Cell, Volume 137 Supplemental Data Loss of GABAergic Signaling by AgRP Neurons to the Parabrachial Nucleus Leads to Starvation

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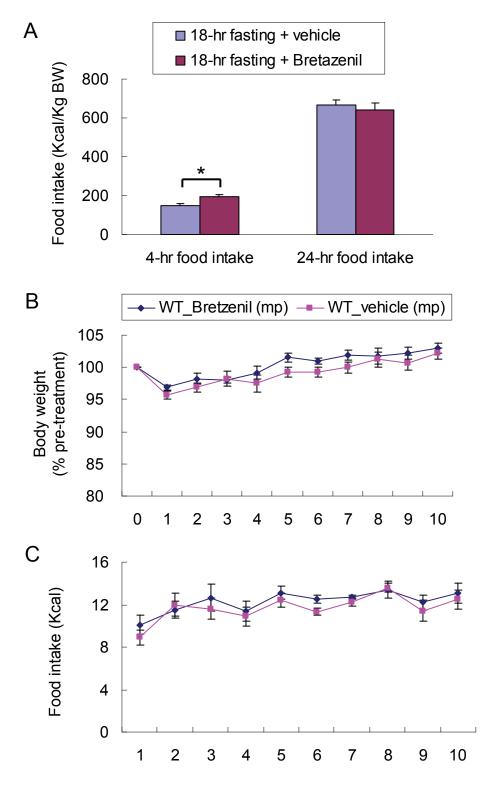


Figure S1. Short- and long-term effects of bretazenil on the regulation of food intake and body weight.

(A) Cumulative food intake of fasting wild-type mice after treatment of bretazenil (0.2 mg/kg, ip) or vehicle.

(**B**) Percentage of initial body weight of wild-type mice which are implanted mini pumps filled with either bretazenil (2.5 mg/ml) or vehicle.

(C) Intake of liquid diet by the mice described in B.

N = 6 for each group. *, p < 0.05, ANOVA. Error bars represent the SEM.

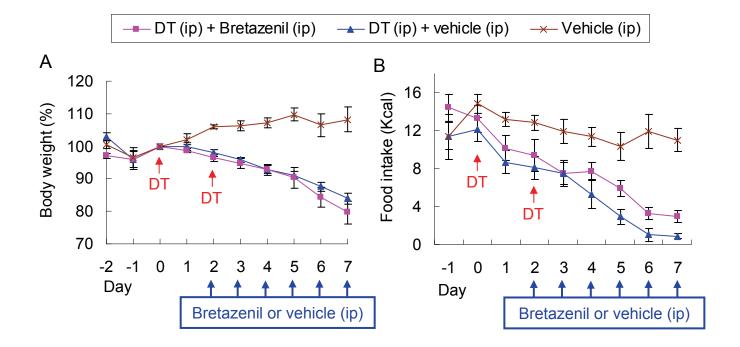


Figure S2. Effects of daily intraperitoneal injection of bretazenil on body weight and food intake. (*A*) Percentage of initial body weight of $Agrp^{DTR/+}$ mice after two intramuscular injections of DT or vehicle, then followed by i.p. injection of either bretazenil (0.2 mg/kg) or vehicle for 6 consecutive days. (*B*) Intake of liquid diet by the mice described in *A*.

N = 6 - 8 for each group. Error bars represent the SEM.

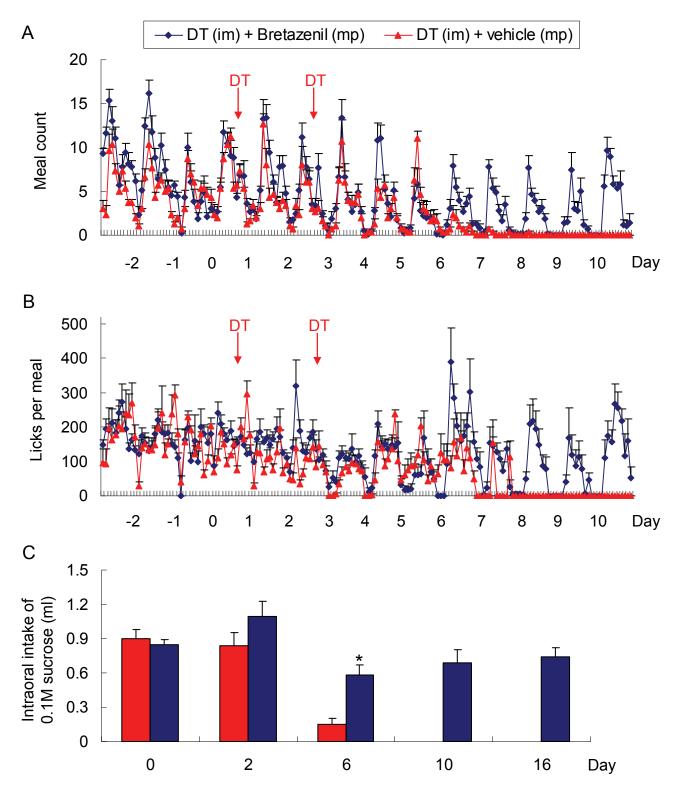


Figure S3. Appetitive and consummatory responses in AgRP-ablated mice are rescued by chronic administration of bretazenil

