500 or INOVA600 spectrometers. Chemical shifts are reported in ppm from tetramethylsilane with reference to internal residual solvent [¹H NMR: CHCl₃ (δ 7.24), C₆HD₅ (δ 7.15), HDO (δ 4.70); ¹³C NMR: CDCl₃ (δ 77.0), C₆D₆ (δ 128.0)]. The following abbreviations are used to designate the multiplicities; s = singlet, d = doublet, t = triplet, q =quartet, m = multiplet, and br = broad peak. High-resolution mass spectrometry (HRMS) for ESI (electrospray ionization) was recorded on a Bruker Daltonics microTOF focus mass spectrometer equipped with Agilent HPLC system (1100 series).

N-(4-Methoxybenzyl)furfurylamine (S1). To a stirred solution of 4-methoxybenzylamine (6.5 mL, 50 mmol) in benzene (100 mL) was added furfural (5.0 mL, 50 mmol) and the mixture was heated to reflux with Dean-Stark condenser. After 12 h, the mixture was cooled to rt and concentrated under reduced pressure. To a stirred solution of the residue in methanol (50 mL) at 0 °C were added NaBH₄ (3.0 g, 74 mmol) in one portion and TFA (4.1 mL, 55 mmol) successively. After 1 h, the mixture was concentrated under reduced pressure, diluted with EtOAc (200 mL), washed with aqueous NaOH (2 M, 100 mL) and brine (100 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (20 g, hexane/EtOAc = 7:3) to give *N*-(4-methoxybenzyl)furfurylamine (**S1**, 10.4 g, 97%) as a colorless oil; IR (film) 3324, 1512, 1456, 1247, 1174, 1035, 919, 813, 737 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.35 (dd, *J* = 1.8 Hz, 0.9 Hz, 1 H), 7.22 (d, *J* = 8.7 Hz, 2 H), 6.84 (d, *J* = 8.7 Hz, 2 H), 6.30 (dd, *J* = 3.0, 1.8 Hz, 1 H), 6.16 (d, *J* = 2.4 Hz, 1 H), 3.78 (s, 3 H), 3.75 (s, 2 H), 3.70 (s, 2 H); ¹³C NMR (125 MHz, CDCl₃) δ 158.5, 153.7, 141.6, 131.8, 129.3, 113.6, 109.9, 106.8, 55.0, 52.0, 45.1; HRMS (ESI, positive) calcd for C₁₃H₁₆NO₂ [(M+H)⁺] 218.1176, found 218.1182.

The lodoacrylamide S2. To a stirred solution of *N*-(4-methoxybenzyl)furfurylamine (**S1**, 596 mg, 2.77 mmol) in THF (30 mL) at 0 °C were added (*Z*)-3-iodoacryl chloride (629 mg, 2.92 mmol)¹ and Cs_2CO_3 (1.43 mg, 4.38 mmol), and allowed to warm to rt. After 3 h, the mixture was poured into saturated aqueous NH₄Cl (30 mL) and extracted with EtOAc (50 mL). Organic layer was washed with brine (50 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (20 g, hexane/EtOAc = 8:2) to give the iodoacrylamide **S2** (840 mg, 74%) as a pale yellow oil, which was unstable and used immediately for the next reaction without characterization.

The 7-Oxanorbornene 5. A solution of the iodoacrylamide **S2** (268 mg, 0.65 mmol) in toluene (7.0 mL) was heated to reflux. After 12 h, the mixture was cooled to rt and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (20 g, hexane/EtOAc = 2:8) to give **5** (232 mg, 87 %) as a white solid; IR (film) 2932, 1684, 1512, 1246, 1175, 1032, 707 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.19 (d, *J* = 8.7 Hz, 2 H), 6.86 (d, *J* = 8.7 Hz, 2 H), 6.50 (d, *J* = 5.7 Hz, 1 H), 6.34 (dd, *J* = 5.7, 2.1 Hz, 1 H), 5.27 (d, *J* = 1.2 Hz, 1 H), 4.54 (d, *J* = 14.4 Hz, 1 H), 4.42 (d, *J* = 14.4 Hz, 1 H), 3.86 (d, *J* = 8.1 Hz, 1 H), 3.78 (s, 3 H), 3.70 (d, *J* = 12.0 Hz, 1 H), 3.49 (d, *J* = 12.0 Hz, 1 H), 2.43 (d, *J* = 7.5 Hz, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ 169.9, 159.0, 136.1, 135.3, 129.3, 127.7, 114.1, 90.3, 88.7, 55.2, 50.1, 47.2, 46.2, 18.4; HRMS (ESI, positive) calcd for C₁₆H₁₇INO₃ [(M+H)⁺]

¹ Prepared from (*Z*)-3-iodoacrylic acid by treatment with oxalyl chloride and DMF in benzene. For the synthesis of (*Z*)-3-iodoacrylic acid, see: R. Takeuchi, K. Tanabe, S. Tanaka, *J. Org. Chem.* **2000**, *65*, 1558-1561.

398.0248, found 398.0252.

The Allyl Ether 6. To a stirred solution of allyl alcohol (20.6 μL, 0.30 mmol) in DMF (1.0 mL) at rt was added NaH (60% in mineral oil, 12 mg, 0.3 mmol). After 30 min, a solution of 5 (19.9 mg, 0.05 mmol) in DMF (1.0 mL) was added via a cannula, and the mixture was stirred at -10 °C for 1.5 h. The mixture was then poured into saturated aqueous NH₄CI (10 mL) and was extracted with EtOAc (20 mL). The extract was washed with water (3 \times 10 mL) and brine (10 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (0.5 g, hexane/EtOAc = 5:5) to give the allyl ether 6 (633 mg, 73%) as a yellow solid: IR (film) 2910, 1686, 1513, 1470, 1247, 1112, 1032, 839, 720 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.14 (d, J = 8.5 Hz, 1 H), 6.84 (d, J = 8.5 Hz, 1 H), 6.51 (d, J = 6.0 Hz, 1 H), 6.42 (dd, J = 11.5, 2.0 Hz, 1 H), 5.89 (ddd, J = 16.8, 11.0, 10.5 Hz, 1 H), 5.31 (dd, J = 17.0, 1.5 Hz, 1 H), 5.19, (dd, J = 10.5, 1.5 Hz, 1 H), 4.99 (dd, J = 4.5, 2.0 Hz, 1 H), 4.50 (d, J = 14.5 Hz, 1 H), 4.35 (d, J = 14.5 Hz, 1 H), 4.35 (d, J = 5.0, 2.0 Hz, 1 H), 4.13 (dd, J = 12.5, 5.5 Hz, 1 H), 4.05 (dd, J = 12.5, 5.5 Hz, 1 H), 3.77 (s, 3 H), 3.75 $(d, J = 11.5 Hz, 1 H), 3.42 (d, J = 11.5 Hz, 1 H), 2.23 (d, J = 2.0 Hz, 1 H); {}^{13}C NMR (125 MHz, 1 H);$ CDCl₃) § 172.1, 159.1, 134.8, 133.9, 129.3, 128.0, 117.7, 114.1, 90.1, 79.6, 78.3, 71.6, 55.3, 55.2, 49.2, 46.0; HRMS (ESI, positive) calcd for C₁₉H₂₂NO₄ [(M+H)⁺] 328.1549, found 328.1542.

Metathesis Reaction of 5. To a stirred solution of the 7-oxanorbornene **5** (50.0 mg, 0.122 mmol) in DCM (3.0 mL) at rt were added vinyl acetate (56.7 μ L, 0.611 mmol) and Hoveyda-Grubbs catalyst 2nd generation (3.80 mg, 6.11 μ mol) under argon atmosphere. After stirring for 14 h, the mixture was concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (4 g, hexane/EtOAc = 7:3) to give the heterobicycle **10** (a brown oil, 18.5 mg, 31%) as a mixture four diastereomers as judged from LC-MS and ¹H NMR spectra.

The Heterotricycle 11. To a stirred solution of the 7-oxanorbornene **6** (295.3 mg, 0.86 mmol) in DCM (331 mL) at rt were added vinyl acetate (2.7 mL, 4.3 mmol) and Hoveyda-Grubbs catalyst 2^{nd} generation (2.7 mg, 4.3 µmol) under argon atmosphere. After 6 h, the mixture was concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (4 g, hexane/EtOAc = 6:4) to give the heterotricycles **11** (291.6 mg, 88%, *E/Z* = 5:4) and **12** (29.5 mg, 10%) as colorless oils.

Data for 11. IR (film) 2935, 1757, 1684, 1513, 1247, 1213, 1088, 1035, 818, 763 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.33 (d, *J* = 12.2 Hz, 1.3 H), 7.16 (d, *J* = 9.0 Hz, 2 H), 7.13 (d, *J* = 9.0 Hz, 2 H), 6.97 (d, *J* = 7.0 Hz, 1 H), 6.85 (d, *J* = 9.0 Hz, 2.6 H), 6.83 (d, *J* = 9.0 Hz, 2 H), 6.05 (dd, *J* = 10.8, 3.0 Hz, 2.6 H), 5.96 (m, 1 H), 5.57 (d, *J* = 12.3 Hz, 1.3 H), 5.27 (d, *J* = 7.0

Hz, 1 H), 4.51-4.45 (m, 2.3 H), 4.37-4.31 (m, 4.6 H), 4.19-3.96 (m, 6.9 H), 3.78 (s, 3.9 H), 3.77 (s, 3 H); 13 C NMR (125 MHz, CDCl₃) δ 170.3, 170.2, 167.7, 166.5, 159.2, 159.2, 137.4, 133.1, 131.4, 131.0, 129.6, 129.5, 127.9, 127.6, 122.0, 121.9, 116.5, 115.6, 114.2, 114.1, 85.4, 82.9, 82.5, 78.6, 76.6, 73.4, 72.6, 64.4, 64.1, 60.5, 59.9, 58.6, 57.2, 55.3, 46.1, 46.0, 20.6, 20.5; HRMS (ESI, positive) calcd for C₁₉H₂₂NO₄ [(M+H)⁺] 386.1604, found 386.1602.

Data for 12. IR (film) 2935, 1684, 1512, 1246, 1090, 1032, 764 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.13 (d, *J* = 8.4 Hz, 2 H), 6.82 (d, *J* = 8.4 Hz, 2 H), 6.05 (dd, *J* = 10.1, 3.6 Hz, 1 H), 5.98 (m, 1 H), 5.96 (dd, *J* = 16.7, 10.5 Hz, 1 H), 5.29 (d, *J* = 16.7 Hz, 1 H), 5.28 (s, 1 H), 5.11 (d, *J* = 10.5 Hz, 1 H), 4.49 (d, *J* = 14.7 Hz, 1 H), 4.34 (d, *J* = 3.0 Hz, 1 H), 4.33 (d, *J* = 14.7 Hz, 1 H), 4.15 (dd, *J* = 16.4, 3.9 Hz, 1 H), 4.08-4.00 (m, 2 H), 3.78 (s, 3 H), 3.42 (d, *J* = 11.1 Hz, 1 H), 3.31 (d, *J* = 11.1 Hz, 1 H), 3.15 (br s, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ 170.4, 159.2, 138.6, 131.0, 129.4, 127.7, 122.1, 114.9, 84.9, 78.7, 73.3, 64.2, 59.8, 57.0, 55.3, 46.0; HRMS (ESI, positive) calcd for C₁₉H₂₂NO₄ [(M+Na)⁺] 328.1543, found 328.1550.

The 7-Oxanorbornene 3 (Tandem Ugi/Diels-Alder Reaction). To a stirred solution of furfural (500 µL, 5.31 mmol) in methanol (25 mL) at rt were added 4-methoxybenzylamine (459 µL, 3.54 mmol), (Z)-iodoacrylic acid (750 mg, 3.54 mmol), and benzyl isocyanide (647 μ L, 5.31 mmol). After stirring at 50 °C for 8.5 h, the mixture was concentrated under reduced pressure and the residue was diluted with chloroform (200 mL). The solution was washed with saturated aqueous NaHCO₃ (100 mL), saturated aqueous NH₄Cl (100 mL), and brine (100 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (20 g, hexane/EtOAc = 6:4) to give the 7-oxanorbornene 3 (1.29 g, 68%) as a white solid: IR (film) 2921, 1684, 1512, 1384, 1247, 1029, 701 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.36-7.30 (m, 3 H), 7.22 (d, J = 7.0 Hz, 2 H), 7.09 (d, J = 8.0 Hz, 2 H), 6.81 (d, J = 8.0 Hz, 2 H), 6.35 (d, J = 6.0 Hz, 1 H), 5.96 (br s, 1 H), 5.25 (d, J = 1.5 Hz, 1 H), 5.04 (d, J = 15.5 Hz, 1 H), 5.96 (br s, 1 H), 5.25 (d, J = 1.5 Hz, 1 H), 5.04 (d, J = 15.5 Hz, 1 H), 4.45 (dd, J = 14.5, 5.5 Hz, 1 H), 4.36 (dd, J = 14.5, 5.5 Hz, 1 H), 3.94 (s, 1 H), 3.89 (d, J = 15.5 Hz, 1 H), 3.86 (d, J = 15.5 Hz, 1 H), 3.76 (s, 3 H), 2.62 (d, J = 7.5 Hz, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ 170.9, 166.8, 159.3, 137.3, 135.2, 135.0, 129.4, 128.9, 128.0, 128.0, 126.8, 114.3, 92.0, 88.8, 61.6, 55.3, 49.2, 45.4, 43.9, 18.2; HRMS (ESI, positive) calcd for $C_{24}H_{24}IN_2O_4$ [(M+H)⁺] 531.0775, found 531.0782.

The 7-Oxanorbornene Allyl Ether 4a. To a stirred solution of allyl alcohol (7.80 mL, 11.3 mmol) in DMF (22 mL) at rt was added NaH (60% in mineral oil, 452 mg, 11.3 mmol). After 30 min, a solution of the iodide **3** (1.011 g, 1.89 mmol) in DMF (44 mL) was added *via* a cannula, and the mixture was stirred at same temperature for 1.5 h. The mixture was poured into saturated aqueous NH_4CI (100 mL) and extracted with EtOAc (100 mL). The extract

was washed with water (3 × 30 mL) and brine (50 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (20 g, hexane/EtOAc = 7:3) to give the allyl ether **4a** (633 mg, 73%) as a yellow solid: IR (film) 3298, 2921, 1669, 1513, 1247, 1055, 701 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.35-7.28 (m, 3 H), 7.18 (d, *J* = 6.5 Hz, 2 H), 7.06 (d, *J* = 8.5 Hz, 2 H), 6.39 (s, 2 H), 6.03 (br t, *J* = 5.0 Hz, 1 H), 5.87 (m, 1 H), 5.30 (dd, *J* = 17.5, 1.5 Hz, 1 H), 5.19 (d, *J* = 10.0 Hz, 1 H), 4.98 (d, *J* = 4.0 Hz, 1 H), 4.80 (d, *J* = 15.0 Hz, 1 H), 4.40 (dd, *J* = 14.8, 6.0 Hz, 1 H), 4.33 (s, 1 H), 4.32 (dd, *J* = 14.8, 6.0 Hz, 1 H), 4.10 (dd, *J* = 12.8, 5.0 Hz, 1 H), 4.03 (d, *J* = 14.5 Hz, 1 H), 4.02 (dd, *J* = 12.8, 5.0 Hz, 1 H), 3.97 (s, 1 H), 3.75 (s, 3 H), 2.45 (d, *J* = 2.0 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 173.4, 166.8, 159.1, 137.5, 134.6, 133.7, 133.5, 129.3, 128.7, 127.8, 127.7, 126.9, 117.6, 91.7, 79.4, 78.5, 71.3, 63.4, 55.1, 54.3, 45.3, 43.7; HRMS (ESI, positive) calcd for C₂₇H₂₉N₂O₅ [(M+H)⁺] 461.2074, found 461.2071.

