

*MEIS1* Intronic Risk Haplotype Associated with Restless Legs Syndrome Affects mRNA and Protein Expression  
Levels

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**Supplementary data**

**Supplementary Table S1. Primer List for All PCR Amplifications**

Region	Fragment	Forward_Primer	Reverse_Primer	Amplicon length	Opt. T'
<b>Long-range PCR</b>	<b>LRP01</b>	GCAGGCCTACGATTGGTATTTCGGAGATC	TCATATTCTGTGCACAGAGCCAGTCTA	6861	57
	<b>LRP02</b>	TTGGGAGACGAAGAGGGTCAGTTCAGAG	AAGCAGGAGCCAGATCACCCCCTCAGT	6986	59
	<b>LRP03</b>	ACCCAGCAGTGTAATTTCCCTCCAGAT	ACCTCCCTCCCTTATCCCTAACTTCAG	6858	55
	<b>LRP04</b>	CATGGAGCTCCCGTGCTTCCAGGAATAC	TCATTTGCAGACATTCTTTCCCTCGTT	6919	58
	<b>LRP05</b>	TACATATACCTCAGTGCAGCAGTACTG	ATCAGTCCCTTTTACTTTGATCTTCAA	6927	55
	<b>LRP06</b>	ACTTAGGCAATAGAGACCTTTGAAGATC	CATTTTCCACCCACTTAGCTGCTGTFTA	6838	55
	<b>LRP07</b>	CCAGCTAAGGTTATAAACAGCAGCTAAG	AGCATTAAACCTGCACATGTGTATTAGG	6845	55
	<b>LRP08</b>	TCTCTGCAGCCAAACCAGCCTAATACA	ACTTGAGGGACATTTTCTACAGCAAACA	6853	55
	<b>LRP09</b>	TTGCTGTAGAAAATGTCCCTCAAGTAAC	TAATTAGCCCTGAGCCTCATCTGTCGTA	6870	55
	<b>LRP10</b>	GAGGGAATTTATGCATCATAACAGAGAT	AATTATTTGGGGTCATGATGGACATCTA	6852	55
	<b>LRP11</b>	TCTGGGATAATGATAAAATACACTAAAG	ACACTGTCCACCTTATAATGATTTTGAG	6879	53
	<b>LRP12</b>	CCATGCAGAAGACAAGAGGCCCCCTCAA	ACGTGTTTAGAAAACAAGGCCCATCAGC	2992	57
<b>Conserved region</b>	<b>C01</b>	ATAAATGGCATTAAAAGCGGGGAGGTAA	TGTTTGCCATATTCAGTTTTTCTTGGTA	394	53
	<b>C02.A</b>	TATTGTTGGCCACACTTTAGGATTTTAT	GATTGCTTTATTCCTTTATTGAAGTTCA	607	52
	<b>C02.B</b>	TTGGCTTTATTGACTTTCATGGGAGTTG	TATAACACCCTAAAATGGCCACCATGTC	685	55
	<b>C03</b>	GACATGGTGGCCATTTTAGGGTGTATA	ATTGTGGGGGCATAGAGATGGTTCCTAT	720	56
	<b>C04</b>	GGAACCATCTCTATGCCCCACAATAGG	CAGGGAAGAGGGACCCAAATCAACTGTC	640	58
	<b>C05</b>	GTAAGTGTACCTTTGCACTTGAAAGT	AATTAAGAATTGTCCGTTGGAGTTTGTGTC	571	51
	<b>C06</b>	TGGCCCACTGACACAGGAAGTTCTGATC	GCACAGGCTCGGGCATTATAAGACTCTA	882	58
	<b>C07.A</b>	AGAGCCCAGGAAGTTCAGGGAGAGGAT	ACCCTGCTCCTTTCCGACACAAGTTAGG	825	59
	<b>C07.B</b>	GGCCGGCGGTTCTCCAAGTTTGTAGT	CAAGCAATGCATAGTGCCTGCCTAGAAG	926	57
	<b>C07.C</b>	ATCTGGCTCCTGCTTGAAAGGTTTCTCC	TTCTCTGACTGGGCCAAGGTCATGTACC	826	57
<b>C08</b>	CTTTGGAAGAAGTTCTTTAGACCTGACT	TTGGGGTATCTATCTAAGAAATAAGTGC	530	53	
<b>C09</b>	TAAGAAGAAAGTTAGACAAACCCTAGAG	CTATGTAACCTTGCTACAACCTGTCTAA	610	52	