(*A*, *B*) Total number of meals (*A*) and average meal size (as shown by licks per meal, *B*) were measured in DT-treated, $Agrp^{DTR/+}$ mice after subcutaneous implantation of mini-osmotic pumps loaded with either bretazenil (n = 12) or vehicle (n = 10). Meals are defined as the number of consecutive 10-s bins that contain five or more licks, and each meal must be separated by 12 or more 10-s bins in which no significant licking activity was recorded.

(C) Consumption of 0.1 M sucrose solution infused through cheek fistulas was measured in the mice described in A, B. The control animals without bretazenil (gray bars) did not survive beyond day 6. *, p < 0.01, ANOVA. Error bars represent the SEM.

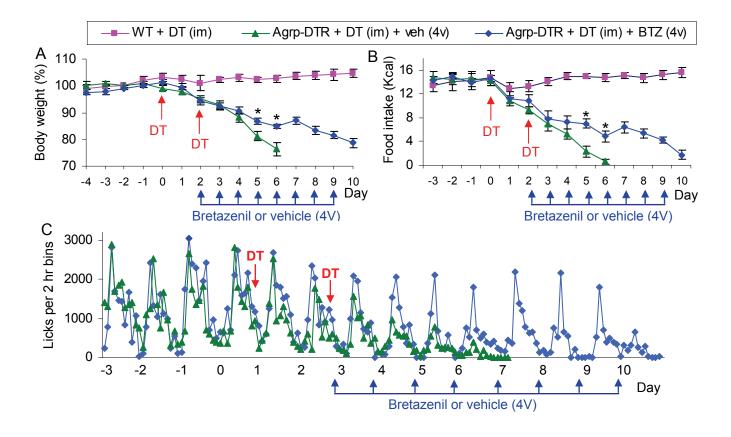
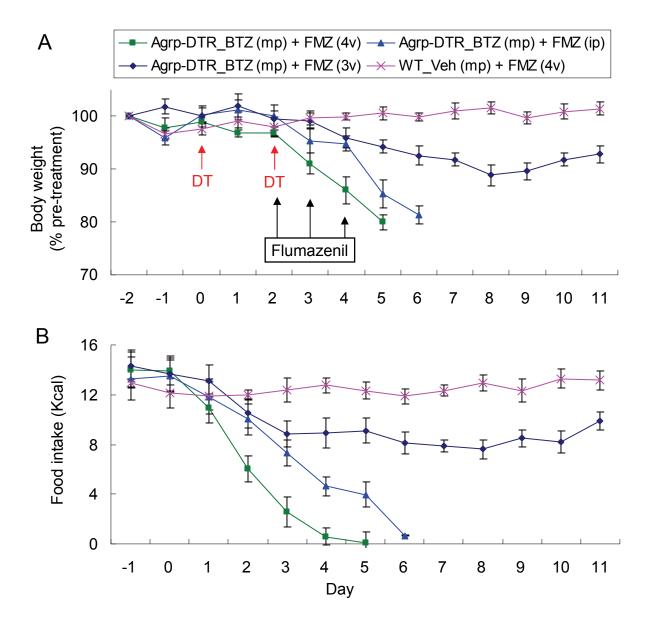


Figure S4. 4th ventricle administration of bretazenil alleviates starvation in adult mice after ablation of AgRP neurons.

(**A**) Percentage of initial body weight of wild-type mice and $Agrp^{DTR/+}$ mice after intramuscular injections of DT, then followed by daily 4v injection of either bretazenil (1 µg/mouse/day) or vehicle.

(B) Liquid diet intake of the mice described in A.

(C) Licking activity of two groups of $Agrp^{DTR/+}$ mice described in **A**. The average number of licks in 2-hr bins is plotted in both groups. *, p < 0.01, ANOVA.





(A) Percentage of initial body weight of DT-treated, $Agrp^{DTR/+}$ and wide-type mice after implantation of minipumps loaded with bretazenil or vehicle, followed by administration of flumazenil either intraperitoneally or into the 3rd or 4th ventricle during Day 2 – Day 4.

(B) Intake of liquid diet by the mice described in A.

