Supplementary Materials

I ALPHA ANGLES

Supplementary Fig. 1: The α angle is the angle made by two successive α -carbons.

II ALIGNMENT MATRIX

a)

Supplementary Fig. 2: Examples of dynamic programming matrices using Swelfe. We compute only the upper half of the matrix. The diagonal is null. The best alignment is written in bold. In reality this matrix is not kept in memory as the SIM algorithm is used (Huang and Miller, 1991; Huang, et al., 1990).

- a) Alignment of sequence data.
- b) Alignment of structures coded as a string of α -angles (scores are decimal numbers but were truncated to fine down the scheme).

III SCORES

The scores used in the SIM alignment were adapted at each level. The nucleic acids substitution scores take into account the frequency of each nucleotide to produce more meaningful alignments (Achaz, et al., 2007). For amino acids sequence alignments we allow the use of any of the standard BLOSUM or PAM matrices. The empirical structural score for two matching α angles increases when the circular difference between them decreases and also accounts for the fact that some angles are much more frequent than others (Supplementary Fig. 3). This results in very frequent angles, e.g. originating from α -helices or β-sheets, having a lower score.

The default gap opening and extension parameters were set after analysis of an extensive range of values (Supplementary Table 1). For nucleic acid sequences, penalty for a gap opening corresponds to about 4 identities. For amino acid sequences it corresponds to between 1 and 2 identitie(s). For structures it corresponds to about 6 or 7 identities. Structural opening gaps are therefore more costly but if we decrease this penalty, we rapidly increase structural alignments with high RRMSD.

Supplementary Table 1 : Default scores used at each level (S_{i,j}). pi, pj : frequencies of nucleic acids for DNA sequences and normalized frequencies of α angles i and j in PDB for structures. Δangle is the angular difference between angles i and j (from 0 to 180°). The β parameter allows to give more or less weighting to angle frequencies p_i and p_i .

		Gap	
	Matching score	opening	extension
DNA sequence	$S_{i,j} = 0.5$ x $\sigma_{(i,j)}$ x $log_4(p_ip_j)$		-1
	$\sigma_{(i,j)} = 1$ if $i \neq j$; $\sigma_{(i,j)} = -1$ if $i = j$		
Amino acid sequence	BLOSUM or PAM matrix	-8	-3
Three dimensional	$S_{i,j} = 30*(1-p_i)(1-p_i)(1-β) + β$] - Δangle	-200°	-50°
structure	$\beta = 0.4$		

Supplementary Fig. 3: frequencies of alpha angles in PDB structures (black) and in repeats found with β parameter = 0.4 (orange), β parameter = 0.5 (red), β parameter = 0.7 (blue), β parameter = 0.3 (violet). 500 best hits are kept for each $β$ parameter values.

IV WATERMAN AND VINGRON METHOD TO ASSESS STATISTICAL SIGNIFICANCE

The Waterman and Vingron method (Waterman and Vingron, 1994) was used to calculate the significance of the repeats identified by dynamic programming. The general idea of the method is to generate a large number of random sequences (see Supplementary Fig. 4) with same length and composition as the gene (codons) or the protein (amino-acids) of interest. For each random sequence we identify the best local alignment score with the same algorithm as for the real sequence. The probability to get a score better than *t* in such a random sequence is then given by

$$
P(P > t) = e^{-\gamma n^2 p^t}
$$

where n is the length of the sequence. The parameters p and γ are estimated by a weighted linear regression:

 $log(-log(P(P > t))) = log(\gamma n^2) + t log(p)$

The use of weighted regression is motivated by the observation that random sequences may have sometimes the same score and this should be accounted for. The length of the sequence is corrected by subtracting the size of the mean length of local alignments for random sequences because sequences are finite (Mott, 2000).

We also implemented the "declumping estimation" method. In this case, we generate fewer random sequences (default 20) with the method described above. Then we calculate $~50$ repeats for each random sequence with the declumping algorithm (successive repeats are independent), and we obtain about 1000 scores. In this case $\gamma n^2 p^t$ can be estimated by the average number of scores that exceed a threshold (E). The parameters p and γ are estimated by a weighted linear regression :

$$
\log(E) = \log(\gamma n^2) + t \log(p).
$$

Number of random sequences for statistics, first method.

We generate sequences of 100 AA, 300 AA and 1000 AA with the same composition in amino acids as the average composition of proteins in Uniprot. We make four different sets of experiments with 50 (blue curve in Fig. 4), 100 (green curve), 500 (orange curve) and 1000 (red curve) random sequences with same length and composition. With Swelfe, we define for each random sequence the score-threshold for a p-value of 0.01 with the Waterman-Vingron method. In the figure we represent the distribution of the thresholds for each set of experiments. We also compute the score distribution on an experiment with 10 000 random sequences (black curve). The vertical line represents the observed 99% threshold for the cumulative distribution of scores. Therefore, threshold values close to the vertical line are the most accurate. We conclude that 100 random sequences are sufficient to produce a threshold that is quite close to the real one while requiring limited computed time. Results were similar for DNA sequences (data not shown).

Supplementary Fig. 4 : Score-threshold obtained on 50 (blue), 100 (green), 500 (orange) and 1000 (red) random sequences of the same length and composition as the benchmark sequence for a p-value of 0.01.

Supplementary figure 5 : number of random sequences for statistics, declumping estimation method.

The simulations are the same as above but with the declumping estimation method. We find that 20 random sequences provide a reasonable approximation. Results were similar for other sequence length ranges (data not shown). The threshold is slightly underestimated by the declumping estimation method in proteins.

a) protein sequences (300 AA)

b) nucleic acid sequences (1000 nt)

Supplementary Fig. 5 : Score-threshold obtained on 1 (yellow), 10 (blue), 20 (green), 30 (orange), 50(red) and 80 (violet) random sequences with same length and composition as the benchmark sequence for a p-value of 0.01.

V COMPARAISON WITH DALI

Supplementary Table 2 - Analysis of similarities between protein structures lacking secondary structure elements in (Novotny, et al., 2004) when using Swelfe (A) and DALI (B). We only indicate the results for the structures that were available on the DALI server. The numbers indicate the rank of the hit between the structures in the analysis of the structures for similarities against the reference database. NS means a non-significant hit.

Supplementary Table 3 – Analysis of similarities between protein structures regarded as difficult by (Novotny, et al., 2004). The table indicates the score of the significant matches using Swelfe and DALI.

Supplementary Table 4 – Analysis of similarities between the structures of cyclophilins used by (Novotny, et al., 2004) when using Swelfe (A) and DALI (B). The numbers indicate the rank of the hit between the structures in the analysis of the structures for similarities against the reference database.

VI RRMSD

A) RRMSD of repeats

b) RMSD of repeats

Supplementary Fig.6 : a) : Best repeats were calculated on 9537 3D structures with a score threshold (250), then the Relative RMSD is calculated. The great majority of the repeats are below 0.5 which is the threshold for significant repeats (Betancourt and Skolnick, 2001). b) RMSD calculated after the RRMSD threshold of 0.5.

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