Region	Fragment	Forward_Primer	Reverse_Primer	Amplicon length	Opt. T'
	<b>C10</b>	TTTGGTTAAGCCTTACTACAGGGACAAA	CTACTACCTGCCCATGTGCCTACCTGTA	585	55
	<b>C11</b>	GTCTGGGGAGGGAAATGGAGTCGATTAT	ACCCAGGTCAGAGGTGCATAACTACAA	433	57
	<b>C12</b>	GGGGTGAAGAATCTTTCCTCATGCAGAT	CCAACGGAAAGATCATTAATCAAGTTCG	826	56
	<b>C13.A</b>	GTTCTATTGTGGCAGCTCCATTGAGACA	GACATTCTTCCCCTCGTTTTTGGTAAA	755	57
	<b>C13.B</b>	TGTAATAATTAAGCTGGGAGGCAGAATC	TCACCTCCAACACTACAGCGTTATGAAGA	651	54
	<b>C14</b>	TGTATTCCCACTGCCTTGTGTACTTTAT	GTAGTCTACAATGCCATCAGAGGTCTGT	596	55
	<b>C15</b>	ACATACATATAGCTACCCCTCTTAATA	AGATGGCTGTTAGATCTTTAACCTTTTC	493	52
	<b>C16</b>	AAATAAAGGGCTCAGTAGTCCTCAAAC	TGAGAGCCATACCTACCAAACCTAGTA	485	56
	<b>C17</b>	CTAGTTATAACCAGTTTTAGTGGGTA	TTTTGTTAAAGGAGAGCCAGATAAATAT	865	52
	<b>C18</b>	CTTTAGTTTGATTGCAAGATAAGAGTTT	CTGGGAAAATATTGTTTAGCTAGTTGAT	893	51
	<b>C19</b>	GTGGGCATTTGTTTTCAAGCTGGTATC	ATTGCCTTACAGATGCCCTGTCTAGCA	895	57
	<b>C20</b>	TTGGTGGAGAATTAAGAAAGCGGATGGA	TTCCCCTCCTGCATTGACTGTGAATAGC	387	57
	<b>C21.A</b>	CTGCAAGTTTATTTAAAGTGGGAGAAGT	AACGGCTATAACAAAGCAATAGAGGTAG	543	53
	<b>C21.B</b>	TCCAACATATTCAGCCAAGCAACAGAAC	ACTGGGATGGGAGGGGATTTTAAGTTG	643	56
	<b>C22</b>	ATGAGTTATCCCTTCGCTGGGGTTC	AAGGCCTATGTCACTCCTTGCCATCCTC	628	58
	<b>C23</b>	TATATATATATCCGAACAACCTGAA	TACCTGGAGCTCAAAGGGTAGTA	826	53
	<b>C24</b>	TGAAATTTTGAGAGCCATATTACTACCC	CACCACTGGCACTATGTGTCATATAAAG	484	52
	<b>C25</b>	AGGCGTCTGCTACATCCTTTGGTCATGG	GCTTAGGGGAGGAGCTGAGGATATGGTT	640	56
	<b>C26</b>	GGCAATATGGTGTTTTAGAGGGAGAACG	TTTCACTTTTGAATCAACAAGCCACCAG	668	56
	<b>C27.A</b>	TGTGCAATACTGTTTCTAAATAAAGTGT	CTGTTAATGTAGGTGCAATAGACATCAA	499	52
	<b>C27.B</b>	TGGGGAAACCTTATCCTCTTTGAAGTAA	CAGCCTTGAAAATTAAGACACAGTAGCA	620	55
	<b>C28</b>	CAGGCGTGAGCTACTGCACCCACTTATA	GCCTTCTCCCCTCAAATCCCATGTACT	573	55
	<b>C29</b>	TAGCAGCCAAGATGACACTGTTAGAAC	TCCTAGGCAAGGAAAATCACCATACA	660	53
	<b>C30</b>	TCAATGGAGTAACAGGTGCGGTATATTA	GTGGGGAAAAGCTCTGAGTAACTTATG	545	54
	<b>C31</b>	TAAAGTGTGCGCTCTGAAACTCTCATT	GTCCCCTTGATTCACATAGTGAGTTTCA	697	54