N = 6 - 10 per group. Error bars represent the SEM.

	Pair-fed		DT + vehicle (mp)		DT + Bretazenil (mp)	
ARC	A1	A2	A3	A4	A5	A6
PVN	B1	B2	B3	B4	B5	B6
DMH	C1	C2	C3	C4	C5	C6
MPO	D1	D2 .	D3	D4	D5	D6
ΓS	E1	E2	E3	E4:	E5	E6
PAG	F1	F2	F3.	F4	F5	F6 >
PBN	G1	G2	G3	G4	G5	G6
NTS	H1	H2	H3	H4	H5	H6

Figure S6. Chronic administration of bretazenil downregulates *Fos* activation in selected brain regions of mice in which AgRP neurons have been ablated.

(*A1-A6*) Representative pictures of *Fos* in situ hybridization in the ARC of pair-fed $Agrp^{DTR/+}$ mice (*A1, A2*) and DT-treated $Agrp^{DTR/+}$ mice that receive subcutaneous infusion of either vehicle (*A3, A4*) or bretazenil (*A5, A6*) via mini pumps.

(*B1-B6, C1-C6, D1-D6, E1-E6, F1-F6, G1-G6, H1-H6*) Representative pictures of *Fos* in situ hybridization in the PVN, DMH, MPO, LS, PAG, PBN, and NTS, of the mice described in *A1-A6*, respectively.

Scale bar (in *A1*) for *A-F*, 400 μ m; scale bar (in *G1*) for *G-H*, 400 μ m; n = 4-6 for each group. See **Figure 3** of the main text for quantified results.

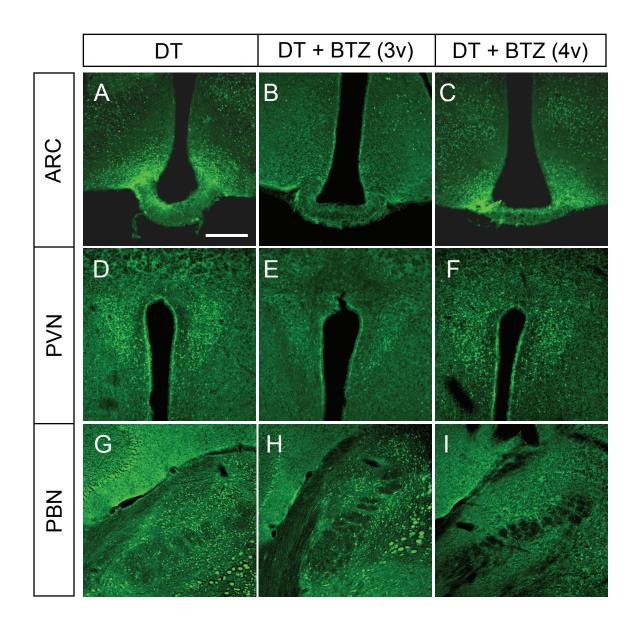


Figure S7. Differentiated effects of chronic icv infusion of bretazenil on Fos activation in AgRP neuron-ablated mice.

(A-C) Anti-Fos immunostaining in the ARC of DT-treated $Agrp^{DTR/+}$ mice, in some that also receive either 3rd or 4th ventricle administration of bretazenil via mini pumps.

(*D-I*) Fos immunostaining in the PVN, and PBN, of the mice described in *A-C*, respectively. Scale bar (in *A*) for *A-I*, 400 μ m; n = 6 for each group.

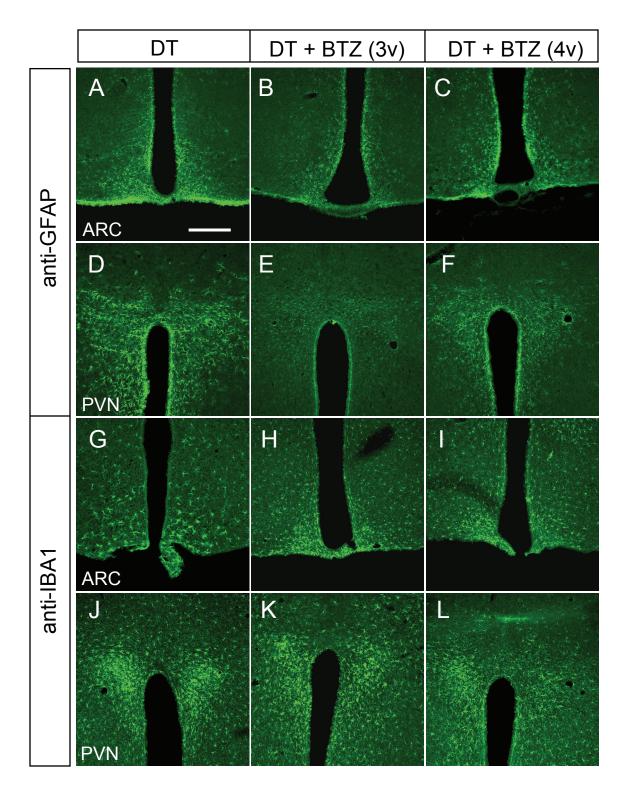


Figure S8. Differentiated effects of chronic icv infusion of bretazenil on gliosis in AgRP neuron-ablated mice.

