

Supplementary Information

Probing the Fibrate Binding Specificity of Rat Liver Fatty Acid Binding Protein

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Supplementary Table 1: FABP sequence alignments

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Human   MSFSGKYQLQSQENFEAFMKAIGLPEELIQKGKDIKGVSEIVQNGKHFKFTITAGSKVIQ 60
Rat     MNFSGKYQVQSQENFEPFMKAMGLPEDLIQKGKDIKGVSEIVHEGKKVKLTITYGSKVIH 60
        *.*****:*****.****:****:*****:*****:*****:*** ***:
        :

Human   NEFTVGEECELETMTGEKVKTVVQLEGDNKLVTFKNIKSVTELNGDIITNTMTLGDIVF 120
Rat     NEFTLGEECELETMTGEKVKAVVKMEGDNKMVTFKGIKSVTEFNGDTITNTMTLGDIVY 120
        *****:*****:****:****:*****.*****:*** *****:
        :

Human   KRISKRI 127
Rat     KRYSKRI 127
        **:****
    
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B

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Heart   --MADAFVGTWKLVDKSNFDDYMKSLGVGFATRQVASMTKPTTIEKNGDTITIKTHS-T 57
Adipocyte --MCDAFVGTWKLVSSENFFDDYKVEVGVGFATRQVAGMAKPNLIISVEGDLVIRSES-T 57
Testis  --MIEPFLGTWKLVSSENFFENYVRELGVCEPRKVACLIKPSVSI SFNGERMDIQAGS-A 57
Epidermal MASLKDLEGKRWLVESHGFEDYMKELGVGLALRKMGMAMAKPDCIITLDNNTLVKTES-T 59
Liver   ----MNFSGKYQVQSQENFEPFMKAMGLPEDLIQKGKDIKGVSEIVHEGKKVKLTITY-G 55
Ileal   ----MAFTGKYEFESEKNYDEFMKRLGLPDEVIERGRNFKIITEVQQDGENFTWSQSYSG 56
Intestinal ----MAFDGTWKVDRNENYEFMEKMGINVVKRKLGADHNLKLTITQEGNKFTVKESS-N 55
        : *... ..:: :. : : :..
        :

Heart   FKNTEISFQLGVEFDEVTADDRKVKSVVTLDGGKLVHVQKWD--GQETTLTRELSDGKLI 115
Adipocyte FKNTEISFQLGVEFDEITPDDRKVKSIITLDGGVLVHVQKWD--GKSTTIKRRDGDGLV 115
Testis   CRNTEISFQLGEEFEETTADNRKVKSLITFEGGSMIQIRWL--GKQTTIKRRIVDGRMV 115
Epidermal VKTTVFSTLGEKFDETTADGRKTEVCTFTD GALVQHQKWE--GKESTITRKLKDGKMV 117
Liver    SKVIHNEFTLGEECELETMTGEKVKAVVKMEGD-NKMVTFK--GIKSVT--EFNGDTIT 110
Ileal    GNIMSNKFTIGKECEMQTMGGKKFKATVKMEG--GKVVADFP--NYHQTS--EVVGDGLV 110
Intestinal FRNIDVVFELGVDFAYSLADGTELTGTWTMEGNKLVGKFKRVDNGKELI AVREISGNELI 115
        . : * . . : :. . . . . . . . . :
        :

Heart   LTLTHGNVSTRTYEKEA 133
Adipocyte VECVMKGVSTRVYERA- 132
Testis   VECTMNNVSTRTYERV- 132
Epidermal VECVMNNAICTRVYEKVQ 135
Liver    NTMTLGDIVYKRVSKRI- 127
Ileal    EISTIGDVTYERVSKRVA 128
Intestinal QTYTYEGVEAKRIFKKE- 132
        . . * :
    
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(A) Sequence alignment of the primary structures of rat FABP proteins. (* = conservative residue, : = conserved substitution, . = semi-conserved substitution. Alignment conducted using Cluster W 2.0) (B) Comparison of human and rat L-FABP. Sequence alignment of the primary structures of L-FABP.

Supplementary Table 2. Binding site occupancy

Percentage occupancy at each site for rL-FABP (300 μ M) at a 3-fold molar excess of each of the fibrate drugs used in the current study.