Region	Fragment	Forward_Primer	Reverse_Primer	Amplicon length	Opt. T'
	<b>C32</b>	ATGTGAATCAAGGGGACAAATGTCTCAG	ACTGGAGCAATGCAGACCGGATCTAAGA	567	57
	<b>C33</b>	GTGGCTGATTGATGGACTTCTTATGATT	CCCTCAGGGAAAGAGGTCTAGTTATCTC	482	54
	<b>C34</b>	TTTACTTATGCCTCTTAAAAATGACAGGA	TTAAAAACAACATCACCTTTTCGACCTTA	478	53
	<b>C35</b>	AGACAGAATTCCTTCCCTCTAAGAGTCT	AACCATTAAACAGACAGGGACAGGTTT	583	52
	<b>C36</b>	GACTTGTCACCTTTTGGAGAGGCTACAA	CTCCGGGCATTAATAAACCTGTAACCAA	848	55
	<b>C37</b>	AGTCCCATTTTGCCAAGATCTAGTTTTA	GAAGTATCGCCATTTTAAGTCATAATT	854	54
<b>Predicted gene</b>	<b>PRD01.A</b>	TGGGATGGGGAGAGGTGACAAGAGTAGC	GCATTCCTAACAAGCTCCCAGGGACTGC	769	60
	<b>PRD01.B</b>	TACCCATGAGGTCACCTACTCAAAATGG	TGTCCAGGTAGATTTCTTTGCAGACAAA	765	57
	<b>PRD01.C</b>	TTTCAGAAACGATTGGATTTTCAGATAG	GTACAATTCTGTTTCATCCGACCACATC	628	56
	<b>PRD02</b>	AGCGCCTGGCACATTCTCACTTCTGAT	TTGGTGAACCCCTGAGTAAGGGCTGTG	497	60
	<b>PRD03</b>	TTTTCTGTGGTTTACCAAGTTGGCTGTG	CCTGGTCTCTTCCCCAAAACTGAATTG	378	55
	<b>PRD04</b>	CAGGGACTTCATATACAAGAACTGATAT	CCTAATTCTATTTTAATGCCACTATTA	889	52
<b>Exon</b>	<b>E01</b>	GACTGATTCAAGGGAAGCGAGCG	CGGCCACGTTCAATTTAATCTCAC	578	57
	<b>E02</b>	GGAAGGACCCAGCTGTATTGACC	CAAGGGCACAGAGAAAGAGGGAAG	398	59
	<b>E03</b>	CTGGGGGAGGGGAGGAAAAGG	GGTGAGTGGGGATGCAGATGGGTG	536	60
	<b>E04</b>	CCTCCCCGAGAGCCGTAGTTGC	GAGGGCGTTGGAGGTGGGAGGTAG	362	59
	<b>E05</b>	GGGGTGGGCTGGAGATGGTAG	GGCGTTTGATCCCAGTTTTAGTG	344	56
	<b>E06</b>	ATTGCCCTGTGTTTCCCCTATT	AAATTAAGCGACAAGAAACAGG	524	53
	<b>E07</b>	GTTGAAGGGGGATGGGAAAGGTG	TGGCCAGGTGACAGACAGTTAAGC	445	57
	<b>E08</b>	GCCTAGGCCTGTCTTTCTCTG	GTTAACAAAATCGCAATCGTGAAT	537	53
	<b>E09</b>	GGTTCTGCAAGTATCCTAAGTAGCT	TGAATCAACAAGCCACCAGG	420	53
	<b>E10</b>	CCAAGTGCATTCATCTTTCCCTC	GCCAGGCGTTCCTATACTACTCC	261	56
	<b>E11</b>	AGCCTGCTATGTTCTGTCTCTTTC	GATCCTGGCTTCCCCTCTG	368	54
	<b>E12</b>	GTCATCCCCTCATCAACACAG	TGGGCAAGGAGAACATAAGAA	423	54
	<b>E13.A</b>	GGAAAATAGTGGCAAAATGTGAGTT	GGTCCAGAGTAGATGCCAAGAATG	584	56

