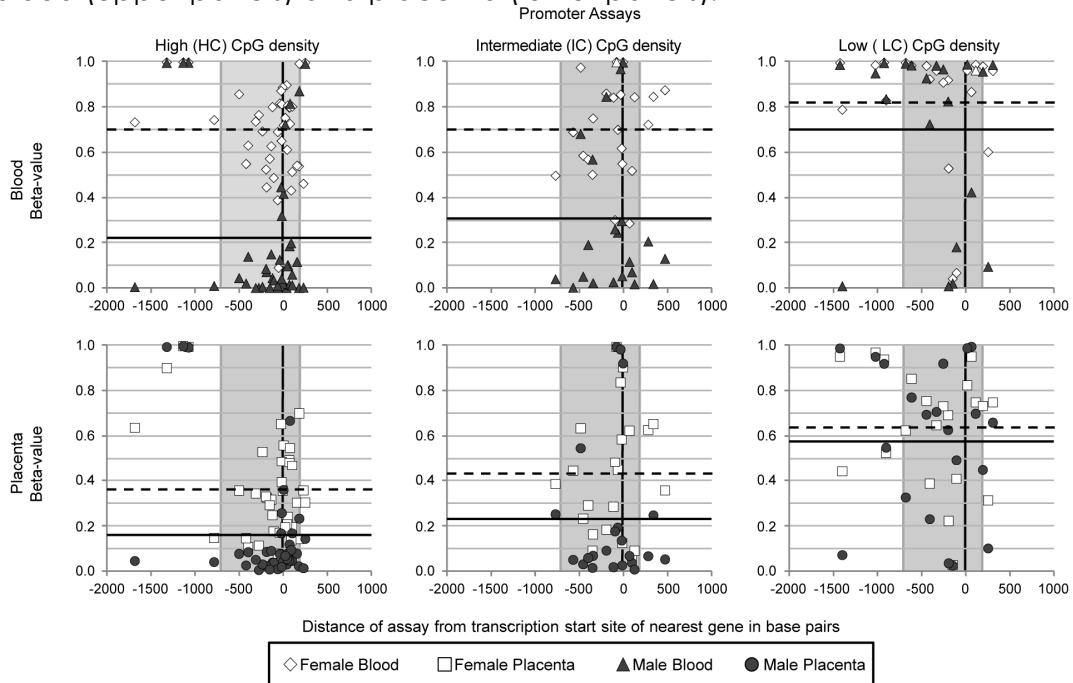


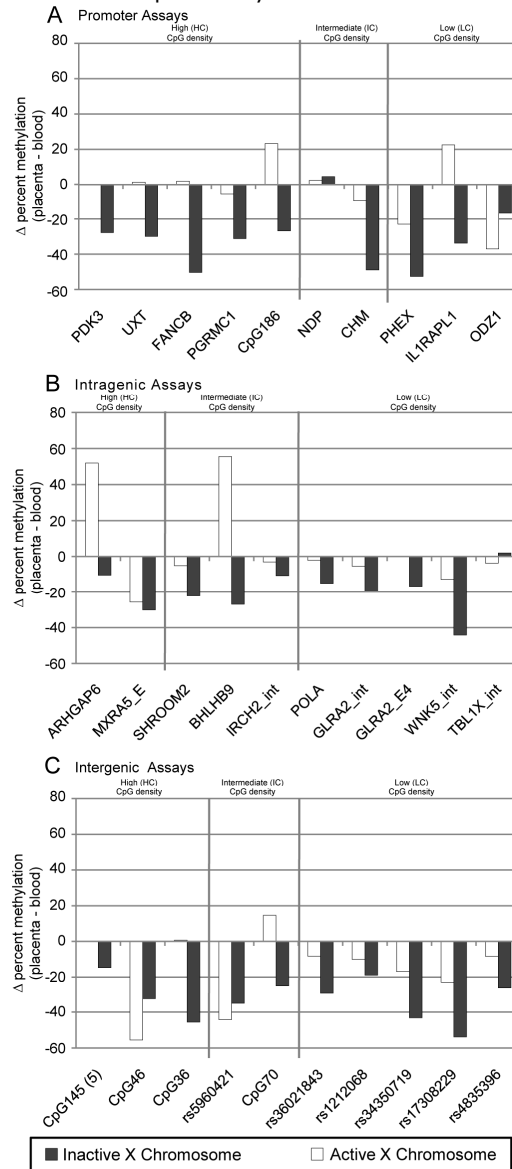
Supplementary Figure 1: Effect of distance from transcription start site on methylation. Beta-values for female blood (white diamond), female placenta (white square), male blood (black triangle) and male placenta (black circle) versus distance from the TSS for each X-linked assay present on Illumina GoldenGate panel. The average beta-value for each sex and tissue is shown as a dashed horizontal line for females and a solid horizontal line for males. Black vertical line marks the TSS (0 bp) and the grey area contains the promoter region as defined by Weber *et al.* (700 bp upstream to 200 bp downstream of the TSS) (10). Assays were separated based on the CpG density (HC, IC and LC) of the 500 bp around each assay with MeXiP being observed in HC and IC in both blood (upper panels) and placenta (lower panels).