Ligand	% Occupancy	
	1 st Site	2 nd Site
Fenofibrate	100	100
Gemfibrozil	100	79
Bezafibrate	N/A	93
Ciprofibrate	100	85
Clofibrate	99	N/A
Fenofibric Acid	100	96

Supplementary Table 3. Temperature dependence of binding affinities

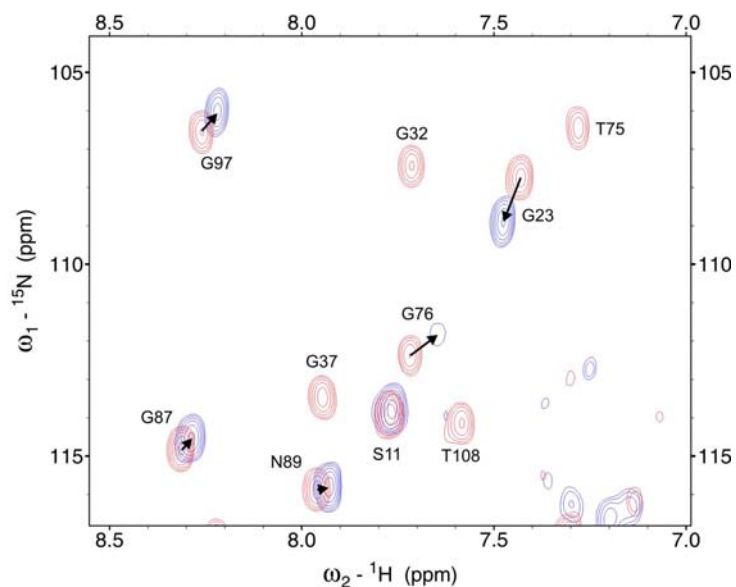
Binding affinity constants of fenofibric acid, fenofibrate and clofibrate for L-FABP measured from steady state fluorescence at different temperatures, used for van't Hoff determination of thermodynamic parameters.

Temperature (°C)	Fenofibric Acid (μM)	Fenofibrate (μM)	Clofibrate (μM)
5	$K_{d1}=0.094 \pm 0.01$ $K_{d2}=18 \pm 2.2$	$K_{d1}=0.018 \pm 0.0020$ $K_{d2}=0.13 \pm 0.020$	$K_{d1}=6.0 \pm 2.9$
10	$K_{d1}=0.10 \pm 0.02$ $K_{d2}=19 \pm 1.9$	$K_{d1}=0.023 \pm 0.0010$ $K_{d2}=0.15 \pm 0.010$	$K_{d1}=6.7 \pm 1.7$
15	$K_{d1}=0.16 \pm 0.04$ $K_{d2}=20 \pm 1.8$	$K_{d1}=0.024 \pm 0.0020$ $K_{d2}=0.16 \pm 0.020$	$K_{d1}=7.3 \pm 1.4$
20	$K_{d1}=0.22 \pm 0.030$ $K_{d2}=23 \pm 2.0$	$K_{d1}=0.027 \pm 0.0030$ $K_{d2}=0.20 \pm 0.020$	$K_{d1}=7.8 \pm 2.5$
25	$K_{d1}=0.34 \pm 0.020$ $K_{d2}=24.0 \pm 2.1$	$K_{d1}=0.032 \pm 0.0040$ $K_{d2}=0.25 \pm 0.020$	$K_{d1}=8.9 \pm 2.8$
30	$K_{d1}=0.36 \pm 0.040$ $K_{d2}=27 \pm 2.5$	$K_{d1}=0.041 \pm 0.0040$ $K_{d2}=0.27 \pm 0.040$	$K_{d1}=9.6 \pm 1.9$
37	$K_{d1}=0.42 \pm 0.030$ $K_{d2}=28 \pm 3.0$	$K_{d1}=0.062 \pm 0.006$ $K_{d2}=0.32 \pm 0.030$	$K_{d1}=11 \pm 3.4$
42	$K_{d1}=\text{ND}$ $K_{d2}=29 \pm 3.5$	$K_{d1}=\text{ND}$ $K_{d2}=0.36 \pm 0.040$	$K_{d1}=12 \pm 3.0$