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	<b>E13.B</b>	ATACAGGAGACCCAACAATGAGTG	CCAGGCTAGAAAGAGGGAGAGA	573	54
	<b>E13.C</b>	CTCTCGCCTAGGATTTTCAGCC	TGCCAACTCATACCAAAGTCTAC	602	53
	<b>E13.D</b>	GTCCCATGCAACAACCAC	GGGGGGCAGTAAACAAATTTTTC	544	53
<b>RT-PCR</b>	<b>meis1_1'F</b>	GATTGGCCGAGCACTCCT			
	<b>meis1_1eF</b>	AGGTCCCGTAGACCGAAGAT			
	<b>meis1_3R</b>		ATGACTCTGACGAGCAGACG		
	<b>meis1_3'F</b>	TCCAACCTCAGATTTTCTCTCTG			
	<b>meis1_6R</b>		GGCATTTCCTTTCAAACA		
	<b>meis1_7F</b>	ACGGCATCTACTCGTTCAGG			
	<b>meis1_10F</b>	AGTGCAGCCCATGATAGACC			
	<b>meis1_10R</b>		GGTCTATCATGGGCTGCACT		
	<b>meis1_12R</b>		TCATGCCCATTCCTCATA		
	<b>meis1_13R</b>		TTGATGCTGACATTGGCATT		

**Supplementary Table S2. Summary of Sequence Variants Detected in Sequencing of *MEIS1* Exons and Flanking Regions**

Variant	Classification	Position	Genomic region	ID	Variant	Allelewise		Genotypewise			
						MAF case/control	<i>P</i> value	Genotype frequency* case/control		<i>P</i> value	
<b>v1</b>	intronic	66520690	intron 3	rs2271856	C/T	0.384/0.356	0.357	0.155/0.140	0.457/0.432	0.388/0.428	0.637
<b>v2</b>	intronic	66522040	intron 4	rs41285949	G/A	0.000/0.002	0.265	0.000/0.004	1.000/0.997	0.000/0.000	0.265
<b>v3</b>	intronic, ins/del	66522148	intron 5	novel ins/del	(CA)del	0.000/0.002	0.265	0.000/0.004	1.000/0.997	0.000/0.000	0.265
<b>v4</b>	intronic	66522155	intron 5	novel SNP	A/G	0.002/0.002	0.915	0.004/0.004	0.997/0.997	0.000/0.000	0.915
<b>v5</b>	intronic	66522155	intron 5	novel SNP	C/G	0.002/0.000	0.214	0.004/0.000	0.997/1.000	0.000/0.000	0.214
<b>v6</b>	exonic, syn	66544852	Exon 7	rs13005707	A/G	0.010/0.011	0.954	0.020/0.021	0.980/0.980	0.000/0.000	0.954
<b>v7</b>	intronic	66649410	intron 11	novel SNP	C/T	0.053/0.042	0.402	0.008/0.000	0.090/0.084	0.902/0.916	0.206
<b>v8</b>	3'UTR	66651786	Exon 13	rs2861108	T/G	0.025/0.016	0.360	0.053/0.033	0.947/0.968	0.000/0.000	0.355
<b>v9</b>	3'UTR	66652131	Exon 13	novel SNP	C/T	0.002/0.000	0.238	0.004/0.000	0.996/1.000	0.000/0.000	0.237
<b>v10</b>	3'UTR	66652293	Exon 13	novel SNP	G/C	0.000/0.004	0.114	0.000/0.007	1.000/0.993	0.000/0.000	0.114
<b>v11</b>	3'UTR	66652436	Exon 13	novel SNP	T/A	0.002/0.004	0.645	0.004/0.007	0.996/0.993	0.000/0.000	0.645
<b>v12</b>	3'UTR	66652634	Exon 13	novel SNP	A/G	0.000/0.004	0.114	0.000/0.007	1.000/0.993	0.000/0.000	0.114
<b>v13</b>	3'UTR	66652636	Exon 13	novel SNP	A/G	0.000/0.002	0.264	0.000/0.004	1.000/0.997	0.000/0.000	0.264
<b>v14</b>	3'UTR	66652708	Exon 13	novel SNP	G/T	0.002/0.000	0.219	0.004/0.000	0.996/1.000	0.000/0.000	0.219
<b>v15</b>	3'UTR	66652752	Exon 13	novel SNP	A/G	0.000/0.002	0.260	0.000/0.004	1.000/0.996	0.000/0.000	0.260
<b>v16</b>	3'UTR	66652898	Exon 13	novel SNP	G/A	0.033/0.038	0.656	0.000/0.004	0.066/0.076	0.934/0.921	0.650
<b>v17</b>	3'UTR	66653295	Exon 13	novel SNP	A/G	0.002/0.000	0.217	0.004/0.000	0.996/1.000	0.000/0.000	0.217
<b>v18</b>	3'UTR	66653332	Exon 13	novel SNP	G/T	0.000/0.002	0.262	0.000/0.004	1.000/0.996	0.000/0.000	0.262
<b>v19</b>	3'UTR	66653340	Exon 13	novel SNP	A/G	0.000/0.004	0.113	0.000/0.007	1.000/0.993	0.000/0.000	0.112