The 7-Oxanorbornene Butenyl Ether 4b. With the same procedure for the synthesis of **4a**, **4b** (407 mg, 49%) was obtained as a yellow solid starting from **3** (900 mg, 1.75 mmol), NaH (430 mg, 10.41 mmol), and 3-buten-1-ol (903 mL, 10.41 mmol).

Data for 4b. IR (film) 3074, 2912, 1667, 1513, 1247, 1031, 820, 713 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.35-7.27 (m, 3 H), 7.18 (d, *J* = 10.0 Hz, 2 H), 7.06 (d, *J* = 9.0 Hz, 2 H), 6.78 (d, *J* = 9.0 Hz, 2 H), 6.37 (dd, *J* = 8.5, 6.5 Hz, 2 H), 6.11 (br t, *J* = 5.5 Hz, 1 H), 5.74 (m, 1 H), 5.05 (dd, *J* = 17.0, 2.0 Hz, 1 H), 5.00 (d, *J* = 10.5 Hz, 1 H), 4.96 (dd, *J* = 4.5, 1.5 Hz, 1 H), 4.80 (d, *J* = 15.0 Hz, 1 H), 4.39 (dd, *J* = 14.1, 6.0 Hz, 1 H), 4.33 (dd, *J* = 14.1, 6.0 Hz, 1 H), 4.27 (dd, *J* = 4.5, 2.0 Hz, 1 H), 4.02 (d, *J* = 15.0 Hz, 1 H), 3.97 (s, 1 H), 3.74 (s, 3 H), 3.60-3.53 (m, 2 H), 2.42 (d, *J* = 2.0 Hz, 1 H), 2.29 (dd, *J* = 7.0, 6.8 Hz, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ 173.5, 166.8, 159.2, 137.5 (× 2), 134.6, 133.5, 129.4, 128.7, 127.9, 127.8, 127.0, 116.6, 114.1, 91.7, 79.5, 79.0, 69.9, 63.5, 55.2, 54.2, 45.4, 43.7, 33.9; HRMS (ESI, positive) calcd for C₂₈H₃₁N₂O₅ [(M+H)⁺] 475.2223, found 475.2215.

The 7-Oxanorbornene *N*-Allyl *N*-Ns Amide 4c. To a stirred solution of the iodide 3 (201.1 mg, 0.38 mmol) in DMF (5.0 mL) at rt were added *N*-allyl 2-nitrobenzenesulfonamide (138 mg, 0.57 mmol) and Cs₂CO₃ (1.14 mmol). After stirring at 50 °C for 10 h, the mixture was cooled to rt, poured into saturated aqueous NH₄Cl (20 mL), and extracted with EtOAc (50 mL). The extract was washed with water (3 × 10 mL) and brine (30 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (4 g, hexane/EtOAc = 5:5) to give the Ns amide **4c** (244.5 mg, 100%) as a yellow solid: IR (film) 3087, 2933, 1695, 1541, 1513, 1359, 1247, 1173, 1031, 737, 589 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.15 (dd, *J* = 7.3, 2.0 Hz, 1 H), 7.76-7.70 (m, 2 H), 7.64 (dd, *J* = 7.3, 2.0 Hz, 1 H), 7.35-7.29 (m, 3 H), 7.19 (d, *J* = 7.0 Hz, 1 H), 7.03 (d, *J* =

9.0 Hz, 2 H), 6.79 (d, J = 9.0 Hz, 2 H), 6.50 (dd, J = 6.0, 1.5 Hz, 1 H), 6.35 (d, J = 6.0 Hz, 1 H), 5.98 (br t, J = 5.5 Hz, 1 H), 5.68 (m, 1 H), 5.18 (d, J = 11.5 Hz, 1 H), 5.16 (s, 1 H), 5.07 (d, J = 11.5 Hz, 1 H), 4.83 (d, J = 15.0 Hz, 1 H), 4.46 (t, J = 3.5 Hz, 1 H), 4.43 (dd, J = 15.0, 6.5 Hz, 1 H), 4.46 (t, J = 3.5 Hz, 1 H), 4.43 (dd, J = 15.0, 6.5 Hz, 1 H), 3.94 (d, J = 15.0 Hz, 1 H), 3.92 (s, 1 H), 3.88 (br s, 2 H), 3.75 (s, 3 H), 2.93 (d, J = 3.5 Hz, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ 172.7, 166.7, 159.3, 128.5, 137.2, 135.7, 134.0, 133.9, 133.5, 132.1, 132.0, 130.8, 129.5, 128.9, 128.0, 127.8, 126.6, 124.3, 117.9, 114.2, 91.2, 81.9, 62.9, 59.8, 55.2, 50.4, 49.8, 45.4, 43.8; HRMS (ESI, positive) calcd for C₃₃H₃₃N₄O₈ [(M+H)⁺] 645.2014, found 645.2018.

The 7-Oxanorbornene *N*-Butenyl *N*-Ns Amide 4d. With the same procedure for the synthesis of 4c, 4d (1.04 g, 85%) was obtained as a yellow solid starting from 3 (1.00 g, 1.87 mmol), Cs_2CO_3 (1.22 g, 3.74 mmol), and *N*-(3-butenyl) 2-nitrobenzenesulfonamide (719 mg, 2.81 mmol).

Data for 4d. IR (film) 3088, 2933, 1695, 1542, 1512, 1355, 1246, 1173, 1032, 681, 589 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.17 (d, *J* = 8.0 Hz, 1 H), 7.77-7.65 (m, 3 H), 7.36-7.30 (m, 3 H), 7.21 (d, *J* = 7.0 Hz, 2 H), 7.03 (d, *J* = 8.5 Hz, 2 H), 6.79 (d, *J* = 8.5 Hz, 2 H), 6.47 (d, *J* = 6.0 Hz, 1 H), 6.37 (d, *J* = 6.0 Hz, 1 H), 6.03 (br s, 1 H), 5.61 (m, 1 H), 5.21 (d, *J* = 4.0 Hz, 1 H), 5.01 (d, *J* = 5.5 Hz, 1 H), 4.99 (s, 1 H), 4.86 (d, *J* = 15.0 Hz, 1 H), 4.43 (dd, *J* = 14.5, 5.5 Hz, 1 H), 4.36 (dd, *J* = 14.5, 5.5 Hz, 1 H), 3.95 (d, *J* = 15.0 Hz, 1 H), 3.93 (s, 1 H), 3.75 (s, 3 H), 3.21 (m, 2 H), 2.81 (d, *J* = 4.0 Hz, 1 H), 2.43 (m, 1 H), 2.08 (m, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ 172.7, 166.8, 159.4, 137.0, 136.0, 133.9, 133.8, 133.6, 132.2, 132.1, 130.3, 129.6, 129.0 (× 2), 128.1, 127.9, 126.5, 124.4, 121.2, 117.7, 114.3, 91.1, 82.2, 62.9, 60.0, 55.3, 50.9, 47.4, 45.5, 44.0, 33.9; HRMS (ESI, positive) calcd for C₃₄H₃₅N₄O₈S [(M+H)⁺] 659.2170, found 659.2179.

The TFA amide 4e. To a stirred solution of **4d** (1.00 g, 1.50 mmol) in acetonitrile (20 mL) at 0 °C were added thiophenol (308 μ L, 3.00 mmol) and Cs₂CO₃ (733 mg, 2.3 mmol). After stirring at rt for 2.5 h, the mixture was then poured into saturated aqueous NaHCO₃ (100 mL) and extracted with DCM (2 × 100 mL). The combined extracts were dried over Na₂SO₄ and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (20 g, methanol/chloroform = 1:9) to give the amine (628 mg, 88%) as a pale yellow oil, which was used in the next reaction without characterization.

To a stirred solution of the amine thus obtained (426.8 mg, 0.099 mmol) in DCM (10 mL) at 0 °C were added TEA (137.5 μ L, 0.992 mmol) and TFAA (138 μ L, 0.992 mmol). After stirring at rt for 30 min, the mixture was then poured into saturated aqueous NaHCO₃ (50 mL) and extracted with EtOAc (2 × 100 mL). The combined extracts were washed with brine (30 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was

purified by column chromatography on silica gel (8 g, hexane/EtOAc = 3:1) to give the TFA amide **4e** (379.6 mg, 74%) as a pale yellow oil: IR (film) 2934, 1697, 1540, 1513, 1417, 1246, 1204, 1146, 700 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.35-7.29 (m, 3 H), 7.20 (d, *J* = 7.5 Hz, 2 H), 7.05 (d, *J* = 8.5 Hz, 2 H), 6.79 (d, *J* = 8.5 Hz, 2 H), 6.34 (d, *J* = 5.5 Hz, 1 H), 6.21 (d, *J* = 6.0 Hz, 1 H), 6.11 (br t, *J* = 5.0 Hz, 1 H), 5.68 (m, 1 H), 5.68 (s, 1 H), 5.16 (d, *J* = 17.0 Hz, 1 H), 5.08 (d, *J* = 10.5 Hz, 1 H), 4.88 (d, *J* = 14.5 Hz, 1 H), 4.43-4.34 (m, 3 H), 3.97 (d, *J* = 17.0 Hz, 1 H), 3.95 (s, 1 H), 3.74 (s, 3 H), 3.60 (m, 1 H), 3.15 (m, 1 H), 2.88 (d, *J* = 3.5 Hz, 1 H), 2.65 (m, 1 H), 2.35 (m, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ 172.4, 166.9, 159.3, 157.7, 137.1, 135.3, 134.2, 132.8, 129.5, 128.8, 128.0, 127.8, 126.6, 118.3, 117.3, 114.2, 90.8, 80.7, 62.6, 58.9, 55.2, 51.7, 46.7, 45.4, 43.8, 32.6; HRMS (ESI, positive) calcd for C₃₀H₃₁N₃O₅ [(M+H)⁺] 570.2210, found 570.2198.

The Heterobicycle 8. To a stirred solution of 3 (200 mg, 0.38 mmol) in benzene (5.0 mL) at rt were added vinyl acetate (173.4 μL, 1.87 mmol) and Hoveyda-Grubbs catalyst 2nd generation (2.3 mg, 3.8 μmol) under argon atmosphere. After 14 h, the mixture was concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (4 g, hexane/EtOAc = 7:3) to give the heterobicycle 8 (205.7 mg, 87%, *E/Z* = 13:1) as a colorless oil: IR (film) 3042, 1759, 1683, 1557, 1540, 1513, 1212, 1028, 699 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.42 (d, *J* = 12.0 Hz, 1 H), 7.33-7.28 (m, 3 H), 7.20 (d, *J* = 7.5 Hz, 2 H), 7.02 (d, *J* = 8.5 Hz, 2 H), 6.78 (d, *J* = 8.5 Hz, 2 H), 5.78-5.71 (m, 2 H), 5.43 (d, *J* = 12.0 Hz, 1 H), 5.42 (d, *J* = 14.5 Hz, 1 H), 5.36 (d, *J* = 10.0 Hz, 1 H), 5.05 (d, *J* = 14.5 Hz, 1 H), 4.41 (dd, *J* = 10.5, 7.0 Hz, 1 H), 4.34 (dd, *J* = 14.5 Hz, 1 H), 3.76 (s, 3 H), 3.52 (s, 1 H), 3.92 (dd, *J* = 10.1, 7.0 Hz, 1 H), 3.78 (d, *J* = 14.5 Hz, CDCl₃) δ 169.8, 167.5, 167.3, 159.2, 139.3, 137.3, 133.0, 129.7, 128.7, 128.0, 127.7, 127.0, 121.1, 114.1, 112.6, 87.2, 84.7, 69.9, 55.2, 53.6, 45.3, 43.7, 20.6, 20.0; HRMS (ESI, positive) calcd for C₂₈H₃₀N₂O₆I [(M+H)⁺] 617.1143, found 617.1151.

The Heterotricycle 9a. To a stirred solution of 4a (310 mg, 0.64 mmol) in benzene (5.0 mL) were added vinyl acetate (312.0 μ L, 3.37 mmol) and Hoveyda-Grubbs catalyst 2nd generation (2.1 mg, 3.2 μ mol) under argon atmosphere. After 4 h, the mixture was concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (4 g, hexane/EtOAc = 6:4) to give the heterotricycle 9a (345.9 mg, 100%) as a brown liquid: IR (film) 2835, 1758, 1682, 1541, 1513, 1455, 1246, 1219, 1090, 1031, 700 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.34 (d, *J* = 12.0 Hz, 1 H), 7.33-7.27 (m, 3 H), 7.21 (d, *J* = 6.5 Hz, 2 H), 7.00 (d, *J* = 8.5 Hz, 2 H), 6.79 (d, *J* = 8.5 Hz, 2 H), 6.04 (dd, *J* = 10.0, 3.5 Hz, 1 H), 5.94 (d, *J* = 10.0, 2.5 Hz, 1 H), 5.78 (br t, *J* = 5.5 Hz, 1 H), 5.41 (d, *J* = 12.0 Hz, 1 H),

5.03 (d, J = 14.5 Hz, 1 H), 4.43 (dd, J = 14.5, 6.0 Hz, 1 H), 4.38 (d, J = 3.5 Hz, 1 H), 4.34 (dd, J = 14.5, 6.0 Hz, 1 H), 4.10 (dd, J = 17.0, 3.5 Hz, 1 H), 4.10 (br s, 1 H), 3.98 (d, J = 17.0 Hz, 1 H), 3.76 (s, 3 H), 3.71 (d, J = 14.5 Hz, 1 H), 3.66 (s, 1 H), 3.33 (s, 1 H), 2.05 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 171.6, 167.9 (× 2), 159.5, 139.2, 137.6, 131.4, 130.0, 129.9, 129.0, 128.3, 128.0, 127.4, 122.0, 114.5, 113.2, 85.6, 78.5, 74.0, 71.6, 64.2, 59.0, 55.5, 45.4, 44.1, 21.0; HRMS (ESI, positive) calcd for C₂₉H₃₁N₂O₇ [(M+H)⁺] 519.2126, found 519.2119.