(*A-C*) Anti-GFAP immunostaining in the ARC of DT-treated $Agrp^{DTR/+}$ mice, in some that also receive either 3rd or 4th ventricle administration of bretazenil via mini pumps.

 $(\textit{\textbf{D-F}})$ GFAP immunostaining in the PVN of the mice described in $\textit{\textbf{A-C}}.$

(G-I) Anti-IBA1 immunostaining in the ARC of the mice described in A-C.

(J-L) Anti-IBA1 immunostaining in the PVN of the mice described in A-C.

Scale bar (in *A*) for *A*-*L*, 400 μ m; n = 6 for each group.

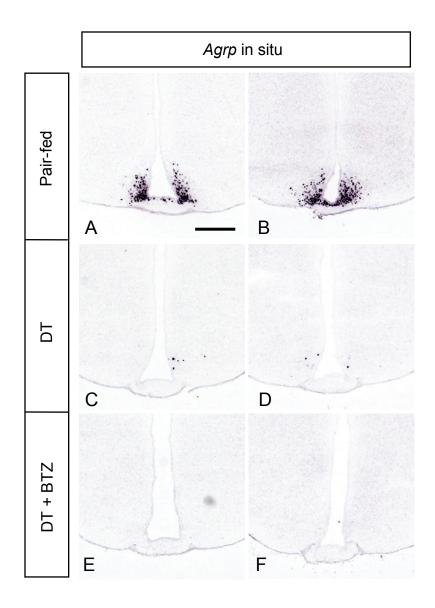


Figure S9. Loss of *Agrp* mRNA following DT treatment

(A-F) Representative pictures of Agrp in situ hybridization in the ARC of $Agrp^{DTR/+}$ mice after either pair feeding (A, B), DT treatment (C, D), or DT treatment plus chronic administration of bretazenil (*E*, *F*). Scale bar (in *A*) for *A*-*F*, 400 μ m; n = 4 - 6 for each group.

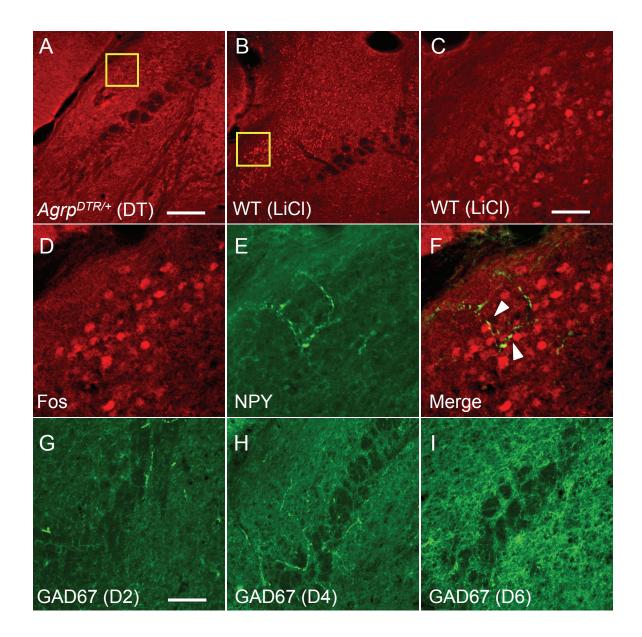


Figure S10. AgRP neurons innervate Fos-positive cells in the PBN (*A*) Fos immunostaining in the PBN of *Agrp*^{DTR/+} mice 4 days after injecting DT.

(B) Fos immunostaining in the PBN of wild-type mice 24 h after injecting LiCl (0.12 g/kg body weight, ip).

(C) Magnified view of the boxed area in B.

(D) Magnified view of the boxed area in A.

(*E*) NPY immunostaining in the same region as *D*.

(F) Merged view of D and E. Arrowheads indicate that varicosities stained with NPY antibody are in close proximity to the Fos-positive neurons in the dorsal lateral region of the PBN.

(G-I) GAD67 staining in the PBN at 2, 4 and 6 days after DT treatment, respectively.

Scale bar in (A) for A, B, 300 µm; in (C) for C-F, 50 µm; in (G) for G-I, 150 µm.