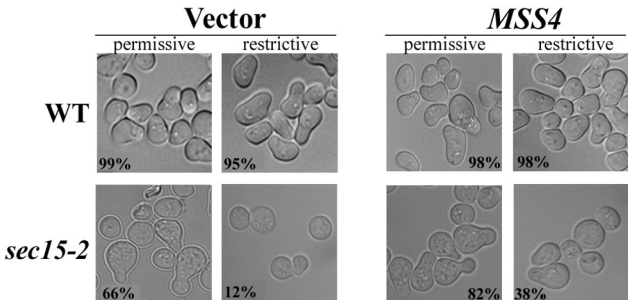
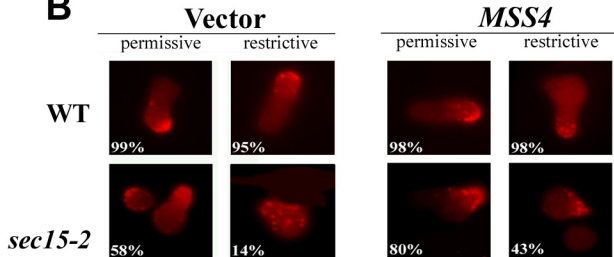


Supplementary Figure Legends

Supplementary Figure S1. *MSS4* over-expression restores shmoo formation and actin polarization in α -factor-treated *sec15-2* cells. (A) *MSS4* over-expression restores shmoo formation in a late *sec* mutant. WT and *sec15-2* cells were transformed either with a control multi-copy vector (pRS426) or a vector over-expressing *MSS4* (pRS426Mss4). Cells were grown to mid-log phase, incubated with 500nM α -factor for 2hr, and either maintained at 26°C (permissive temperature) or shifted to 37°C (restrictive temperature) for 1h. Numbers indicate the percentage of shmooing cells (n=100 cells counted). **(B) *MSS4* over-expression restores actin polarization in α -factor-treated *sec15-2* cells.** Cells from (A) were subjected to fixation and *in situ* labeling with rhodamine-conjugated phalloidin. Numbers indicate the percentage of cells in which polarized actin was observed (n=100 cells counted).

Supplementary Figure S2. *Cdc24* does not play a role in transducing the exocytic signal to *Cdc42* via *Mss4*. (A) *Cdc24* is mislocalized in late *sec* mutants in an *Mss4*-independent fashion. The indicated strains were transformed with a single-copy vector expressing *Cdc24*-GFP (pUG35-*Cdc24*-GFP) from a *MET* promoter. Cells were induced in medium lacking methionine for 1hr and either maintained at 26°C (permissive), shifted for 1hr to 35°C (semi-restrictive), or shifted for 1hr to 37°C (restrictive) prior to visualization by fluorescence confocal microscopy. Numbers indicate the percentage of cells in which bud-tip localization of *Cdc24*-GFP was observed (n=100 cells counted).

(B) CDC24 and MSS4 do not interact genetically. *cdc24-1* cells were transformed either with a control vector (pRS426) or vector over-expressing *MSS4* (pRS426Mss4). *mss4-102* cells were transformed either with a control vector (pUG35) or a single-copy vector over-expressing *CDC24* (pUG35Cdc24-GFP) under the control of a methionine-starvation inducible *MET* promoter. Cells were grown overnight in selective medium, diluted serially (5-fold), and spotted onto solid selective medium lacking methionine. Plates were incubated for 48hrs at the indicated temperatures (^oC). **(C) PI(4,5)P₂ is normally distributed in *cdc24-1* cells.** The indicated WT and *cdc24-1* cells were transformed with a multi-copy vector expressing a GFP-2XPH(PLC δ) fusion probe (pRS426GFP-2XPH(PLC δ)) and maintained at either 26^oC (permissive) or shifted for 1hr to 37^oC (restrictive). Numbers indicate the percentage of cells in which GFP-2XPH(PLC δ) localizes to the PM (n=100 cells counted).

A**B**

Supplementary Figure S1.

A**Cdc24-GFP****+MSS4****37°C****WT****26°C**

100%

35°C

100%

37°C

100%

100%

mss4-102

20%



0%



0%

95%

sec8-9

92%



89%



0%

0%

sec15-2

97%



93%



0%

0%

*cdc42-6***26°C**

98%

33°C

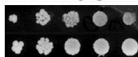
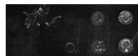
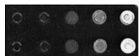
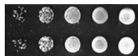
95%

35°C

0%

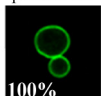
37°C

0%

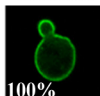
B**26°C****35°C****37°C***cdc24-1*Vector
*MSS4**mss4-102*Vector
*CDC24***C****GFP-2XPH(PLC δ)**

permissive

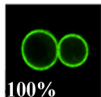
restrictive

WT

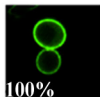
100%



100%

cdc24-1

100%



100%

Supplementary Figure S2.