The Heterotricycle 9b. To a stirred solution of **4b** (298.1 mg, 0.63 mmol) in benzene (7.0 mL) at rt was added vinyl acetate (291.0 μ L, 3.15 mmol) and Hoveyda-Grubbs catalyst 2nd generation (3.9 mg, 6.3 μ mol) under argon atmosphere. After 46 h, the mixture was concentrated under reduced pressure. The Ru catalyst was removed by passing through a short pad of silica gel (6 g, hexane/EtOAc = 4:6). The filtrate was concentrated under reduced pressure to give a residue which was mainly composed of the triene ROM/CM product **9b'** (84% yield on isolation, *E*/*Z* = >20:1).

The residue thus obtained was, without purification, dissolved in benzene (7.0 mL). To the stirred mixture at 69 °C was added Hoveyda-Grubbs catalyst 2nd generation (3.9 mg, 6.3 μ mol). After 21 h, the mixture was cooled to rt and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (6 g, hexane/EtOAc = 6:4) to give the heterotricycle **9b** (281.9 mg, 84%, *E*/*Z* = >20:1) as a brown liquid: IR (film) 2932, 1757, 1674, 1513, 1455, 1246, 1217, 1111, 1031, 699 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.42 (d, *J* = 12.0 Hz, 1 H), 7.32-7.26 (m, 3 H), 7.16 (d, *J* = 7.0 Hz, 1 H), 7.00 (d, *J* = 8.5 Hz, 2 H), 6.79 (d, *J* = 8.5 Hz, 2 H), 5.79 (m, 1H), 5.74 (br s, 1 H), 5.60 (dd, *J* = 11.8, 4.0 Hz, 1 H), 5.40 (d, *J* = 12.0 Hz, 1 H), 5.01 (d, *J* = 14.5 Hz, 1 H), 4.48-4.45 (m, 2 H), 4.42 (dd, *J* = 14.5, 6.0 Hz, 1 H), 3.65 (d, *J* = 1.5 Hz, 1 H), 3.57 (m, 1 H), 3.30 (s, 1 H), 2.34-2.28 (m, 2 H), 2.06 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 171.8, 167.6 (× 2) 159.1, 138.8, 137.4, 129.8, 129.5, 128.5, 127.8, 127.5, 127.1, 125.7, 114.0, 112.3, 83.9, 83.0, 82.5, 71.0, 69.0, 59.9, 55.1, 44.9, 43.6, 30.6, 20.6; HRMS (ESI, positive) calcd for C₃₀H₃₃N₂O₇ [(M+H)⁺] 533.2282, found 533.2281. **Data for the Intermediary Triene 9b'.** IR (film) 2931, 1758, 1675, 1551, 1513, 1453, 1370,

Data for the intermediary mene 95. IR (iiiii) 2931, 1738, 1675, 1551, 1513, 1453, 1475, 1247, 1216, 1108, 1035, 930, 700, 649, 596 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.40 (d, *J* = 12.0 Hz, 1 H), 7.32-7.26 (m, 3 H), 7.21-7.20 (m, 2 H), 7.00 (d, *J* = 8.5 Hz, 2 H), 6.78 (d, *J* = 8.5 Hz, 2 H), 5.94 (m, 1 H), 5.85 (br s, 1 H), 5.72 (m, 1 H), 5.41 (d, *J* = 12.0 Hz, 1 H), 5.26 (d, *J* = 17.0 Hz, 1 H), 5.23 (d, *J* = 10.0 Hz, 1 H), 5.02-4.94 (m, 3 H), 4.43 (dd, *J* = 14.5, 6.0 Hz, 1 H), 4.31 (dd, *J* = 14.5, 6.0 Hz, 1 H), 4.28 (dd, *J* = 8.0, 3.5 Hz, 1 H), 4.21 (d, *J* = 3.5 Hz, 1 H), 3.75 (s, 3 H), 3.71 (d, *J* = 15.0 Hz, 1 H), 3.69 (s, 1 H), 3.54-3.43 (s, 1 H), 2.24 (m, 2 H), 2.08 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 171.9, 167.7, 167.6, 159.3, 138.7, 137.3, 135.0, 133.1, 129.7, 128.7, 128.0, 127.8, 127.1, 119.2, 116.4, 114.2, 112.7, 85.2, 84.8, 83.6, 71.5, 127.1, 119.2, 116.4, 114.2, 112.7, 85.2, 84.8, 83.6, 71.5, 127.1, 119.2, 116.4, 114.2, 112.7, 85.2, 84.8, 83.6, 71.5, 127.1, 119.2, 116.4, 114.2, 112.7, 85.2, 84.8, 83.6, 71.5, 127.1, 119.2, 116.4, 114.2, 112.7, 85.2, 84.8, 83.6, 71.5, 127.1, 119.2, 116.4, 114.2, 112.7, 85.2, 84.8, 83.6, 71.5, 128.0, 127.8, 127.1, 119.2, 116.4, 114.2, 112.7, 85.2, 84.8, 83.6, 71.5, 128.0, 127.8, 127.1, 119.2, 116.4, 114.2, 112.7, 85.2, 84.8, 83.6, 71.5, 128.0, 127.8, 127.1, 119.2, 116.4, 114.2, 112.7, 85.2, 84.8, 83.6, 71.5, 128.0, 127.8, 127.1, 119.2, 116.4, 114.2, 112.7, 85.2, 84.8, 83.6, 71.5, 128.0, 127.8, 127.1, 119.2, 116.4, 114.2, 112.7, 85.2, 84.8, 83.6, 71.5, 128.0, 127.8, 127.1, 119.2, 116.4, 114.2, 112.7, 85.2, 84.8, 83.6, 71.5, 128.0, 127.8, 127.1, 119.2, 116.4, 114.2, 112.7, 85.2, 84.8, 83.6, 71.5, 128.0, 127.8, 127.1, 119.2, 116.4, 114.2, 112.7, 85.2, 84.8, 83.6, 71.5, 128.0, 127.8, 127.1, 119.2, 116.4, 114.2, 112.7, 85.2, 84.8, 83.6, 71.5, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128.0, 128

69.2, 57.3, 55.3, 45.1, 43.8, 34.0, 20.7; HRMS (ESI, positive) calcd for $C_{32}H_{37}N_2O_7$ [(M+H)⁺] 561.2595, found 561.2600.

The Heterotricycle 9c. With the same procedure for the synthesis of **9a**, **9c** (1.07 g, 97%) was obtained as a brown solid starting from **4c** (1.00 g, 1.42 mmol), Hoveyda-Grubbs catalyst 2^{nd} generation (19.6 mg, 0.032 mmol), and vinyl acetate (727 µL, 7.85 mmol).

Data for 9c. IR (film) 2940, 1758, 1696, 1542, 1513, 1370, 1245, 1172, 1031, 682, 583 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.42 (d, *J* = 8.0 Hz, 1 H), 7.81-7.68 (m, 3 H), 7.34-7.30 (m, 3 H), 7.20 (d, *J* = 7.5 Hz, 2 H), 7.05 (d, *J* = 8.5 Hz, 2 H), 6.83 (d, *J* = 8.5 Hz, 2 H), 5.91 (dd, *J* = 10.5, 5.0 Hz, 1 H), 5.80-5.77 (m, 2 H), 5.46 (d, *J* = 12.5 Hz, 1 H), 5.04 (d, *J* = 14.5 Hz, 1 H), 4.95 (t, *J* = 6.5 Hz, 1 H), 4.75 (d, *J* = 6.5 Hz, 1 H), 4.38 (dd, *J* = 14.5, 5.5 Hz, 1 H), 4.32 (dd, *J* = 14.5, 5.5 Hz, 1 H), 4.19 (dd, *J* = 18.5, 5.0 Hz, 1 H), 3.81 (s, 3 H), 3.77 (d, *J* = 14.5 Hz, 1 H), 3.16 (d, *J* = 4.0 Hz, 1 H), 2.11 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 172.2, 167.6, 166.9, 159.3, 128.0, 138.1, 137.4, 133.9, 132.3, 132.1, 129.7, 128.6 (× 2), 128.2, 128.0, 127.6, 127.4, 126.8, 126.1, 124.2, 114.2, 112.6, 84.5, 72.8, 70.6, 57.9, 55.2, 54.6, 45.0, 43.6, 40.3, 20.6; HRMS (ESI, positive) calcd for C₃₅H₃₅N₄O₁₀S [(M+H)⁺] 703.2068, found 703.2076.

The Heterotricycle 9d. With the same procedure for the synthesis of **9b**, **9d** (220.1 mg, 90%) was obtained as a brown solid starting from **4d** (225.9 mg, 0.343 mmol), Hoveyda-Grubbs catalyst 2^{nd} generation (21.5 mg, 0.034 mmol), and vinyl acetate (158.9 μ L, 1.720 mmol).

Data for 9d. IR (film) 3033, 2975, 1757, 1697, 1542, 1246, 1165, 681 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.43 (d, *J* = 8.0 Hz, 1 H), 7.81 (t, *J* = 7.5 Hz, 1 H), 7.72 (t, *J* = 7.5 Hz, 2 H), 7.60 (d, *J* = 8.0 Hz, 1 H), 7.31 (d, *J* = 13.0 Hz, 1 H), 7.31-7.27 (m, 3 H), 7.18 (d, *J* = 5.5 Hz, 2 H), 6.97 (d, *J* = 8.5 Hz, 2 H), 6.78 (d, *J* = 8.5 Hz, 2 H), 5.96 (dt, *J* = 11.5, 3.5 Hz, 1 H), 5.69 (br t, *J* = 5.5 Hz, 1 H), 5.60 (m, 1 H), 5.51 (d, *J* = 13.0 Hz, 1 H), 5.09 (d, *J* = 14.5 Hz, 1 H), 4.81 (d, *J* = 5.5 Hz, 1 H), 4.35 (d, *J* = 5.5 Hz, 2 H), 3.83 (s, 3 H), 3.77 (d, *J* = 9.5 Hz, 1 H), 3.77 (s, 3 H), 3.59 (d, *J* = 15.0 Hz, 1 H), 3.54 (s, 1 H), 3.45 (m, 1 H), 2.69 (s, 1 H), 2.67 (m, 1 H), 2.37 (d, *J* = 17.5 Hz, 1 H), 2.10 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 170.8, 167.6, 166.8, 159.4, 147.6, 138.0, 137.1, 133.8, 132.9, 132.7, 132.1, 129.9, 128.7, 128.2, 128.0, 127.8, 126.7, 123.7, 120.6, 114.3, 112.1, 84.2, 82.4, 70.8, 65.6, 59.8, 55.2, 45.0, 43.7, 42.8, 33.3, 20.6; HRMS (ESI, positive) calcd for C₃₆H₃₇N₄O₁₀S [(M+H)⁺] 717.2225, found 714.2218.

The Heterotricycle 9e. With the same procedure for the synthesis of **9b**, **9e** (363.0 mg, 94%) was obtained starting from **4e** (354.0 mg, 0.620 mmol), Hoveyda-Grubbs catalyst 2^{nd} generation (38.8 mg, 0.062 mmol), and vinyl acetate (287.9 µL, 3.11 mmol).

Data for 9e. IR (film) 2934, 1760, 1696, 1514, 1210, 1035, 700 cm⁻¹; ¹H NMR (500 MHz,

CDCl₃, ca 7:3 mixture of rotamers) δ 7.41 (d, *J* = 12.5 Hz, 0.3 H), 7.35 (d, *J* = 12.5 Hz, 0.7 H), 7.31-7.18 (m, 5 H), 7.02 (d, *J* = 8.0 Hz, 0.6 H), 7.00 (d, *J* = 8.0 Hz, 1.4 H), 6.80 (d, *J* = 12.5 Hz, 0.7 H), 6.78 (d, *J* = 8.0 Hz, 0.6 H), 6.11 (br s, 1 H), 5.94 (dt, *J* = 11.5, 4.0 Hz, 0.7 H), 5.88 (dt, *J* = 11.5, 4.0 Hz, 0.3 H), 5.65 (d, *J* = 12.5, 0.7 H), 5.64 (d, *J* = 12.5 Hz, 0.3 H), 5.57 (m, 1 H), 5.19 –5.02 (m, 2 H), 4.76 (d, *J* = 5.0 Hz, 1.4 H), 4.46 (dd, *J* = 14.5, 6.0 Hz, 1 H), 4.39-4.35 (m, 2 H), 4.15 (t, *J* = 6.5 Hz, 0.7 H), 4.04-3.85 (m, 3.3 H), 3.85-3.77 (m, 6 H), 3.74-3.65 (m, 3 H), 3.55 (m, 1 H), 3.23 (d, *J* = 11.5, 1.4 H), 2.72 (m, 0.3 H), 2.50-2.22 (m, 1.7 H), 2.10 (s, 0.9 H), 2.08 (s, 2.1 H); ¹³C NMR (125 MHz, CDCl₃) δ 171.2, 170.8, 167.7, 167.6, 166.9, 166.8, 166.5, 166.3, 159.4, 159.3, 138.3, 137.9, 137.8, 137.7, 137.5, 137.3, 130.0 (× 2), 128.5, 128.4, 128.2, 128.1, 127.5, 127.2, 126.9, 126.7, 121.2, 120.4, 114.2 (× 2), 113.0, 112.6; HRMS (ESI, positive) calcd for C₃₆H₃₇N₄O₁₀S [(M+H)⁺] 717.2225, found 717.2217.

The *N*-Boc Imide S3a. To a stirred solution of the *N*-Bn amide 9a (290.0 mg, 0.56 mmol) in DCM (5.0 mL) at 0 °C were added Boc₂O (396 μL, 1.69 mmol), TEA (310 μL, 2.24 mmol) and DMAP (34 mg, 0.28 mmol). After 2.5 h, the mixture was diluted with EtOAc (20 mL), washed with saturated aqueous NH₄Cl (20 mL) and brine (20 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (6 g, hexane/EtOAc = 7:3) to give the *N*-Boc imide S3a (329 mg, 95%) as a white solid: IR (film) 2892, 2836, 1697, 1513, 1250, 1147, 1032, 848, 700 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.36 (d, *J* = 12.0 Hz, 1 H), 7.29-7.21 (m, 5 H), 6.96 (d, *J* = 8.0 Hz, 2 H), 6.78 (d, *J* = 8.0 Hz, 1 H), 6.02-5.96 (m, 2 H), 5.41 (s, 1 H), 5.22 (d, *J* = 12.0 Hz, 2 H), 4.84 (d, *J* = 15.0 Hz, 1 H), 4.80 (d, *J* = 13.5, 3.0 Hz, 1 H), 3.97 (d, *J* = 13.5 Hz, 1 H), 3.75 (s, 3 H), 3.74 (d, *J* = 14.5 Hz, 1 H), 3.29 (s, 1 H), 2.02 (s, 3 H), 1.30 (s, 9 H); ¹³C NMR (125 MHz, CDCl₃) δ 172.1, 171.6, 167.3, 159.2, 151.9, 139.1, 137.2, 130.8, 130.0, 128.3, 127.4, 127.1, 122.1, 114.1, 112.5, 85.9, 84.8, 78.1, 73.9, 69.7, 63.9, 58.7, 55.2, 47.8, 45.3, 27.6, 20.6; HRMS (ESI, positive) calcd for C₃₄H₃₉N₂O₉ [(M+H)⁺] 619.2650, found 619.2660.

