## **Supporting Information**

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**Fig. S1.** Western blot analysis of Xenopus oocytes isolated plasma membrane proteins. (*A*) Immunodetection of both RD (right lane) and RD $\Delta$ KESYY (left lane, indicated RD $\Delta$ K) by an  $\alpha$ 1 antibody. (*B*) Immunodetection with an antibody against the C terminus (anti-KETYY; gift from Jim Kyte, University of California, San Diego) which recognizes the Xenopus  $\alpha$ 1 C-terminal sequence KESYY (right lane) and confirms the successful truncation of the mutant (left lane). Enriched plasma membrane fractions were obtained from  $\approx$ 40–50 oocytes according to the method of Hill et al. (1). Twenty-five mg of plasma membrane protein were separated via 10% SDS PAGE (2) and then electrotransferred to PVDF membrane in buffer containing 10 mM CAPS (pH = 11.0), 10% methanol, at constant current (180 mA) for 2 h (3). After blocking with 10% dry milk in PBS, the membrane was probed with anti-NKA antibody MA1–16731 (Affinity Bioreagents) (1:2,500), followed by incubation with HRP-conjugated goat anti-mouse 2° antibody. Bands were detected via supersignal substrate kit (Pierce, Rockford, IL) according to the manufacturer's protocol.

- 1. Hill WG, et al. (2005) Isolation and characterization of the Xenopus oocyte plasma membrane: A new method for studying activity of water and solute transporters. Am J Physiol Renal Physiol 289:F217–F224.
- 2. Laemmli UK (1970) Cleavage of structural proteins during the assembly of the head of bacteriophage T4. Nature 227:680-685.
- 3. Matsudaira P (1987) Sequence from picomole quantities of proteins electroblotted onto polyvinylidene difluoride membranes. J Biol Chem 262:10035–10038, 1987.



**Fig. S2.** Voltage dependence of Na/K pump related currents in NMG<sub>0</sub><sup>+</sup>. Maximal K<sub>0</sub><sup>+</sup>-induced current ( $I_{max}$ ) from Hill fits (at zero Na<sub>0</sub><sup>+</sup>) are shown for RD (filled black squares, n = 15) and for RD $\Delta$ KESYY (open red circles, n = 17) pumps. Ouabain-sensitive currents (at zero K<sub>0</sub><sup>+</sup>) are plotted as a function of voltage for the RD pumps (filled black diamonds, n = 9) and for the RD $\Delta$ KESYY (open red triangles, n = 10). Note the presence of an inward current in RD pumps that disappears in RD $\Delta$ KESYY pumps.



**Fig. S3.**  $[Na_{0}^{+}]$ -dependence of inward steady state Na/K pump current at zero  $K_{0}^{+}$ . (A) Continuous TEVC recording illustrating the maneuvers performed to measure the  $[Na_{0}^{+}]$  dependence of the inward leak through the Na/K pump at  $V_{h} = -50$  mV. Sharp vertical deflections represent pulses from -200 mV to +40 mV applied to measure the current's voltage dependence. (B)  $[Na_{0}^{+}]$ -dependence of the ouabain-sensitive steady state current measured at -160 mV normalized to the current observed in 125 mM Na<sub>0</sub><sup>+</sup>. The line represents a linear fit to the average data from 5 oocytes.



**Fig. S4.** Simplified kinetic scheme of Na/K pump function. Greek letters  $\alpha$  and  $\beta$  indicate reaction rates in the forward and reverse directions, respectively, for each step in the scheme. For additional details, see *SI Appendix*.

## **Other Supporting Information Files**

SI Appendix (PDF)

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