Supporting Information

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Fig. S1. Cartoon of optical tweezers assay tailored for bacteriophage λ .



Fig. S2. ATP concentration (μ M) vs. velocity of translocation (bp/s) in WT and Y46F. Measurements were taken with an optical tweezers assay using a force feedback \approx 5 pN as described in the text. In red is the fit to the Hill equation with a constraint on V_{max} described in the text. Errors are SE.



Fig. S3. Linear-linear plot version of Fig. S2: ATP concentration (μ M) vs. velocity of translocation (bp/s) in WT and Y46F. In red is the fit to the Hill equation described in the text with a constraint on V_{max} . Discarding the 2- μ M data point for the WT ATP concentration vs. velocity plot gives an $n = 0.91 \pm 0.11$ value when fit to the Hill equation.



Fig. S4. Percentage of delayed initiation in packaging events ($t_{start} > 0 s$) in various terminases measured with an optical tweezers assay. Packaging events were initiated with an \approx 5-pN force clamp.



Fig. S5. The crystal structure of T4's gp17 with ATP modeled to be inserted in the catalytic pocket. The adenine-binding or Q motif is zoomed in. The analogous residues to λ gpA's Y46 and Q47 in T4 (Y142, Q143) are shown in cyan (courtesy of K. Kondabagil and V. Rao).



Fig. S6. Histograms of stalling dependence on filling in various terminases: The final lengths (converted to filling percentages) at which the translocating complexes stalled were measured and compiled in bins of 5% filling. Packaging events are normalized to the bin with the largest number of events.

Table S1. Velocities for DNA translocation	on of the λ packaging moto	r measured at various ATP con	centrations used for Figs. S2 and S3
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ATP concentration, μ M	WT velocity, bp/s	SE	Y46F velocity, bp/s	SE
2	128	37	_	
10	141	26	113.3	9.56
30	289	72		_
50	366	57	254.7	39.8
100	370	61	354.6	23.7
500	557	151		_
1,000	582	46.2	402	22