The Ester Aldehyde 13a. To a stirred solution of the imide S3a (290.0 mg, 0.56 mmol) in methanol (15 mL) at -20 °C was added K₂CO₃ (36.6 mg, 0.27 mmol). After 5 h, the mixture was poured into saturated aqueous NH₄Cl (30 mL), and the mixture was extracted with EtOAc (50 mL). The extract was washed with brine (50 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (6 g, hexane/EtOAc = 7:3) to give the ester aldehyde **13a** (178 mg, 84%) as a white solid: IR (film) 2954, 1745, 1696, 1513, 1441, 1248, 1030, 684 cm⁻¹; ¹H NMR (500 MHz, C₆D₆) δ 9.67 (s, 1 H), 703 (d, *J* = 8.5 Hz, 2 H), 6.67 (d, *J* = 8.5 Hz, 2 H), 5.53 (dd, *J* = 5.5, 2.0 Hz, 1 H), 5.32 (dd, *J* = 10.0, 3.5 Hz, 1 H), 4.97 (d, *J* = 14.5 Hz, 1 H), 4.39 (d, *J* = 2.0

Hz, 1 H), 4.34 (s, 1 H), 3.94 (s, 1 H), 3.92 (d, J = 14.5 Hz, 1 H), 3.60 (dd, J = 17.0, 4.0 Hz, 1 H), 3.42 (d, J = 17.0 Hz, 1 H), 3.30 (s, 1 H), 3.22 (s, 3 H), 3.09 (s, 3 H), 2.87 (d, J = 16.5 Hz, 1 H), 2.45 (d, J = 16.5 Hz, 1 H); ¹³C NMR (125 MHz, C_6D_6) δ 198.4, 170.5, 169.8, 159.8, 130.9, 130.1, 128.3, 122.0, 114.4, 85.1, 78.8, 74.1, 69.4, 63.9, 58.2, 54.7, 51.8, 49.1, 45.5; HRMS (ESI, positive) calcd for $C_{21}H_{23}NO_7Na$ [(M+Na)⁺] 424.1367, found 424.1366.

The *N***-Boc Imide S3b.** With the same procedure for the synthesis of S3a, S3b (311.7 mg, 95%) was obtained as a white solid starting from **9b** (275.0 g, 0.52 mmol), Boc₂O (363.6 μ L, 1.55 mmol), DMAP (19.1 mg, 0.15 mmol), and TEA (214.9 μ L, 1.55 mmol).

Data for S3b. IR (film) 2834, 1692, 1513, 1249, 1147, 847, 630 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.42 (d, *J* = 12.5 Hz, 1 H), 7.28-7.21 (m, 5 H), 6.96 (d, *J* = 8.5 Hz, 2 H), 6.78 (d, *J* = 8.5 Hz, 2 H), 5.75 (dt, *J* = 12.0, 5.5 Hz, 1 H), 5.66 (dd, *J* = 12.0, 4.0 Hz, 1 H), 5.37 (s, 1 H), 5.22 (d, *J* = 12.5 Hz, 1 H), 4.83 (d, *J* = 12.5 Hz, 1 H), 4.77 (d, *J* = 14.5 Hz, 1 H), 4.69 (d, *J* = 15.0 Hz, 1 H), 4.58 (br s, 1 H), 4.47 (d, *J* = 3.0 Hz, 1 H), 3.91 (m, 1 H), 3.77-3.74 (m, 4 H), 3.58 (m, 1 H), 3.27 (s, 1 H), 2.30 (m, 2 H), 2.04 (s, 3 H), 1.30 (s, 9 H); ¹³C NMR (125 MHz, CDCl₃) δ 172.2, 172.0, 167.3, 159.2, 151.8, 139.2, 137.1, 130.0, 129.1, 128.2, 128.1, 127.4, 127.0, 126.4, 114.1, 111.9, 84.7, 84.5, 83.4, 82.3, 69.5, 69.0, 60.1, 55.2, 47.8, 45.3, 30.4, 27.6, 20.6; HRMS (ESI, positive) calcd for C₃₅H₄₁N₂O₉ [(M+H)⁺] 633.2807, found 633.2803.

The Ester Aldehyde 13b. With the same procedure for the synthesis of **13a**, **13b** (147.1 mg, 83%) was obtained as a white solid starting from **S3b** (270.0 mg, 0.43 mmol) and K_2CO_3 (29.5 mg, 0.21 mmol).

Data for 13b. IR (film) 2953, 1743, 1698, 1514, 1436, 1249, 1177, 1027, 683 cm⁻¹; ¹H NMR (500 MHz, C_6D_6) δ 9.86 (s, 1 H), 7.16 (d, *J* = 8.5 Hz, 2 H), 6.80 (d, *J* = 8.5 Hz, 2 H), 5.60 (dd, *J* = 11.8, 3.5 Hz, 1 H), 5.45 (m, 1 H), 5.06 (d, *J* = 15.0 Hz, 1 H), 4.67 (s, 1 H), 4.66 (d, *J* = 14.5 Hz, 1 H), 4.48 (s, 1 H), 4.11 (d, *J* = 14.5 Hz, 1 H), 3.65 (m, 1 H), 3.44 (s, 1 H), 3.36 (s, 3 H), 3.22 (s, 3 H), 3.21 (m, 1 H), 3.08 (d, *J* = 17.3 Hz, 1 H), 2.62 (d, *J* = 17.3 Hz, 1 H), 1.85 (m, 2 H); ¹³C NMR (125 MHz, C_6D_6) δ 198.6, 171.0, 169.8, 159.8, 130.1, 128.9, 128.5, 126.7, 114.4, 83.8, 83.7, 82.5, 69.2, 68.8, 59.5, 54.7, 51.8, 48.7, 45.6, 30.0; HRMS (ESI, positive) calcd for $C_{22}H_{25}NO_7Na$ [(M+H)⁺] 438.1523, found 438.1518.

The N-Boc Imide S3c. With the same procedure for the synthesis of **S3a**, **S3c** (1.00 g, 90%) was obtained as a pale yellow solid starting from **9c** (1.00 g, 1.42 mmol), Boc_2O (1.66 mL, 7.10 mmol), DMAP (86.7 mg, 0.71 mmol), and TEA (984 μ L, 7.10 mmol).

Data for S3c. IR (film) 2930, 1696, 1544, 1370, 1147 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.52 (d, *J* = 8.0 Hz, 1 H), 7.78 (t, *J* = 8.0 Hz, 1 H), 7.71 (t, *J* = 8.0 Hz, 1 H), 7.65 (d, *J* = 8.0 Hz, 1 H), 7.25-7.13 (m, 6 H), 6.94 (d, *J* = 9.0 Hz, 2 H), 6.77 (d, *J* = 9.0 Hz, 2 H), 5.86 (dd, *J* = 10.5 Hz, 1 H), 5.78 (d, *J* = 10.5 Hz, 1 H), 5.30 (d, *J* = 12.5 Hz, 1 H), 5.25 (s, 1 H), 5.02 (dd, *J* = 7.5, 2.5 Hz, 1 H), 4.82 (d, *J* = 14.5 Hz, 1 H), 4.76 (d, *J* = 7.5 Hz, 1 H), 4.70 (d, *J* = 14.0 Hz, 1 H), 4.48 (d, *J* = 14.5 Hz, 1 H), 4.26 (dd, *J* = 18.0, 5.5 Hz, 1 H), 3.75 (s, 3 H), 3.75 (d, *J* = 14.0 Hz, 1 H), 3.46 (d, *J* = 18,0 Hz, 1 H), 3.12 (s, 1 H), 2.02 (s, 3 H), 1.28 (s, 9 H); ¹³C NMR (125 MHz, CDCl₃) δ 172.9, 171.8, 167.2, 159.2, 151.8, 148.0, 138.2, 137.1, 133.8, 132.7, 132.5, 132.1, 130.1, 128.2, 128.1, 127.9, 127.4, 126.4, 126.1, 123.9, 114.1, 113.0, 84.7, 84.4, 72.3, 68.9, 58.4, 55.8, 55.2, 47.6, 40.2, 27.6, 20.5; HRMS (ESI, positive) calcd for C₄₀H₄₃N₄O₁₂S [(M+H)⁺] 803.2593, found 803.2591.

The Ester Aldehyde 13c. With the same procedure for the synthesis of **13a**, **13c** (604.4 mg, 81%) was obtained as a pale yellow solid starting from **S3c** (1.03 g, 1.28 mmol) and K_2CO_3 (88.3 mg, 0.64 mmol).

Data for 13c. IR (film) 3002, 1748, 1698, 1541, 1508, 1362, 1248, 1172, 1030, 582 cm⁻¹; ¹H NMR (500 MHz, C_6D_6) δ 9.44 (s, 1 H), 8.44 (d, J = 8.0 Hz, 1 H), 7.02-6.97 (m, 3 H), 6.72-6.99 (m, 3 H), 6.56 (t, J = 7.5 Hz, 1 H), 5.18 (ddd, J = 10.8, 2.0, 2.0 Hz, 1 H), 5.10 (ddd, J = 10.8, 3.5, 3.5 Hz, 1 H), 4.93 (d, J = 14.5 Hz, 1 H), 4.76 (dd, J = 6.8, 3.5 Hz, 1 H), 4.28 (s, 1 H), 4.21 (br s, 1 H), 3.92 (d, J = 14.5 Hz, 1 H), 3.92 (d, J = 18.0 Hz, 1 H), 3.51 (d, J = 4.0 Hz, 1 H), 3.48 (d, J = 14.5 Hz, 1 H), 3.23 (s, 3 H), 3.11 (s, 3 H), 2.71 (dd, J = 17.5, 1.0 Hz, 1 H), 2.31 (dd, J = 17.5, 1.0 Hz, 1 H); ¹³C NMR (125 MHz, C_6D_6) δ 197.7, 171.8, 169.5, 159.9, 148.8, 133.8, 132.5, 131.6, 131.2, 130.1, 127.4, 127.3, 125.5, 123.8, 114.6, 84.6, 73.7, 68.9, 59.5, 54.7, 54.4, 51.8, 48.7, 45.5, 41.5; HRMS (ESI, positive) calcd for $C_{27}H_{29}N_3O_{10}S$ [(M+H)⁺] 586.1489, found 586.1490.

The *N***-Boc Imide S3e.** With the same procedure for the synthesis of S3a, S3e (351.3 mg, 83%) was obtained as a pale yellow solid starting from **9e** (363.6 mg, 0.58 mmol), Boc_2O (407.9 mg, 1.74 mmol), DMAP (35.4 mg, 0.29 mmol), and TEA (241.1 μ L, 1.74 mmol).

Data for S3e. IR (film) 2894, 1702, 1513, 1210, 1146, 848 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, ca 1:1 mixture of rotamers) δ 7.32 (d, *J* = 13.0 Hz, 0.5 H), 7.29-7.19 (m, 5.5 H), 6.97-6.94 (m, 2 H), 6.79-6.78 (d, *J* = 8.0 Hz, 2 H), 5.94-5.86 (m, 1 H), 5.70-5.62 (m, 1 H), 5.47 (d, *J* = 13.0 Hz, 0.5 H), 5.45 (d, *J* = 13.0 Hz, 0.5 H), 5.36 (s, 0.5 H), 5.35 (s, 0.5 H), 5.35 (s, 0.5 H), 5.18 (d, 0.5 H), 4.90-4.83 (m, 3.5 H), 4.58 (d, *J* = 14.5 Hz, 0.5 H), 4.54 (d, *J* = 14.5 Hz, 0.5 H), 4.40-4.28 (m, 1 H), 3.95-3.87 (m, 1 H), 3.76 (s, 3 H), 3.73-3.62 (m, 2 H), 3.22 (d, *J* = 3.5 Hz, 1 H), 2.67 (m, 0.5 H), 2.47-2.22 (m, 1.5 H), 2.05 (s, 3 H), 1.34-1.28 (m, 9 H); ¹³C NMR (125 MHz, CDCl₃, selected) δ 171.6, 171.1, 167.2, 159.3, 159.3, 156.8, 151.7, 139.1, 137.3, 137.0, 130.3, 128.0 (× 2), 127.4, 126.4, 121.7, 120.9, 117.4, 114.1, 113.0, 84.7, 83.4, 76.1, 70.1, 64.2, 60.9, 55.1, 47.7, 45.5, 41.6, 32.6, 27.5, 20.5; HRMS (ESI, positive) calcd for C₃₇H₄₁N₃O₉F₃ [(M+H)⁺] 728.2789, found 728.2787.

The Ester Aldehyde 13e. With the same procedure for the synthesis of 13a, 13e (219.4 mg,

91%) was obtained as a pale yellow solid starting from **S3e** (345.0 mg, 0.48 mmol) and K_2CO_3 (32.8 mg, 0.24 mmol).