\* genotype frequency in the order of homozygous minor allele / heterozygous /homozygous major allele; MAF: minor allele frequency

Supplementary Table S3. Marker Information and Case-Control Association Tests

dbSNP	Location	Region	MAF case	MAF control	Allelic association*	Genotypic association*
rs6546232	66,512,364	Intergenic	0.350	0.337	0.65	0.90
rs13033745	66,513,907	Intergenic	0.347	0.336	0.70	0.93
rs11883967	66,527,366	Intron6	0.376	0.362	0.62	0.70
rs6716792	66,541,347	Intron6	0.452	0.475	0.43	0.40
rs11692504	66,577,567	Intron7	0.391	0.385	0.84	0.87
rs4547518	66,580,072	Intron7	0.260	0.340	$3.02 \times 10^{-03}$	$8.59 \times 10^{-03}$
rs12373638	66,584,746	Intron7	0.491	0.447	0.13	0.34
rs6721499	66,597,095	Intron8	0.297	0.375	$4.76 \times 10^{-03}$	<b><math>5.52 \times 10^{-04}</math></b>
rs4300815	66,600,036	Intron8	0.250	0.331	$2.47 \times 10^{-03}$	$4.52 \times 10^{-03}$
rs4544423	66,603,521	Intron8	0.318	0.415	<b><math>6.64 \times 10^{-04}</math></b>	<b><math>7.56 \times 10^{-04}</math></b>
rs6742861	66,603,656	Intron8	0.235	0.285	0.07	0.22
rs6728018	66,603,769	Intron8	0.237	0.287	0.07	0.23
C13B_2	66,604,068	Intron8	0.190	0.081	<b><math>1.81 \times 10^{-07}</math></b>	<b><math>2.21 \times 10^{-6}</math></b>
rs11688599	66,610,054	Intron8	0.251	0.329	$3.88 \times 10^{-03}$	$8.84 \times 10^{-03}$
rs4316931	66,614,409	Intron8	0.251	0.282	0.23	0.40
rs7603236	66,615,915	Intron8	0.314	0.404	$1.63 \times 10^{-03}$	$2.08 \times 10^{-03}$
rs12469063	66,617,812	Intron8	0.364	0.222	<b><math>8.12 \times 10^{-08}</math></b>	<b><math>1.19 \times 10^{-07}</math></b>
rs11681729	66,618,204	Intron8	0.241	0.209	0.23	0.21
rs9789535	66,619,082	Intron8	0.396	0.445	0.09	0.11
rs17625724	66,622,479	Intron8	0.237	0.290	0.04	0.15
rs17625742	66,622,658	Intron8	0.232	0.294	0.02	0.08
rs11678796	66,623,539	Intron8	0.253	0.335	$2.43 \times 10^{-03}$	$5.68 \times 10^{-03}$
rs2300478	66,634,957	Intron9	0.373	0.243	<b><math>1.37 \times 10^{-06}</math></b>	<b><math>9.41 \times 10^{-06}</math></b>
rs2300483	66,641,765	Intron9	0.530	0.432	<b><math>7.73 \times 10^{-04}</math></b>	$3.39 \times 10^{-03}$
rs2300486	66,644,908	Intron9	0.292	0.287	0.84	0.72
rs1000756	66,648,338	Intron10	0.305	0.357	0.06	0.14

MAF: minor allele frequency. \* Significant results are highlighted in bold.

