## **Supporting Information**

Guo et al. 10.1073/pnas.0906606106

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Fig. S1. AICAR treatment does not promote Akt Ser-473 or Akt Thr-308 phosphorylation. U87 or U87-EGFR cells, were treated with AICAR (0.5 mM) for up to 24 h, then EGF (10 ng/mL) for 15 min, and the effect on signaling proteins was determined by immunoblot analysis with indicated antibodies.



**Fig. 52.** Activation of AMPK through an alternative mechanism inhibits glioblastoma growth with preferential inhibition in EGFR-activated tumor cells. 2, 4-dinitrophenol (DNP) activates AMPK by lowering ATP via inhibition of oxidative phosphorylation, thus increasing the AMP/ATP ratio. (*A*) Western blot analysis of effect of DNP on AMPK and ACC. U87-EGFRVIII cells were plated and 24 h later treated with DNP (0.5 mM) for up to 8 h. AMPK activation was determined by probing cellular lysates with antibodies against AMPK Thr-172 and ACC Ser-79 phosphorylation. (*B*) U87-EGFRVIII cells were plated and 24 h later, treated with a range of concentrations of DNP in 1% FBS conditions for 3 days. Relative cell number was assessed using the WST assay (Chemicon). (C) DNP is significantly more effective at blocking the growth of EGFRVIII reGFR expressing glioblastoma cells. U87, U87/EGFR, and U87/EGFRVIII cell lines were treated by the WST assay (Chemicon). Control cells were treated by ethanol, the diluent, alone (1:1,000).



**Fig. S3.** Effect of AICAR and rapamycin treatment on U87 glioblastoma cell colony growth. U87 glioblastoma cells were seeded at a density of 500 cells per 60-mm dish, 24 h later, medium was changed and AICAR (0.5 mM) and/or rapamycin (1 nM) was added. The medium, including AICAR or rapamycin, was replenished every 3 days. Three weeks after initial treatment, cultures were fixed with 4% formaldehyde (Sigma) in PBS and stained with a 0.5% crystal violet solution (Sigma) in 25% methanol. Colonies of more than 50 cells were counted. Control cells were treated with the diluent, ethanol (1:1,000).

AC PNAS







**Fig. S5.** Effect of inhibition of HMG-CoA reductase and ACC on glioblastoma cell colony growth. U87 and U87-EGFRvIII glioblastoma cells were seeded at a density of 500 cells per 60-mm dish, and 24 h later, medium was changed and atorvastatin (Ator 1  $\mu$ M) and/or TOFA (10  $\mu$ g/mL) was added. The medium, including atorvastatin (Ator) and TOFA, was replenished every 3 days. Three weeks after initial treatment, cultures were fixed with 4% formaldehyde (Sigma) in PBS and stained with a 0.5% crystal violet solution (Sigma) in 25% methanol. Colonies of more than 50 cells were counted. The experiment was repeated 3 times. Colony number was normalized with control group, into which DMSO alone was added, *P* value was determined using student *t* Test. (*A* and *B*) Evidence for enhanced efficacy by combining atorvastatin and TOFA in blocking clonogenic growth of U87-EGFRVIII cells (*B*), \*, *P* < 0.05 compared with TOFA treatment. No such combination benefit is detected in non-EGFRVIII expressing U87 cells (*A*). Control cells treated by DMSO (1:1,000).



**Fig. S6.** Addition of mevalonate (Mev) and palmitate (Pal) rescued cell growth inhibited by atorvastatin (1  $\mu$ M) and TOFA (10  $\mu$ g/mL). U87-EGFRvIII cells were plated, 24 h later they were treated with either atorvastatin and TOFA at indicated doses, mevalonate and palmitate (100  $\mu$ M) or atorvastatin (Ator) and TOFA plus mevalonate and palmitate. Cells were treated for 3 days and relative cell growth was determined by the WST assay. \*, *P* < 0.0001 compared with control cells treated DMSO (1:1,000).



Fig. S7. AICAR treatment did not induce apoptotic cell death in vivo. TUNEL immunohistochemical staining was performed on AICAR-treated or vehicle (PBS)-treated xenografts. No increase in TUNEL positive cells was detected after treatment with AICAR (quantitation not shown as no difference was seen). EGFRVIII expressing tumors have lower baseline apoptosis.

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