Data for 13e. IR (film) 2933, 1748, 1698, 1541, 1508, 1145, 669 cm⁻¹; ¹H NMR (500 MHz, C₆D₆, ca 1:1 mixture of rotamers) δ 9.58 (br s, 0.5 H), 9.55 (br s, 0.5 H), 6.99 (d, *J* = 8.5 Hz, 1 H), 6.94 (d, *J* = 8.5 Hz, 1 H), 6.66 (d, *J* = 8.5 Hz, 1 H), 6.62 (d, *J* = 8.5 Hz, 1 H), 5.31-5.25 (m, 1 H), 5.16-5.02 (m, 3 H), 4.89 (d, *J* = 5.5 Hz, 0.5 H), 4.21 (s, 0.5 H), 4.08 (s, 0.5 H), 4.06 (dd, *J* = 7.3, 5.0 Hz, 0.5 H), 3.93 (dd, *J* = 7.3, 5.0 Hz, 0.5 H), 3.82-3.74 (m, 1.5 H), 3.41 (t, *J* = 13.5 Hz, 0.5 H), 3.31-3.22 (m, 6 H), 3.15 (s, 1.5 H), 3.10 (t, *J* = 12.0 Hz, 0.5 H), 2.90 (s. 0.5 H), 2.64 (br s, 0.5 H), 2.45-2.38 (m, 1.5 H), 2.35-2.32 (m, 1 H), 1.78 (br t, *J* = 18.0 Hz, 0.5 H), 1.66 (br d, *J* = 20.0 Hz, 0.5 H), 1.50 (br d, *J* = 20.0 Hz, 0.5 H), 0.91 (m, 0.5 H); ¹³C NMR (125 MHz, C₆D₆) δ 197.6, 197.4, 170.4, 170.3, 169.8 (× 2), 172.1, 167.5, 161.1, 160.1, 128.8 (× 2), 137.8, 137.1, 130.6, 130.5, 127.7, 127.5, 122.0, 120.7, 114.8 (× 2), 82.9, 82.8, 77.0, 76.8, 69.5, 68.5, 65.9, 65.8, 65.0, 60.8, 60.0, 54.9, 52.1, 49.5, 45.7, 45.6, 43.4, 42.1, 32.8, 32.1, 29.9, 27.3; HRMS (ESI, positive) calcd for C₂₄H₂₅F₃N₂O₇Na [(M+Na)⁺] 533.1506, found 533.1493.

The Diester S4a. To a stirred solution of the aldehyde 13a (222.7 mg, 0.55 mmol) in tert-butanol (15.0 mL) and water (5.0 mL) at rt were added 2-methyl-2-butene (290.8 μL, 2.75 mmol), NaH₂PO₄·2H₂O (94.2 mg, 0.60 mmol), and NaClO₂ (148.3 mg, 1.65 mmol). After 5 h, the mixture was diluted with DCM (50 mL), and the mixture was washed with hydrochloric acid (1 M, 20 mL) and brine (20 mL). The organic layer was dried over Na₂SO₄ and concentrated under reduced pressure. The residue was dissolved in methanol (15.0 mL) and cooled to 0 °C. TMSCHN₂ (2 M in Et₂O, 0.84 mL, 1.68 mmol) was added, and the mixture was allowed to warm to rt. After stirring for 30 min, the mixture was concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (6 g, hexane/EtOAc = 4:6) to give the diester S4a (235.4 mg, 94 %) as a white solid: IR (film) 2953, 1744, 1698, 1513, 1437, 1248, 1178, 1049, 822, 684 cm⁻¹; ¹H NMR (500 MHz, $CDCl_3$) δ 7.10 (d, J = 8.5 Hz, 2 H), 6.82 (d, J = 8.5 Hz, 2 H), 6.02 (dd, J = 10.5, 3.5 Hz, 1 H), 5.93 (m, 1 H), 4.81 (d, J = 14.5 Hz, 1 H), 4.39 (d, J = 3.0 Hz, 1 H), 4.34 (s, 1 H), 4.15 (dd, J = 17.3, 3.5 Hz, 1 H), 4.03 (d, J = 17.3 Hz, 1 H), 4.01 (s, 1 H), 4.00 (d, J = 14.5 Hz, 1 H), 3.77 (s, 3 H), 3.61 (s, 3 H), 3.58 (s, 3 H), 3.37 (s, 1 H), 3.11 (d, J = 16.5 Hz, 1 H), 2.75 (d, J = 16.5 Hz, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ 170.8, 170.3, 170.1, 159.2, 130.6, 129.7, 127.1, 122.5, 114.1, 85.1, 78.3, 73.6, 68.2, 64.1, 57.7, 55.2, 52.4, 51.7, 45.4, 40.1; HRMS (ESI, positive) calcd for C₂₂H₂₅NO₈Na [(M+Na)⁺] 454.1472, found 454.1477.

The Lactam 14a. To a stirred solution of the *N*-PMB amide **S4a** (187.3 mg, 0.43 mmol) in CH₃CN (10.0 mL) and water (2.4 mL) at -10 °C was added a solution of CAN (1.19 mg, 2.17

mmol) in water (6.6 mL) portionwise. After 5 h, the mixture was poured into saturated aqueous Na₂S₂O₃ (20 mL) and extracted with EtOAc (3 × 50 mL). The combined extracts were washed with saturated aqueous NaHCO₃ (20 mL) and brine (20 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (5 g, hexane/EtOAc = 4:6) to give the lactam **14a** (95.2 mg, 71%) as a white solid: IR (film) 2953, 1747, 1698, 1508, 1436, 1250, 1211, 1087, 848, 688 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 6.16 (s, 1 H), 6.05 (dd, *J* = 10.0, 4.0 Hz, 1 H), 5.98 (ddd, *J* = 10.0, 4.0, 2.0 Hz, 1 H), 4.44 (s, 1 H), 4.29 (d, *J* = 2.0 Hz, 1 H), 4.15 (s, 1 H), 4.13 (dd, *J* = 16.5, 4.0 Hz, 1 H), 4.00 (d, *J* = 16.5 Hz, 1 H), 3.67 (s, 3 H), 3.60 (s, 3 H), 3.28 (s, 1 H), 3.14 (d, *J* = 17.5 Hz, 1 H), 2.85 (d, *J* = 17.5 Hz, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ 174.1, 170.4, 130.9, 122.0, 87.3, 78.1, 73.2, 64.7, 64.1, 56.7, 52.5, 51.6, 40.3; HRMS (ESI, positive) calcd for C₁₄H₁₇NO₇Na [(M+Na)⁺] 334.0897, found 334.0899.

The Diester S4b. With the same procedure for the synthesis of **S4a**, **S4b** (139.3 mg, 73%) was obtained as a colorless oil starting from **13b** (147.1 mg, 0.35 mmol), 2-methyl-2-butene (226.2 μ L, 2.14 mmol), NaH₂PO₄·2H₂O (73.3 mg, 0.47 mmol), NaClO₂ (115.3 mg, 1.28 mmol), and TMSCHN₂ (2 M in Et₂O, 0.35 mL, 0.70 mmol).

Data for S4b. IR (film) 2953, 1744, 1698, 1513, 1437, 1248, 1047, 820 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.08 (d, *J* = 8.5 Hz, 2 H), 6.81 (d, *J* = 8.5 Hz, 2 H), 5.73 (dt, *J* = 11.5, 5.5 Hz, 1 H), 5.58 (dd, *J* = 11.5, 3.5 Hz, 1 H), 4.77 (d, *J* = 14.5 Hz, 1 H), 4.49-4.47 (m, 2 H), 4.30 (s, 1 H), 4.02 (d, *J* = 14.5 Hz, 1 H), 3.93 (m, 1 H), 3.77 (s, 3 H), 3.66 (m, 1 H), 3.62 (s, 3 H), 3.56 (s, 3 H), 3.34 (s, 1 H), 3.11 (d, *J* = 17.0 Hz, 1 H), 2.72 (d, *J* = 17.0 Hz, 1 H), 2.38-2.26 (m, 2 H); ¹³C NMR (125 MHz, CDCl₃) δ 171.4, 170.3, 170.0, 159.2, 129.7, 128.3, 127.2, 127.1, 114.1, 83.7, 83.3, 81.8, 68.7, 68.0, 58.9, 55.2, 52.3, 51.6, 45.3, 39.4, 29.9; HRMS (ESI, positive) calcd for C₂₃H₂₇NO₈Na [(M+Na)⁺] 468.1629, found 468.1634.

The Lactam 14b. With the same procedure for the synthesis of **14a**, **14b** (67.4 mg, 71%) was obtained as a white solid starting from **S4b** (130 mg, 0.29 mmol) and CAN (801 mg, 1.46 mmol).

Data for 14b. IR (film) 2953, 1743, 1715, 1436, 1362, 1211, 1046, 669 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 5.92 (br s, 1 H), 5.81 (ddd, *J* = 11.5, 5.0, 5.0 Hz, 1 H), 5.64 (dd, *J* = 11.5, 2.5 Hz, 1 H), 4.54 (br s, 1 H), 4.40 (s, 1 H), 4.37 (br s, 1 H), 3.96 (ddd, *J* = 11.5, 5.5, 4.5 Hz, 1 H), 3.69 (s, 3 H), 3.64 (s, 3 H), 3.62 (ddd, *J* = 11.5, 5.5, 4.5 Hz, 1 H), 3.29 (s, 1 H), 3.22 (d, *J* = 17.5 Hz, 1 H), 2.92 (d, *J* = 17.5 Hz, 1 H), 2.35 (dd, *J* = 5.5, 5.0 Hz, 2 H); ¹³C NMR (125 MHz, CDCl₃) δ 174.6, 170.4, 170.3, 129.7, 126.1, 85.8, 82.5, 82.0, 68.9, 64.6, 58.0, 52.5, 51.6, 39.6, 30.3; HRMS (ESI, positive) calcd for C₁₅H₁₉NO₇Na [(M+Na)⁺] 348.1054, found 348.1060.

The Diester S4c. With the same procedure for the synthesis of **S4a**, **S4c** (589.6 mg, 93%) was obtained as a white solid starting from **13c** (604.4 g, 1.03 mmol), 2-methyl-2-butene (547.0 μ L, 5.16 mmol), NaH₂PO₄·2H₂O (177.0 mg, 1.13 mmol), NaClO₂ (278 mg, 3.09 mmol), and TMSCHN₂ (2 M in Et₂O, 1.03 mL, 2.06mmol).

Data for S4c. IR (film) 2953, 1745, 1699, 1544, 1513, 1248, 1172, 1031, 682 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.36 (dd, *J* = 7.5, 1.0 Hz, 1 H), 7.82-7.68 (m, 3 H), 7.12 (d, *J* = 8.5 Hz, 2 H), 6.88 (d, *J* = 8.5 Hz, 2 H), 6.00 (dt, *J* = 11.0, 4.0 Hz, 1 H), 5.80 (dd, *J* = 11.0, 3.0 Hz, 1 H), 4.94 (d, *J* = 14.5 Hz, 1 H), 4.70 (t, *J* = 7.5 Hz, 1 H), 4.63 (br s, 1 H), 4.19 (s, 1 H), 4.10 (dt, *J* = 16.0, 2.0 Hz, 1 H), 3.94 (d, *J* = 14.5 Hz, 1 H), 3.91 (d, *J* = 16.0 Hz, 1 H), 3.82 (s, 3 H), 3.70 (d, *J* = 5.0 Hz, 1 H), 3.67 (s, 3 H), 3.64 (s, 3 H), 2.97 (d, *J* = 16.5 Hz, 1 H), 2.82 (d, *J* = 16.5 Hz, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ 171.5, 169.3 (× 2), 159.3, 148.2, 134.1, 132.2, 132.0, 131.4, 129.8, 129.7, 126.8, 125.9, 124.2, 11.3, 84.7, 73.1, 63.4, 58.7, 55.2, 52.8, 52.5, 51.9, 45.2, 40.9, 39.7; HRMS (ESI, positive) calcd for $C_{28}H_{29}N_3O_{11}SNa$ [(M+Na)⁺] 638.1415, found 638.1392.

The Lactam 14c. With the same procedure for the synthesis of **14a**, **14c** (178.4 mg, 78%) was obtained as a white solid starting from **S4c** (278.4 mg, 0.45 mmol) and CAN (1.24 g, 2.26 mmol).

Data for 14c. IR (film) 2922, 1715, 1541, 1362, 1253, 1166, 683, 584 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.19 (dd, *J* = 5.5, 4.5 Hz, 1 H), 7.72-7.64 (m, 3 H), 6.02 (br s, 1 H), 5.91 (dd, *J* = 10.0, 1.0 Hz, 1 H), 5.74 (d, *J* = 10.0 Hz, 1 H), 4.91 (t, *J* = 7.5 Hz, 1 H), 4.80 (br s, 1 H), 4.41 (s, 1 H), 4.13 (d, *J* = 19.0 Hz, 1 H), 3.87 (d, *J* = 19.0 Hz, 1 H), 3.75 (s, 3 H), 3.59 (s, 3 H), 3.37 (d, *J* = 7.5 Hz, 1 H), 3.05 (d, *J* = 16.5 Hz, 1 H), 2.81 (d, *J* = 16.5 Hz, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ 173.2, 169.3, 169.2, 148.0, 134.0, 132.4, 132.1, 131.9, 126.4, 124.5, 87.5, 73.7, 65.6, 58.0, 52.8, 52.0, 50.8, 40.8, 40.1; HRMS (ESI, positive) calcd for C₂₀H₂₂N₃O₁₀S [(M+H)⁺] 496.1020, found 496.1020.

The Diester S4e. With the same procedure for the synthesis of **S4a**, **S4e** (183.7 mg, 85%) was obtained as a pale yellow oil starting from **13e** (210.4 mg, 0.41 mmol), 2-methyl-2-butene (218.0 μ L, 2.06 mmol), NaH₂PO₄-2H₂O (70.7 mg, 0.46 mmol), NaClO₂ (111.3 mg, 1.24 mmol), and TMSCHN₂ (2 M in Et₂O, 0.41 mL, 0.81 mmol).

Data for S4e. IR (film) 2954, 1745, 1702, 1513, 1438, 1205, 1167, 1035 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, ca 6:4 mixture of rotamers) δ 7.12-7.01 (m, 2 H), 6.85-6.82 (m, 2 H), 5.98-5.90 (m, 1 H), 5.60-5.54 (m, 1 H), 5.07 (d, *J* = 14.5, 0.4 H), 5.00 (d, *J* = 14.5 Hz, 0.6 H), 4.99 (d, *J* = 5.5 Hz, 0.6 H), 4.72 (d, *J* = 5.5 Hz, 0.4 H), 4.15 (s, 0.6 H), 4.11-4.03 (m, 1.4 H), 3.90-3.85 (m, 2 H), 3.79-3.78 (m, 3 H), 3.70 (s, 1.2 H), 3.66-3.64 (m, 4.8 H), 3.56-3.51 (m, 0.6 H), 3.43 (s, 0.4 H), 3.37 (s, 0.6 H), 2.98-2.94 (m, 1.4 H), 2.82-2.69 (m, 1 H), 2.50-2.31 (m, 2 H); ¹³C

NMR (125 MHz, CDCl₃) δ 170.3 (× 2), 169.4, 169.1, 168.9, 168.7, 159.4, 159.3, 157.1, 156.5, 137.6, 137.0, 130.0, 129.9, 126.9, 126.8, 121.6, 120.5, 117.3, 115.0, 114.2, 114.1, 82.8, 82.6, 75.9, 75.7, 68.4, 67.4, 65.3, 64.6, 59.3, 58.6, 55.2 (× 2), 52.5, 52.4, 52.1, 52.0, 45.1 (× 2), 43.0, 41.6, 39.9, 39.7, 32.7, 29.6; HRMS (ESI, positive) calcd for C₂₅H₂₇N₂O₈F₃Na [(M+Na)⁺] 563.1612, found 563.1609.