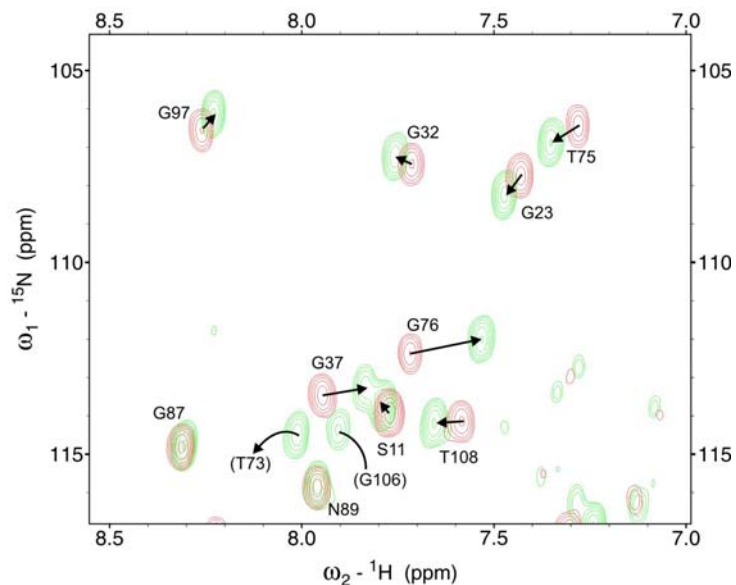
K_{d1} High affinity site; K_{d2} low affinity site; ND, Not detected; Affinity values are the mean \pm the standard deviation of three independent measurements.

Supplementary Figure 1. ^1H - ^{15}N HSQC data for rL-FABP

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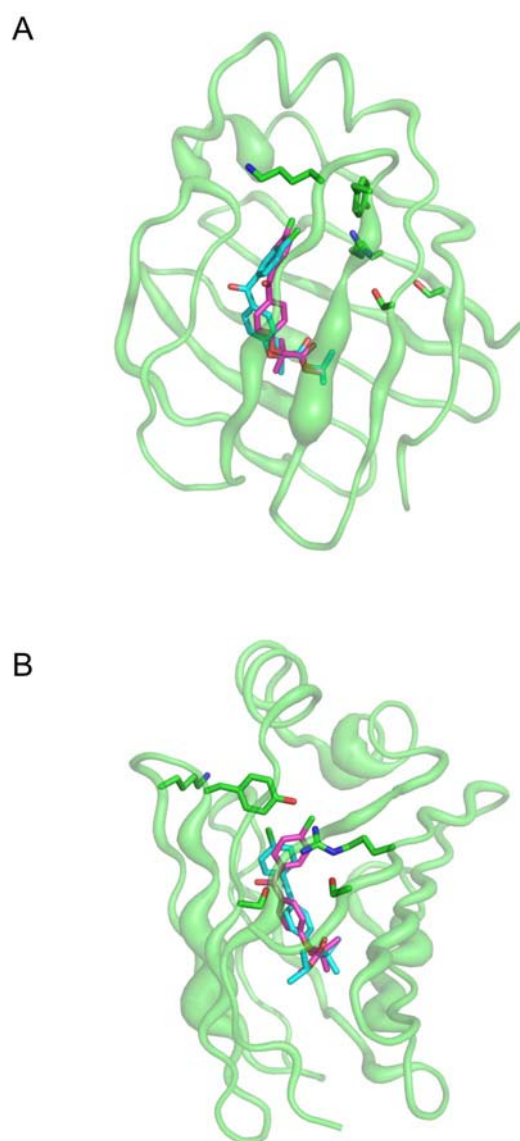