**Supplementary Table S4. Clinical Information on the Autopsy Brain Tissues from 28 Individuals with RLS**

Brain #	Age	Sex	PMI	RLS diagnosis confirmation	Family history	RLS age at onset (years)	RLS progression (years symptoms progressed to daily)	RLS Treatment
B4958	77	F	11	RLS	no	6	yes (7)	vicodin
B5041	76	F	19	RLS	yes	10	yes	zanaflex
B5065	84	F	17	RLS	yes	42	yes (20-40)	bromocriptine, propoxyphene, iron
B5164	106	F	3	RLS	yes	101	yes (1)	
B5347	53	F	4	RLS	no	<45	yes (3)	carbidopa/levodopa, neurontin
B5462	77	F	20	RLS	yes	43	yes (15)	carbidopa/levodopa, mirapex, iron, codiene
B5609	86	F	17	RLS	no	45-50	yes (20)	carbidopa/levodopa, permax, percocet, ativan
B5655	85	F	15	RLS	unknown	20	yes	codeine.
B5739	52	M	8	RLS	unknown	unknown	unknown	
B5909	71	F	19	RLS	unknown	unknown	unknown	
B6032	89	F	5	RLS	unknown	unknown	unknown	
B6050	76	F	20	RLS	unknown	unknown	unknown	
B6073	85	M	24	RLS	unknown	unknown	unknown	
B6084	82	F	19	RLS	unknown	28	yes (15)	carbidopa/levodopa, xanax, effexor, clariton D
B6106	75	F	22	RLS	unknown	unknown	unknown	
B6264	83	F	27	RLS	yes	5	yes (15+)	demerol, codeine, klonopin, durgesic patch, neurtonin
B6343	87	F	22	RLS	yes	childhood	yes	permax
B6402	87	M	22	RLS	unknown	74	unknown	mirapex
B6410	81	M	14	RLS	unknown	70	no	oxycodone, gabapentin, clonazepam, iron
B6441	83	F	17	RLS	no	35	yes (45)	carbidopa/levodopa, zanex
B6490	73	F	22	RLS	no	35	yes (23)	permax, clonazepam
B6504	90	M	16	RLS	yes	20	yes (15)	carbidopa/levodopa, permax
B6607	87	M	12	RLS	unknown	55	yes	carbidopa/levodopa, mirapex, iron
B6619	77	M	14	RLS	yes	50	no	
B6644	91	F	9	RLS	yes	40	yes	carbidopa/levodopa, vit E
B6719	78	F	13	RLS	yes	childhood	yes	ultram, ambien, vicodin, hydrocodone, neurontin
B6772	88	F	15	RLS	unknown	35	yes	iron, clonazepam
B6945	93	F	20	RLS	yes	14	yes	valium, carbidopa/levodopa

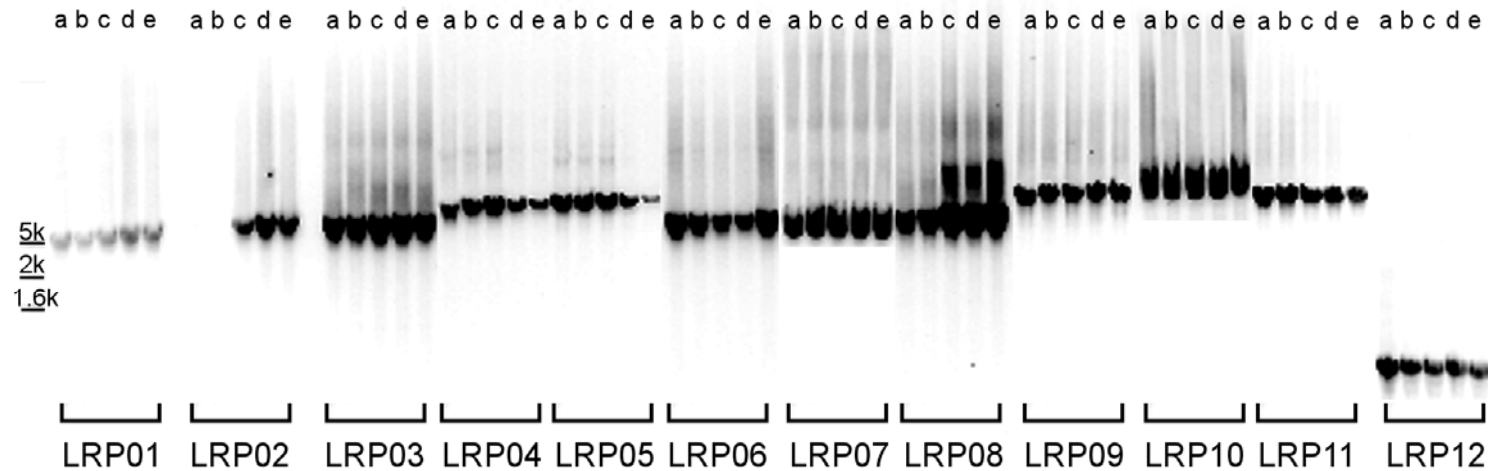
PMI: post-mortem interval.