The Lactam 14e. With the same procedure for the synthesis of **14a**, **14e** (109.4 mg, 80%) was obtained as a pale yellow solid starting from **S4e** (175.5 mg, 0.33 mmol) and CAN (891 mg, 1.63 mmol).

Data for 14e. IR (film) 2930, 1716, 1436, 1209, 1146, 1046, 730 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, ca 1:1 mixture of rotamers) δ 6.15 (br s, 0.5 H), 6.09 (br s, 0.5 H), 6.03-5.97 (m, 1 H), 5.78-5.72 (m, 1 H), 5.04 (d, J = 6.0 Hz, 0.5 H), 4.75 (d, J = 6.0 Hz, 0.5 H), 4.49-4.45 (m, 2 H), 4.29 (s, 0.5 H), 4.24 (s, 0.5 H), 4.06 (dd, J = 13.5, 4.5 Hz, 0.5 H), 3.85 (d, J = 8.0 Hz, 1 H), 3.77 (s, 1.5 H), 3.76 (s, 1.5 H), 3.64 (s, 1.5 H), 3.63 (s, 1.5 H), 3.45 (ddd, J = 12.5, 10.5, 3.5 Hz, 0.5 H), 3.40 (d, J = 17.0 Hz, 0.5 H), 3.25-3.24 (m, 1 H), 3.24 (d, J = 17.0 Hz, 0.5 H), 2.96 (d, J = 17.0 Hz, 0.5 H), 2.94 (d, J = 17.0 Hz, 0.5 H), 2.82 (m, 0.5 H), 2.57-2.36 (m, 1.5 H); ¹³C NMR (125 MHz, CDCl₃) δ 173.4, 172.9, 169.6, 169.3 (× 2), 169.1, 157.1, 156.8, 156.4, 156.1, 137.7, 136.5, 121.7, 120.3, 117.4, 117.2, 115.1, 114.9, 85.5, 85.3, 75.6, 75.3, 65.2 (× 2), 64.8, 64.3, 63.6, 59.2, 59.1, 52.7, 52.6, 52.1, 520, 42.6 (× 2), 41.2, 39.9, 39.8, 32.3, 29.1; HRMS (ESI, positive) calcd for C₁₇H₁₉N₂O₇Na [(M+Na)⁺] 443.1037, found 443.1038.

The Pyrrolidine 15a. To a stirred solution of the pyrrolidinone **14a** (65.8 mg, 0.212 mmol) in DCM (2.0 mL) at 0 °C were added MeO₃·BF₄ (94.1 mg, 0.636 mmol) and K₂CO₃ (117.2 mg, 0.848 mmol). After stirring at rt for 4 h, the mixture was diluted with DCM (20 mL), washed with water (10 mL) and brine (10 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The crude imidate thus obtained was used in the next reaction without purification.

To a stirred solution of the above imidate in methanol (2.0 mL) at 0 °C were added NaCNBH₃ (40 mg, 0.636 mmol) and TFA (31.5 μ L, 0.424 mmol). After stirring at rt for 4 h, the mixture was diluted with DCM (20 mL), washed with saturated aqueous NaHCO₃ (20 mL), dried over Na₂SO₄, and concentrated under reduced pressure to a final volume of ca 2 mL. Boc₂O (149 μ L, 0.636 mmol) and TEA (88 μ L, 0.636 mmol) were added, and the mixture was stirred at rt for 2 h. The mixture was then concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (3 g, hexane/EtOAc = 8:2) to give the pyrrolidine **15a** (49.4 mg, 59 %, 3 steps) as a white solid: IR (film) 1742, 1701, 1395, 1366, 1174, 1013, 689 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, ca 7:3 mixture of rotamers) δ 6.01 (br s, 2 H), 4.74 (s, 1 H), 4.29 (br s, 0.7 H), 4.26 (br s, 0.3 H), 4.15 (d, *J* = 16.5 Hz, 1 H), 3.98 (d, *J* = 16.5 Hz, 1 H), 3.94-3.90 (m, 2 H), 3.66 (s, 2.1 H), 3.65 (s, 0.9 H), 3.37 (dd, *J* = 10.0,

4.5 Hz, 0.7 H), 3.32 (m, 0.3 H), 3.17 (d, J = 17.0 Hz, 0.7 H), 3.14 (d, J = 17.0 Hz, 0.3 H), 3.07 (br d, J = 6.0 Hz, 0.3 H), 3.00 (dd, J = 10.0, 4.5 Hz, 0.7 H), 2.68 (d, J = 17.0 Hz, 0.3 H), 2.62 (d, J = 17.0 Hz, 0.7 H); ¹³C NMR (125 MHz, CDCl₃, selected) δ 171.0, 170.3, 154.0, 130.5, 122.7, 92.0, 81.1, 80.4, 73.0, 69.0, 64.0, 52.0, 51.6, 51.5, 48.7, 40.4, 28.2; HRMS (ESI, positive) calcd for C₁₉H₂₇N₁O₈Na [(M+Na)⁺] 420.1629, found 420.1622.

The Pyrrolidine 15b. With the same procedure for the synthesis of **15a**, **15b** (10.4 mg, 71%) was obtained as a white solid starting from **14b** (9.7 mg, 0.030 mmol), $MeO_3 \cdot BF_4$ (13.2 mg, 0.090 mmol), and $NaCNBH_3$ (5.62 mg, 0.089 mmol).

Data for 15b. IR (film) 1745, 1701, 1396, 1171, 669 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, ca 7:3 mixture of rotamers) δ 5.76 (dt, J = 12.0, 4.5 Hz, 1 H), 5.67 (dd, J = 12.0, 4.5 Hz, 1 H), 4.68-4.64 (br s, 2 H), 3.95-3.88 (m, 3 H), 3.71-3.61 (m, 6 H), 3.52-3.48 (m, 1 H), 3.32 (dd, J = 11.5 Hz, 0.7 H), 3.26 (m, 1 H), 3.16 (d, J = 17.0 Hz, 0.7 H), 3.12 (d, J = 17.0 Hz, 0.3 H), 3.05 (dd, J = 10.0, 5.0 Hz, 0.3 H), 2.97 (dd, J = 10.0, 5.0 Hz, 0.7 H), 2.67 (d, J = 17.0 Hz, 0.3 H), 2.60 (d, J = 17.0 Hz, 0.7 H), 2.36-2.26 (m, 2 H); ¹³C NMR (125 MHz, CDCl₃, selected) δ 171.0, 170.3, 154.0, 129.5, 126.8, 90.5, 85.5, 82.1, 80.4, 68.9, 68.8, 52.9, 52.0, 51.5, 49.5, 39.7, 30.6, 28.2; HRMS (ESI, positive) calcd for C₂₀H₂₉NO₈Na [(M+Na)⁺] 434.1785, found 434.1788.

The Pyrrolidine 15c. With the same procedure for the synthesis of **15a**, **15c** (162.1 mg, 86%) was obtained as a pale yellow solid starting from **14c** (160.1 mg, 0.323 mmol), $MeO_3 \cdot BF_4$ (143.3 mg, 0.969 mmol), and $NaCNBH_3$ (101.5 mg, 1.615 mmol).

Data for 15c. IR (film) 2977, 1747, 1698, 1542, 1364, 1168, 682 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, ca 1:1 mixture of rotamers) δ 8.08 (d, *J* = 8.0 Hz, 0.5 H), 8.02 (d, *J* = 8.0 Hz, 0.5 H), 7.74-7.67 (m, 3 H), 5.81 (dd, *J* = 10.8, 3.5 Hz, 0.5 H), 5.75 (dd, *J* = 10.8, 3.5, Hz, 0.5 H), 5.65-5.62 (m, 1 H), 4.63 (d, *J* = 7.0 Hz, 0.5 H), 4.56 (d, *J* = 7.0 Hz, 0.5 H), 4.52 (s, 0.5 H), 4.51-4.45 (m, 1 H), 4.37 (s, 1 H), 4.15 (br d, *J* = 18.0 Hz, 0.5 H), 4.01 (d, *J* = 11.5 Hz, 0.5 H), 3.96 (br d, *J* = 18.0 Hz, 0.5 H), 3.81-3.77 (m, 1 H), 3.69 (s, 1.5 H), 3.67 (s, 1.5 H), 3.62 (dd, *J* = 11.8, 5.5 Hz, 0.5 H), 3.59 (s, 3 H), 3.58 (d, *J* = 11.5 Hz, 0.5 H), 3.51 (dd, *J* = 11.8, 5.5 Hz, 0.5 H), 3.06 (m, 1 H), 2.83 (d, *J* = 15.5 Hz, 0.5 H), 2.77 (d, *J* = 15.5 Hz, 0.5 H), 2.72 (d, *J* = 15.5 Hz, 0.5 H), 2.67 (d, *J* = 15.5 Hz, 0.5 H), 1.46 (s, 4.5 H), 1.39 (s, 4.5 H); ¹³C NMR (125 MHz, CDCl₃) δ 170.3, 170.0, 169.8, 169.7, 154.6, 153.7, 147.8, 147.6, 134.1, 134.0, 132.8, 132.6, 132.4, 132.0, 131.3, 131.0, 126.9, 126.7, 125.1, 124.7 (x2), 124.2, 91.3, 90.7, 80.8, 80.6, 71.9, 71.6, 70.4, 69.8, 59.1, 58.8, 52.2, 52.1, 51.8, 48.5, 48.2, 46.7, 45.4, 40.5, 39.7, 39.5, 28.2 (× 2); HRMS (ESI, positive) calcd for C₂₅H₃₁N₃O₁₁Na [(M+H)⁺] 604.1571, found 604.1566.

The Pyrrolidine 15e. With the same procedure for the synthesis of 15a, 15c (110.1 mg,

84%) was obtained as a pale yellow solid starting from **14c** (109.0 mg, 0.259 mmol), $MeO_3 \cdot BF_4$ (114.9 mg, 0.777 mmol), and $NaCNBH_3$ (48.8 mg, 0.777 mmol).

Data for 15e. IR (film) 1746, 1688, 1394, 1211, 1143, 731 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, ca 6:4 mixture of rotamers) δ 5.98-5.89 (m, 1 H), 5.75-5.68 (m, 1 H), 4.70 (s, 0.6 H), 4.54-4.50 (m, 1.4 H), 4.44-4.37 (m, 1 H), 4.01-3.98 (m, 1 H), 3.92-3.78 (m, 3 H), 3.92-3.78 (m, 3 H), 3.71 (s, 3 H), 3.65 (s, 3 H), 3.45 (m, 0.6 H), 3.14 (m, 0.4 H), 3.00-2.97 (m, 1 H), 2.90-2.81 (m, 1.6 H), 2.73 (t, J = 16.0 Hz, 0.4 H), 2.49 (br s, 0.6 H), 2.42-2.30 (m, 1.4 H), 1.45 (br s, 3.6 H), 1.39 (s. 5.4 H); ¹³C NMR (125 MHz, CDCl₃, selected) δ 170.5, 169.6, 157.3, 153.3, 136.6, 121.7, 117.5, 90.8, 81.0, 75.9, 68.6 (× 2), 54.7, 51.9, 51.0, 42.2, 39.0, 32.7, 29.6, 28.1; HRMS (ESI, positive) calcd for C₂₂H₂₉N₂O₈Na [(M+Na)⁺] 529.1768, found 529.1753.

The Pyrrolidine 15f. To a stirred solution of 15c (141.2 mg, 0.243 mmol) in CH₃CN (3 mL) at 0 °C were added thiophenol (49.8 µL, 0.485 mmol) and Cs₂CO₃ (119.0 mg, 0.365 mmol). After stirring at rt for 1.5 h, the mixture was diluted with chloroform (50 mL), washed with saturated aqueous NaHCO₃ (25 mL), dried over Na₂SO₄, and concentrated under reduced pressure to a final volume of ca 3 mL. Boc₂O (170.9 μL, 0.729 mmol) and pyridine (59 μL, 79.1 mmol) were added, and the mixture was stirred at rt for 2 h. The mixture was then diluted with DCM (50 mL), washed with saturated aqueous NH₄Cl (30 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (2 g, hexane/EtOAc = 8:2) to give the N-Boc pyrrolidine 15f (103.3 mg, 86%) as a colorless solid: IR (film) 2976, 1746, 1702, 1395, 1367, 1171, 681 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, ca 1:1 mixture of rotamers) δ 5.78 (d, J = 9.0 Hz, 1 H), 5.62 (dd, J = 9.0, 2.0 Hz, 1 H), 4.87 (br s, 1 H), 4.58 (br s, 1 H), 4.48 (s, 0.4 H), 4.40 (s, 0.6 H), 4.20 (br d, J = 19.0 Hz, 1 H), 3.93 (d, J = 11.0 Hz, 0.6 H), 3.84-3.82 (m, 0.4 H), 3.69 (s, 3 H), 3.67-3.58 (m, 1 H), 3.60 (s, 3 H), 3.55 (br d, J = 19.0 Hz, 1 H), 2.96-2.91 (m, 1 H), 2.83-2.65 (m, 2 H); ¹³C NMR (125 MHz, CDCl₃, selected) δ 170.6, 169.7, 154.3, 153.6, 125.6, 125.3, 91.2, 80.4, 72.5, 70.5, 56.8, 52.1, 52.0, 51.8, 49.3, 47.6, 46.3, 40.5, 28.2, 28.1; HRMS (ESI, positive) calcd for $C_{24}H_{36}N_2O_9Na$ [(M+Na)⁺] 519.2313, found 519.2294.

The Tetrahydropyran S5a. To a stirred solution of **15a** (5.4 mg, 0.0136 mmol) in methanol (1.0 mL) at rt was added palladium (10 wt% on carbon, 0.55 mg). The mixture was stirred vigorously under hydrogen atmosphere (1atm) for 1 h. The catalyst was then removed by filtration and the filtrate was then concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (0.5 g, hexane/EtOAc = 8:2) to give **S5a** (5.4 mg, 100%) as a white solid: IR (film) 1742, 1701, 1391, 1250, 1169, 1105, 1041, 897 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, mixture of rotamers) δ 4.72 (br s, 1 H), 4.05 (br s, 1 H),

3.84 (m, 2 H), 3.78 (br s, 1 H), 3.67 (m, 6 H), 3.34-3.14 (m, 3 H), 2.92-2.82 (m, 1 H), 2.76-2.66 (m, 1 H), 2.10 (m, 1 H), 1.80 (m, 1 H), 1.64 (m, 1 H), 1.42-1.38 (m, 9 H), 1.32 (m, 1 H); 13 C NMR (125 MHz, CDCl₃, selected) δ 171.1, 170.5, 154.1, 91.7, 81.8, 80.5, 75.8, 69.5, 6.4, 52.0, 51.6, 48.6, 40.3, 28.3, 28.2, 24.9, 19.8; HRMS (ESI, positive) calcd for C₁₉H₂₉NO₈Na [(M+Na)⁺] 422.1785, found 422.1791.