B



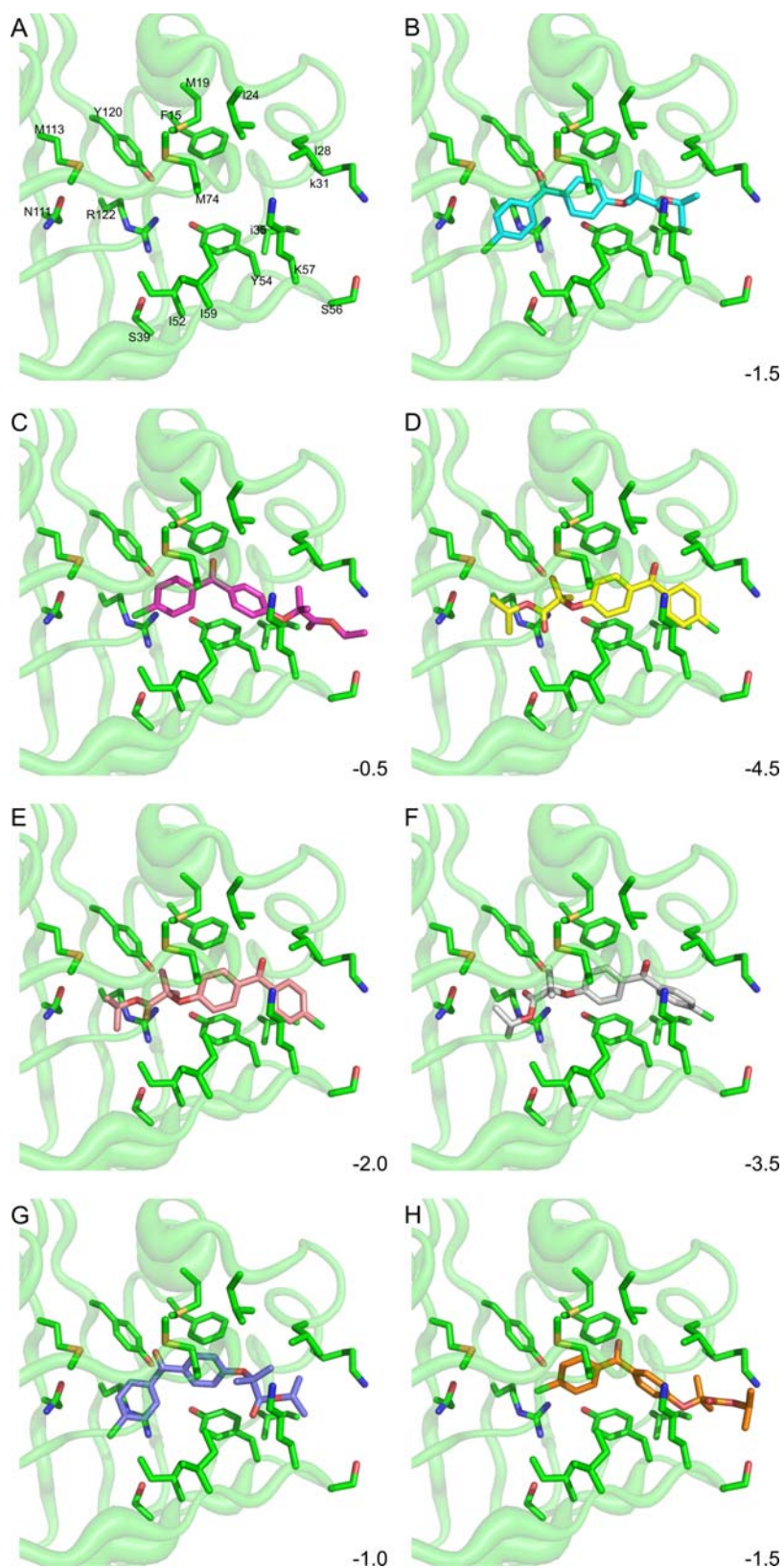
A portion of the HSQC spectrum of apo rL-FABP is shown in red and labeled with the assignment of each peak. (A) The HSQC spectrum of rL-FABP in the presence of three molar equivalents of clofibrate (green) is overlaid onto the spectrum of apo rL-FABP. The presence of clofibrate results in significant perturbation of several of the peaks. (B) The HSQC spectrum of rL-FABP in the presence of three molar equivalents of fenofibric acid (blue) is overlaid onto the spectrum of apo rL-FABP. Some peaks are perturbed upon addition of fenofibric acid, others are perturbed and demonstrate significant broadening (e.g. G76), whilst some are broadened beyond the limit of detection in the experiment (e.g. G32, G37, T75, T108).

Supplementary Figure 2. Docking of fenofibric acid methyl ester



A. A comparison of the docking clusters of fenofibrate (light blue carbon atoms) binding at the high affinity site of rL-FABP with those obtained for the methyl ester of fenofibric acid (magenta carbon atoms), which was docked using the same protocol. The poses for the two compounds are similar, suggesting that for these two compounds, the nature of the small aliphatic group in of the ester is not a dominating factor in driving the binding orientation. **B.** 90 degree rotation of the same.

Supplementary Figure 3. Docking clusters for fenofibrate



The seven docking clusters of fenofibrate along with their scores based on correlation with the CSPs observed in the NMR data.