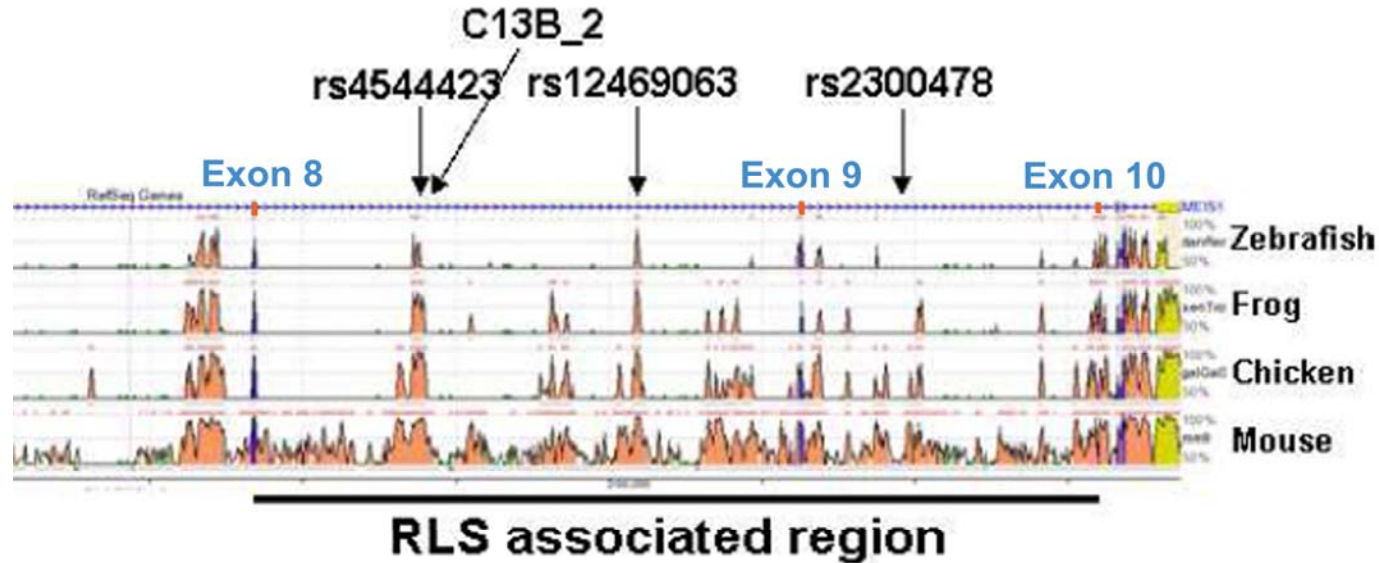
**Supplementary Table S5. Clinical and Demographic Information on RLS Patients and Controls**

<b>RLS Cases</b>	<b>285</b>
Female:male ratio	1.7 (178/107)
Age (years, mean $\pm$ SD)	54 $\pm$ 12
Age of onset (yr, mean $\pm$ SD)	30 $\pm$ 16
Severity score	26 $\pm$ 11 (IRLSS <sup>a</sup> )
Daily or nearly daily symptoms	82%
Familial cases	78%
Ferritin ( $\mu$ g/l)	Female: 72 $\pm$ 51, Male:143 $\pm$ 101
PLMS positive <sup>b</sup>	82%
<b>Controls</b>	<b>285</b>
Female:male	1.3 (162/123)
Age (yr, mean $\pm$ SD)	42 $\pm$ 13