The Oxepane S5b. With the same procedure for the synthesis of **S5a**, **S5b** (2.4 mg, 96%) was obtained as a white solid starting from **15b** (2.5 mg, 0.028 mmol) and palladium (10 wt% on carbon, 0.3 mg).

Data for S5b. IR (film) 1701, 1384, 1070 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, mixture of rotamers) δ 4.67 (br s, 1 H), 3.30 (m, 1 H), 4.10 (m, 1 H), 3.91-3.86 (m, 2 H), 3.67 (m, 6 H), 3.31-3.19 (m, 2 H), 3.14-3.04 (m, 1 H), 2.99-2.89 (m, 1 H), 2.61-2.53 (m, 1 H), 2.09 (m, 1 H), 1.77-1.65 (m, 4 H), 1.42-1.37 (m, 9 H), 1.27-1.20 (m, 1 H); ¹³C NMR (125 MHz, CDCl₃, selected) δ 171.3, 170.3, 154.0, 91.2, 90.0, 83.9, 80.4, 74.0, 68.7, 52.9, 52.0, 51.6, 49.4, 39.6, 32.1, 31.8, 28.2, 21.4; HRMS (ESI, positive) calcd for C₂₀H₃₁NO₈Na [(M+Na)⁺] 436.1941, found 436.1946.

The Piperidine S5f. With the same procedure for the synthesis of **S5a**, **S5f** (32.8 mg, 100%) was obtained as a white solid starting from **15f** (32.8 mg, 0.066 mmol) and palladium (10 wt% on carbon, 3.3 mg).

Data for S5f. IR (film) 1747, 1696, 1393, 1367, 1254, 1165, 1063, 770 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, mixture of rotamers) δ 4.49-4.45 (m, 1 H), 4.28-4.21 (m, 2 H), 3.83-3.75 (m, 1 H), 3.73-3.69 (m, 4 H), 3.67-3.62 (m, 3 H), 3.55-3.50 (m, 1 H), 3.29-3.11 (m, 2 H), 2.76-2.64 (m, 2 H), 1.68-1.37 (m, 22 H); ¹³C NMR (125 MHz, CDCl₃, selected) δ 171.2, 170.2, 156.0, 153.9, 90.9, 80.8, 74.9, 70.3, 61.0, 60.3, 52.2, 52.1, 50.3, 49.3, 41.0, 39.8, 28.6, 28.4, 26.1, 19.4; HRMS (ESI, positive) calcd for C₂₄H₃₈N₂O₉Na [(M+Na)⁺] 521.2469, found 521.2466.

The Azepane S5e. With the same procedure for the synthesis of **S5a**, **S5e** (34.8 mg, 100%) was obtained as a white solid starting from **15e** (34.7 mg, 0.069 mmol) and palladium (10 wt% on carbon, 3.5 mg).

Data for S5e. IR (film) 1747, 1692, 1393, 1209, 1016, 733 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, ca 6:4 mixture of rotamers) δ 5.05 (m, 0.4 H), 4.45-4.40 (m, 1.6 H), 4.33-4.32 (m, 1 H), 4.07 (br d, 0.6 H), 3.98 (m, 0.4 H), 3.89 (m, 0.6 H), 3.81-3.74 (m, 3.4 H), 3.67-3.62 (m, 4 H), 3.19-3.01 (m, 1.4 H), 2.88-2.73 (m, 2.6 H), 2.22 (m, 1 H), 1.85-1.38 (m, 14 H); ¹³C NMR (125 MHz, CDCl₃, selected) δ 170.4, 170.0, 153.9, 153.4, 117.7, 90.5, 81.1, 78.3, 67.7, 66.5, 53.3, 52.1, 52.0, 44.7, 37.6, 30.8, 28.2, 28.1, 27.4, 20.9; HRMS (ESI, positive) calcd for C₂₂H₃₁N₂O₈F₃Na [(M+Na)⁺] 531.1925, found 531.1936.

The Diol S6a. To a stirred solution of 15a (5.40 mg, 0.0136 mmol) in *tert*-butanol (0.2 mL) at rt was added a solution of NMO (100 mg, 0.85 mmol) in water (0.2 mL) and OsO₄ (3.9 mM in *tert*-butanol, 33 μL, 0.0014 mmol). After 3 h, saturated aqueous Na₂S₂O₄ (2 mL) was added, and the mixture was extracted with chloroform (3 × 5 mL). The combined extracts were washed with brine (2 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (0.5 g, methanol/chloroform = 2:98) to give the diol S6a (5.9 mg, 100%) as a white solid: IR (film) 3400, 1743, 1693, 1401, 1250, 1167, 1090 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, mixture of rotamers) δ 4.71 (m, 1 H), 4.16-4.10 (m, 2 H), 4.01 (br s, 1 H), 3.92 (br s, 1 H), 3.81 (m, 1 H), 3.70-3.56 (m, 6 H), 3.44 (m, 1 H), 3.34-3.27 (m, 1 H), 3.08 (m, 1 H), 2.97-2.80 (m, 2 H), 2.64 (m, 1 H), 1.42-1.38 (m, 9 H); ¹³C NMR (125 MHz, CDCl₃, selected) δ 170.8, 170.3, 154.1, 91.9, 81.6, 80.8, 79.2, 69.3, 66.3, 64.4, 64.0, 52.1, 51.7, 50.9, 48.6, 40.2, 28.2; HRMS (ESI, positive) calcd for C₁₉H₂₉NO₁₀Na [(M+Na)⁺] 454.1684, found 454.1678.

The Diol S6b. With the same procedure for the synthesis of **S6a**, **S6b** (2.9 mg, 100%) was obtained as a white solid starting from **15b** (2.7 mg, 6.57 μ mol).

Data for S6b. IR (film) 3406, 1742, 1694, 1394, 1171, 1101, 1074, 904, 756 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, mixture of rotamers) δ 4.62 (br s, 1 H), 4.38 (m, 1 H), 4.19 (br s, 1 H), 4.13-4.09 (m, 1 H), 3.91-3.84 (m, 2 H), 3.68-3.64 (m, 6 H), 3.32-3.24 (m, 1 H), 3.08-3.03 (m, 1 H), 2.98-2.90 (m, 2 H), 2.60-2.51 (m, 2 H), 1.92-1.86 (m, 1 H), 1.80-1.76 (m, 1 H), 1.42-1.38 (m, 9 H); ¹³C NMR (125 MHz, CDCl₃, selected) δ 171.1, 170.2, 153.8, 91.2, 85.5, 86.0, 80.6, 77.2, 71.8, 68.6, 67.7, 52.9, 52.1, 51.8, 49.1, 39.3, 34.7, 28.2; HRMS (ESI, positive) calcd for C₂₀H₃₁NO₁₀Na [(M+Na)⁺] 468.1840, found 468.1840.

The Diol S6f. With the same procedure for the synthesis of S6a, S6f (34.5 mg, 100%) was obtained as a white solid starting from **15f** (32.0 mg, 0.065 mmol).

Data for S6f. IR (film) 3412, 1744, 1698, 1396, 1367, 1170, 1133, 1057, 755 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, ca 7:3 mixture of rotamers) δ 4.70-4.62 (m, 1 H), 4.48 (br s, 0.3 H), 4.39 (br s, 0.7 H), 4.24 (m, 1 H), 3.96 (br s, 1 H), 3.89-3.81 (m, 1 H), 3.73 (br s, 3 H), 3.67-3.56 (m, 4 H), 3.23 (m, 1 H), 3.07-2.99 (m, 1 H), 2.68-2.65 (m, 2 H), 2.55-2.50 (m, 1 H), 2.26 (br s, 1 H), 1.47-1.44 (m, 18 H); ¹³C NMR (125 MHz, CDCl₃, selected) δ 170.8, 170.2, 155.7, 153.5, 90.6, 80.7, 79.0, 70.5, 66.4, 57.7, 52.3, 52.1, 48.8, 47.6, 46.8, 43.2, 38.8, 28.2, 28.1; HRMS (ESI, positive) calcd for C₂₄H₃₈N₂O₁₁Na [(M+Na)⁺] 553.2368, found 553.2366.

The Diol S6e. With the same procedure for the synthesis of S6a, S6e (35.1 mg, 100%) was obtained as a white solid starting from **15f** (33.1 mg, 0.065 mmol).

Data for S6e. IR (film) 3413, 1744, 1690, 1395, 1210, 1144, 1049, 756 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, ca 6:4 mixture of rotamers) δ 5.28 (s, 0.6 H), 5.07 (m, 0.4 H), 4.49-4.31 (m, 3.5

H), 4.00-3.89 (m, 2.5 H), 3.75-3.61 (m, 7 H), 3.29-2.77 (m, 4 H), 2.35-1.80 (m, 4 H), 1.45 (br s, 5.4 H), 1.39 (br s, 3.6 H); ¹³C NMR (125 MHz, CDCl₃, selected) δ 170.5, 170.0, 154.2, 153.6, 115.6, 91.3, 81.7, 78.0, 73.4, 69.7, 68.0, 67.0, 52.4, 52.3, 50.3, 39.6, 37.9, 29.3, 28.4, 28.3; HRMS (ESI, positive) calcd for C₂₂H₃₁N₂O₁₀F₃Na [(M+Na)⁺] 563.1823, found 563.1827.

General Procedures for the Synthesis of the Glutamate Analogues 16-27. A suspension of fully protected glutamate analogues (**15** series, **S5** series, and **S6** series) in hydrochloric acid (6 M, 0.5 mL) was heated at 65 °C for 10 h. The reaction mixture was then cooled to rt and concentrated under reduced pressure. The residue was purified by column chromatography on reversed-phase silica gel (500 mg, water). The active fractions were lyophilized to afford the glutamate analogues **16-27**.

The Glutamate Analogue 16. With the general procedure above, **15a** (13.6 mg, 0.034 mmol) was deprotected to give the glutamate analogue **16** (8.2 mg, 79%) as a white solid: IR (film) 1713, 1634, 1402, 1029 cm⁻¹; ¹H NMR (500 MHz, D₂O) δ 6.11 (dd, *J* = 10.0, 3.5 Hz, 1 H), 5.91 (ddd, *J* = 10.0, 3.5, 2.0 Hz, 1 H), 4.49 (s, 1 H), 4.43 (t, *J* = 1.5 Hz, 1 H), 4.11 (dd, *J* = 17.3, 3.5 Hz, 1 H), 4.05 (d, *J* = 2.5 Hz, 1 H), 4.02 (d, *J* = 17.3 Hz, 1 H), 3.93 (dd, *J* = 12.5, 10.0 Hz, 1 H), 3.20 (d, *J* = 17.0 Hz, 1 H), 3.15 (dd, *J* = 12.5, 8.5 Hz, 1 H), 3.10 (t, *J* = 8.5 Hz, 1 H), 2.88 (t, *J* = 17.0 Hz, 1 H); ¹³C NMR (125 MHz, D₂O/CD₃OD = 15:1) δ 174.0, 168.6, 132.4, 120.4, 90.6, 79.1, 72.8, 67.3, 64.3, 52.2, 45.7, 40.5; HRMS (ESI, positive) calcd for C₁₂H₁₆NO₆ [(M+H)⁺] 270,0978, found 270.0976.

The Glutamate Analogue 17. With the general procedure above, **15b** (9.4 mg, 0.023 mmol) was deprotected to give the glutamate analogue **17** (7.4 mg, 100%) as a white solid: IR (film) 1715, 1621, 1405, 1361, 1075 cm⁻¹; ¹H NMR (500 MHz, D₂O) δ 5.88 (ddd, *J* = 11.5, 5.5, 5.5 Hz, 1 H), 5.60 (dd, *J* = 1.5, 4.0 Hz), 4.75 (s, 1 H), 4.16 (s, 1 H), 4.14 (d, *J* = 3.0 Hz, 1 H), 3.85 (dd, *J* = 17.5, 1 H), 3.12-3.02 (m, 2 H), 2.81 (d, *J* = 17.5 Hz, 1 H), 2.37-2.21 (m, 2 H); ¹³C NMR (125 MHz, D₂O/CD₃OD = 15:1) δ 174.6, 170.0, 132.0, 124.8, 88.8, 83.7, 81.0, 69.1, 67.6, 53.6, 46.2, 40.7, 29.7; HRMS (ESI, positive) calcd for C₁₃H₁₈NO₆ [(M+H)⁺] 284.1129, found 284.1128.

The Glutamate Analogue 18. With the general procedure above, **15f** (23.7 mg, 0.048 mmol) was deprotected to give the glutamate analogue **18** (14.7 mg, 90%) as a white solid: IR (film) 1713, 1624, 1417, 1257, 1085, 967 cm⁻¹; ¹H NMR (500 MHz, D₂O) δ 6.11 (dd, *J* = 10.3, 4.5 Hz, 1 H), 6.07 (br d, 10.3 Hz, 1 H), 4.75 (br s, 1 H), 4.34 (s, 1 H), 3.99 (dd, *J* = 12.5, 9.0 Hz, 1 H), 3.86 (d, *J* = 4.0 Hz, 1 H), 3.78 (dd, *J* = 17.3, 4.0 Hz, 1 H), 3.65 (d, *J* = 17.3 Hz, 1 H), 3.48 (t, *J* = 9.0 Hz, 1 H), 3.32 (dd, *J* = 12.5, 9.0 Hz, 1 H), 3.08 (br s, 2 H); ¹³C NMR (125

MHz, $D_2O/CD_3OD = 15:1$) δ 174.2, 168.4, 126.7, 121.7, 89.3, 70.2, 65.5, 60.3, 49.1, 46.2, 42.3, 38.3; HRMS (ESI, positive) calcd for $C_{12}H_{17}N_2O_5$ [(M+H)⁺] 269.1131, found 269.1130.

The Glutamate Analogue 19. With the general procedure above, 15e (31.4 mg, 0.062 mmol) was deprotected to give the glutamate analogue 19 (20.1 mg, 91%) as a white solid: IR (film) 1730, 1624, 1405, 1243, 1087, 991 cm⁻¹; ¹H NMR (500 MHz, D₂O) δ 5.92 (m, 1 H), 5.69 (d, *J* = 11.0 HZ, 1 H), 5.13 (br s, 1 H), 4.15 (s, 1 H), 4.11 (d, *J* = 4.5 Hz, 1 H), 3.97 (dd, *J* = 13.0, 10.5 Hz, 1 H), 3.51 (t, *J* = 8.5 Hz, 1 H), 3.31-3.25 (m, 3 H), 3.09 (d, *J* = 18.0 Hz, 1 H), 2.92 (d, *J* = 18.0 Hz, 1 H), 2.55 (m, 1 H), 2.27 (m, 1 H); ¹³C NMR (125 MHz, D₂O/CD₃OD = 15:1) δ 173.3, 167.9, 127.7, 125.8, 87.9, 78.1, 65.2, 61.9, 51.1, 46.8, 45.4, 38.6, 21.4; HRMS (ESI, positive) calcd for C₁₃H₁₉N₂O₅ [(M+H)⁺] 283.1288, found 283.1296.

The Glutamate Analogue 20. With the general procedure above, **S5a** (5.4 mg, 0.014 mmol) was deprotected to give the glutamate analogue **20** (2.6 mg, 63%) as a white solid: IR (film) 1717, 1704, 1419, 1199, 1104, 1050 cm⁻¹; ¹H NMR (500 MHz, D₂O) δ 4.22 (s, 1 H), 4.18 (s, 1 H), 3.90 (s, 1 H), 3.83 (dd, *J* = 12.8, 10.0 Hz, 1 H), 3.78 (d, *J* = 13.5 Hz, 1 H), 3.32 (t, *J* = 12.0 Hz, 1 H), 3.17 (d, *J* = 16.5 Hz, 1 H), 3.06 (dd, *J* = 12.8, 9.0 Hz, 1 H), 2.95 (t, *J* = 9.0 Hz, 1 H), 2.92 (d, *J* = 16.5 Hz, 1 H), 1.97 (br d, *J* = 14.0 Hz, 1 H), 1.81-1.65 (m, 2 H), 1.34 (d, *J* = 16.5 Hz, 1 H); ¹³C NMR (125 MHz, D₂O/CD₃OD = 15:1) δ 174.2, 168.7, 89.9, 78.9, 75.4, 67.5, 66.5, 52.7, 45.2, 40.4, 23.5, 19.2; HRMS (ESI, positive) calcd for C₁₂H₁₈NO₆ [(M+H)⁺] 272.1129, found 272.1133.

The Glutamate Analogue 21. With the general procedure above, **S5b** (2.4 mg, 0.014 mmol) was deprotected to give the glutamate analogue 21 (1.4 mg, 77%) as a white solid: IR (film) 1716, 1635, 1397, 1085, 1062, 979 cm⁻¹; ¹H NMR (500 MHz, D₂O) δ 4.39 (ddd, *J* = 7.0, 6.0, 3.5 Hz, 1 H), 4.22 (s, 1 H), 4.10 (d, *J* = 3.5 Hz, 1 H), 4.03 (d, *J* = 12.0 Hz, 1 H), 3.91 (t, *J* = 11.0 Hz, 1 H), 3.29 (m, 1 H), 3.11 (d, *J* = 16.5 Hz, 1 H), 3.09 (d, *J* = 7.5 Hz, 1 H), 2.93 (dd, *J* = 11.0, 7.5 Hz, 1 H), 2.82 (d, *J* = 16.5 Hz, 1 H), 2.06 (m, 1 H), 1.71 (m, 1 H), 1.64-1.60 (m, 3 H), 1.25 (m, 1 H); ¹³C NMR (125 MHz, D₂O/CD₃OD = 15:1) δ 174.6, 170.2, 90.2, 88.9, 83.6, 74.2, 68.2, 53.4, 46.9, 41.0, 31.5, 31.0, 20.9; HRMS (ESI, positive) calcd for C₁₃H₁₉NO₆ [(M+H)⁺] 286.1293, found 286.1285.

The Glutamate Analogue 22. With the general procedure above, S5f (18.1 mg, 0.036 mmol) was deprotected to give the glutamate analogue 22 (12.3 mg, 100%) as a white solid: IR (film) 1715, 1625, 1404, 1255, 1031, 975 cm⁻¹; ¹H NMR (500 MHz, D₂O) δ 4.47 (br s, 1 H), 4.25 (s, 1 H), 3.90 (t, *J* = 11.5 Hz, 1 H), 3.71 (s, 1 H), 3.37 (d, *J* = 12.5 Hz, 1 H), 3.32 (t, *J* = 9.0 Hz, 1 H), 3.22 (s, 1 H), 3.19 (dd, *J* = 12.5, 9.0 Hz, 1 H), 3.15 (d, *J* = 18.0 Hz, 1 H), 3.02 (d, *J* = 18.0 Hz, 1 H), 2.87 (t, *J* = 12.5 Hz, 1 H), 2.11 (d, *J* = 15.5 Hz, 1 H), 1.87 (t, *J* = 13.0 Hz,

1 H), 1.71-1.63 (m, 2 H); ¹³C NMR (125 MHz, $D_2O/CD_3OD = 15:1$) δ 174.7, 168.4, 88.7, 72.4, 65.5, 59.7, 49.4, 45.6, 43.4, 37.2, 22.7, 16.5; HRMS (ESI, positive) calcd for $C_{12}H_{19}N_2O_5$ [(M+H)⁺] 271.1288, found 271.1291.

The Glutamate Analogue 23. With the general procedure above, **S5e** (23.5 mg, 0.046 mmol) was deprotected to give the glutamate analogue 23 (16.2 mg, 98%) as a white solid: IR (film) 1731, 1624, 1417, 1258, 1084 cm⁻¹; ¹H NMR (500 MHz, D₂O) δ 4.59 (dt, *J* = 7.5, 6.0 Hz, 1 H), 4.33 (s, 1 H), 3.97 (dd, *J* = 12.8, 10.0 Hz, 1 H), 3.86 (d, *J* = 4.5 Hz, 1 H), 3.44 (br s, 1 H), 3.43 (t, *J* = 9.0 Hz, 1 H), 3.28 (dd, *J* = 12.5, 9.0 Hz, 1 H), 3.08 (d, *J* = 18.0 Hz, 1 H), 2.99 (d, *J* = 18.0 Hz, 1 H), 2.94 (dd, *J* = 13.5, 3.5, 3.5 Hz, 1 H), 2.26 (m, 1 H), 1.88-1.60 (m, 4 H), 1.40 (dd, *J* = 13.5, 12.3 Hz, 1 H); ¹³C NMR (125 MHz, D₂O/CD₃OD = 15:1) δ 173.5, 168.0, 89.1, 78.4, 67.9, 65.3, 51.1, 49.4, 47.0, 38.6, 28.3, 26.4, 19.8; HRMS (ESI, positive) calcd for C₁₃H₂₁N₂O₅ [(M+H)⁺] 284.1445, found 184.1449.

The Glutamate Analogue 24. With the general procedure above, S6a (2.4 mg, 0.014 mmol) was deprotected to give the glutamate analogue 24 (2.9 mg, 74%) as a white solid: IR (film) 3419, 1716, 1634, 1403, 1240, 1088 cm⁻¹; ¹H NMR (500 MHz, D₂O) δ 4.39 (s, 1 H), 4.18 (s, 1 H), 4.07 (s, 2 H), 3.86-3.82 (m, 2 H), 3.55 (dd, *J* = 11.0, 5.0 Hz, 1 H), 3.42 (t, *J* = 11.0 Hz, 1 H), 3.13 (d, *J* = 17.0 Hz, 1 H), 3.09-3.00 (m, 2 H), 2.90 (d, *J* = 17.0 Hz, 1 H); ¹³C NMR (125 MHz, D₂O/CD₃OD = 15:1) δ 175.7, 170.3, 92.0, 82.4, 78.6, 78.5, 67.3, 65.8, 65.3, 53.2, 46.9, 42.0; HRMS (ESI, positive) calcd for C₁₂H₁₈NO₈ [(M+H)⁺] 304.1027, found 304.1232.

The Glutamate Analogue 25. With the general procedure above, S6b (5.9 mg, 0.014 mmol) was deprotected to give the glutamate analogue 25 (4.8 mg, 100%) as a white solid: IR (film) 3420, 1748, 1623, 1375, 1223, 1036 cm⁻¹; ¹H NMR (500 MHz, D₂O) δ 4.34 (s, 1 H), 4.28 (dd, *J* = 7.5, 4.5 Hz, 1 H), 4.19 (d, *J* = 4.5 Hz, 1 H), 4.10 (d, *J* = 4.0 Hz, 1 H), 3.96-3.87 (m, 3 H), 3.55 (t, *J* = 13.0 Hz, 1 H), 3.12 (d, *J* = 16.5 Hz, 1 H), 3.11 (dd, *J* = 12.5, 7.0 Hz, 1 H), 2.96 (dd, 10.8, 7.0 Hz, 1 H), 2.86 (d, *J* = 16.5 Hz, 1 H), 1.89 (ddd, *J* = 18.8, 13.0, 4.5 Hz, 1 H), 1.69 (dd, *J* = 13.3, 4.5 Hz, 1 H); ¹³C NMR (125 MHz, D₂O/CD₃OD = 15:1) δ 174.3, 170.2, 90.7, 86.7, 86.0, 77.0, 72.8, 68.4, 68.3, 53.2, 47.0, 40.4, 34.4; HRMS (ESI, positive) calcd for C₁₃H₂₀NO₈ [(M+H)⁺] 318.1183, found 318.1193.

The Glutamate Analogue 26. With the general procedure above, S6f (20.3 mg, 0.038 mmol) was deprotected to give the glutamate analogue 26 (13.9 mg, 97%) as a white solid: IR (film) 3300, 1714, 1627, 1404, 1256, 1101, 1000 cm⁻¹; ¹H NMR (500 MHz, D₂O) δ 4.44 (br s, 1 H), 4.25 (s, 1 H), 4.18 (br s, 1 H), 3.98 (dd, *J* = 7.8, 3.5 Hz, 1 H), 3.89 (dd, *J* = 12.8, 10.0 Hz, 1 H), 3.85 (d, *J* = 2.5 Hz, 1 H), 3.36 (t, *J* = 8.5 Hz, 1 H), 3.22 (t, *J* = 13.5 Hz, 1 H),

2.99 (d, J = 17.5 Hz, 1 H); ¹³C NMR (125 MHz, D₂O/CD₃OD = 15:1) δ 174.7, 168.4, 89.1, 77.8, 65.4, 64.9, 62.8, 57.4, 48.3, 45.6, 41.2, 37.1; HRMS (ESI, positive) calcd for C₁₂H₁₉N₂O₇ [(M+H)⁺] 303.1187, found 303.1190.

The Glutamate Analogue 27. With the general procedure above, S6e (23.3 mg, 0.043 mmol) was deprotected to give the glutamate analogue 27 (16.7 mg, 100%) as a white solid: IR (film) 3350, 1718, 1635, 1405, 1227, 1097, 991 cm⁻¹; ¹H NMR (500 MHz, D₂O) δ 4.37 (t, J = 5.5 Hz, 1 H), 4.31 (s, 1 H), 4.13 (br s, 1 H), 3.98 (dd, J = 13.0, 10.0 Hz, 1 H), 3.96 (d, J = 5.5 Hz, 1 H), 3.92 (d, J = 5.5 Hz, 1 H), 3.46 (t, J = 8.5 Hz, 1 H), 3.31 (d, J = 12.5 Hz, 1 H), 3.16 (m, 1 H), 3.11 (d, J = 18.0 Hz, 1 H), 3.00 (d, J = 18.0 Hz, 1 H), 1.98-1.97 (m, 2 H); ¹³C NMR (125 MHz, D₂O/CD₃OD = 15:1) δ 173.5, 168.0, 88.8, 81.3, 74.8, 71.3, 65.5, 65.2, 51.1, 47.0, 44.0, 38.7, 29.5; HRMS (ESI, positive) calcd for C₁₃H₂₁N₂O₇ [(M+H)⁺] 317.1343, found 317.1352.

Experimental Procedures for Biological Study

Biological assay: Mouse behavioral assay was performed as reported previously.² Behavior of animals treatred with drug was monitored for 6 h. All the animals were maintained according to the National Research Council's Guide for the Care and Use of Laboratory Animals.

Electrophysiological analyses: Effects of **16** were measured in whole-cell patch clamp assays from cultured rat hippocampal neurons. Spontaneous action potentials were recorded using the current-clamp configuration in HEPES-buffered saline solution and K-gluconate internal solution. Excitatory postynaptic currents (EPSCs) were recorded in voltage-clamp in extracellular solution containing bicuculline methiodide and picrotoxin (10 μ M each) to block GABA_A receptors. CsF/CsCl-based internal solution was used for voltage-clamp recordings. All recordings were made at room temperature. Spontaneous action potential and EPSC frequencies were significantly reduced in the presence of **16** (p<0.01, Student's paired t-test, n = 3 for each type of recording).

² R. Sakai, G. T. Swanson, K. Shimamoto, T. Green, A. Contractor, A. Ghetti, Y. Tamura-Horikawa, C. Oiwa, H. Kamiya H. *J. Pharmacol. Exp. Ther.* **2001**, *296*, 650-658.

Proposed Pathway and Its Computational Rationalization for Domino Metathesis Reaction of 7-Oxanorbornenes

The plausible reaction mechanism is shown in the scheme below. To rationalize this pathway, the geometry of two intermediates **E** and **F** were optimized by the semiempirical level of theory employing the PM3 method. First, the distance between the *upper* exocyclic amide carbonyl group and the Ru metal center in **E** was calculated to be 4.621 Å, indicating the association is rather loose and is expected to allow conversion into the alkylidene intermediate **F** smoothly.

Second, formation of a metal-alkylidene group (intermediate **F**) causes the same group moves closer to the *upper* exocyclic amide group, and the steric environment around the alkylidene group of **F** is getting crowded by their interactions. This effect controls the approaching trajectory of the CM substrate (vinyl acetate) to the metal-alkylidene group in **F**, to generate the kinetically favorable *trans*-metallacyclobutane **F'**. The four-membered ring then stereospecifically cleaved to provide the *trans*-vinylic acetate **G**. *Trans*-olefin is usually obtained selectively in the olefin metathesis reaction, and the steric interaction in the intermediate **F** further enhance the selectivity.

Finally, the triene **G** undergoes cyclization by RCM, giving rise to the heterotricycle **9a–9e**.











Scheme. The proposed pathway for the domino metathesis reaction of 7-oxanorbornenes. Chauvin's mechanism (J. L. Herisson, Y. Chauvin, *Makromol. Chem.* **1971**, *141*, 161-176) is modified here to account for our reactions. Structural optimization for the intermediates **E** and **F** was performed under high vacuum at PM3 level of theory. Benzyl (Bn) and 4-methoxybenzyl (PMB) groups were omitted in the calculation for simplicity. All calculations were performed using Spartan software. Spartan, version '06; Wavefunction, Inc., 18401 Von Karman Ave., Suite 370, Irvine, CA 92612 USA.