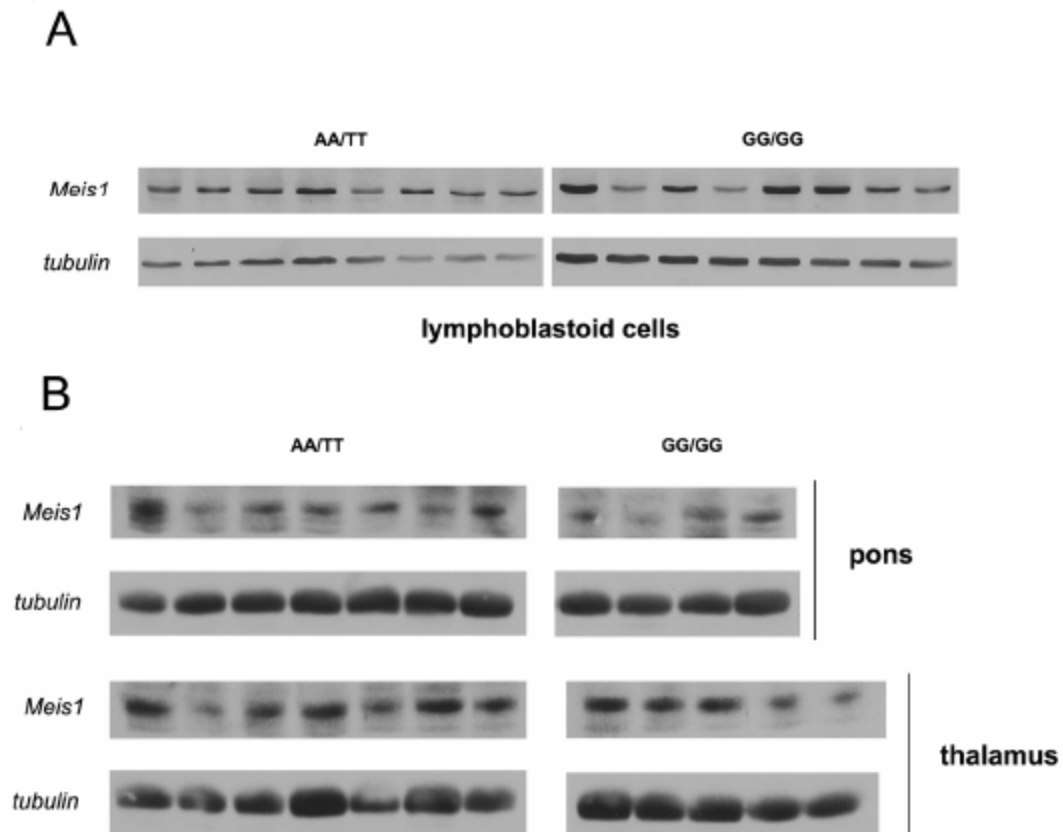
<sup>a</sup> The International RLS Study Group severity scale (SS) contains 10 items with a range of 0 – 4 score/each. SS: 0 – 10, mild; 11 – 20, moderate; 21 – 30, severe; 31 – 40, very severe; <sup>b</sup> The PLMS measurements were performed by one-night polysomnography (PSG) prior to treatment or with reduced dopaminergic medications. The PLMS index > 5.0/hr was classified as positive according to the standard.



**Supplementary Figure S1: Long range PCR results.** The PCR products were electrophoresed on a 0.9% agarose gel (UltraPure, Invitrogen) for 4 hours. Lambda DNA/Hind III Fragment (Invitrogen) was used as the molecular weight standard. Lane a,b,d were samples from RLS patients homozygous for the risk associated haplotype (GG/GG); lane c,e were samples from two non-RLS controls homozygous for haplotype (AA/TT). Fragments were run on three different gels based on different size range and images were assembled according to the ladders. LRP02 was repeated and two failed samples (lane a, b) were reamplified and showed no difference.



**Supplementary Figure S2. A Schematic Showing of Comparative Genomic Analysis of the MEIS1 Gene Using the ECR Browser\*.** Pairwise comparisons of the human sequence with mouse, chicken, frog, and zebrafish are shown from the bottom to the top respectively. Blue peaks represent significantly evolutionary conserved exons, and peach colored peaks represent significantly evolutionary conserved intronic noncoding sequences. Black arrows point to the location of the RLS associated SNPs.  
 \* Ref: Loots, G. and Ovcharenko, I. (2007) ECRbase: database of evolutionary conserved regions, promoters, and transcription factor binding sites in vertebrate genomes. *Bioinformatics*, 23, 122-124. Epub 2006 Nov 7.



**Supplementary Figure S3: Western blot of MEIS1 in RLS lymphoblastoid cell lines and brain tissues.** (A) Western blotting experiments illustrate significant changes in expression levels of MEIS1 in lymphoblastoid cell samples carrying the AA/TT genotype, compared with samples carrying the RLS-associated genotype GG/GG. (B) Western blotting experiments illustrate changes in expression levels of MEIS1 in both thalamus and pons tissue samples of RLS patients carrying the AA/TT genotype, compared with RLS patients carrying the RLS-related genotype GG/GG.

**Method:**

**Protein extraction:** Protein lysates from human thalamus, pons, and lymphoblastoid cells were homogenized in SUB lysis buffer (0.5% SDS, 8M Urea, 2%  $\beta$ -mercaptoethanol), centrifuged 20 min/13,000 rpm at 4 °C. The soluble fractions were quantified and diluted in loading buffer. Protein lysates were prepared three separate times.