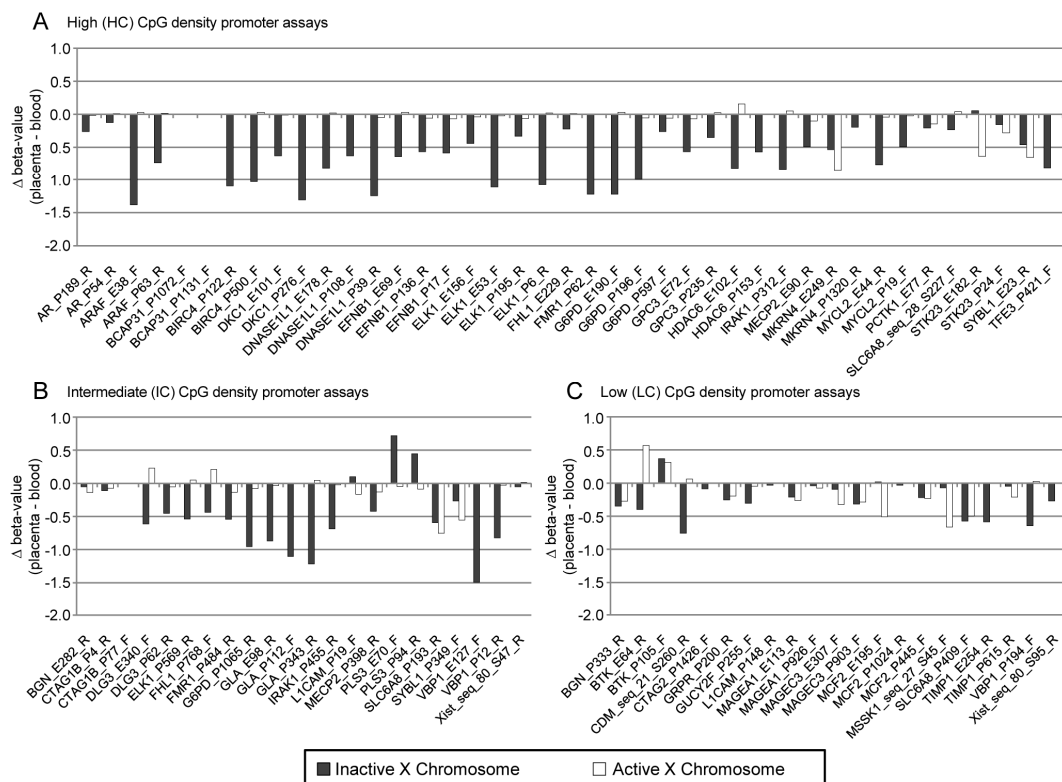
Supplementary Figure 2: Heatmap illustrating the percent methylation as determined by pyrosequencing at all 145 sites examined in the 31 pyrosequencing assays in 6 female blood, 6 female placentas (2 sites each), 6 male blood and 6 male placenta (2 sites each). DNA methylation levels represented as a gradient from blue (high methylation) to yellow (low methylation). Specific samples which failed for a particular assay were replaced with another sample and are marked in red with the replacement sample listed at the end of the figure.

Submitted as separate Excel file "Cotton Sup Fig 2.xls"

Supplementary Figure 3: The Xi shows less placental methylation compared with blood than the Xa at the majority of regions examined across the X chromosome. Percent methylation change from blood to placenta for Xa (white) and Xi (black) at 30 pyrosequencing assays. Negative percent change methylation indicates that blood is more methylated than placenta while positive shows that placenta is more methylated than blood. Assays are separated into CpG density classes, HC, IC and LC, by vertical lines. (A) promoter assays (B) intragenic assays (C) intergenic assays. Xa value is the level of methylation observed in male, Xi value is calculated by subtracting the Xa from the female methylation level multiplied by two.



Supplementary Figure 4: The Xi shows less placental methylation compared with blood than the Xa at the majority of promoters examined on the X chromosome. Beta-value methylation change from blood to placenta for Xa (white) and Xi (black) at Illumina GoldenGate panel assays. Negative percent change methylation indicates that blood is more methylated than placenta while positive shows that placenta is more methylated than blood. Assays are separated into CpG density classes, HC, IC and LC, by vertical lines. (A) promoter assays (B) intragenic assays (C) intergenic assays. Xa value is the level of methylation observed in male, Xi value is calculated by subtracting the Xa from the female methylation level multiplied by two.



Supplementary Table 1: Primer sequences and cycling conditions used in pyrosequencing assays. PCR product size, assay class and CpG class of each assay also listed.

Name ^a	Sequence (5' to 3')	Annealing Temperature	PCR product size	Assay Class	CpG class
PDK3_78_F1 PDK3_78_R1_B PDK3_78_S1	GGTTGTA AAAATTAAGTGTAGGA /Biotin/AACCCAACCCAACAATACAA AAAATTAAGTGTAGGATG	57°C	211	Promoter	HC
UXT_89_F1 UXT_89_R1B UXT_89_S1	GTTAATGGGGGATTGTA AAAAG /Biotin/TCACTTCCTCTACCTCCACCTAT ATGGGGGATTGTA AAAA	57°C	130	Promoter	HC
FANCB_93_F1B FANCB_93_R1 FANCB_93_S1	/Biotin/TTTGGGGAGTGTGTGAAAGTA AACCAAACCCTCAACCTAAATC CCTCAACCTAAATCCCAT	57°C	167	Promoter	HC
PGRMC1_95_F1 PGRMC1_95_R1_B PGRMC1_95_S1	GGGGAAGGGTTATTAAGGAGAG /Biotin/CCCATTCTAAAACCCCTCATCT GGGAAGGGTTATTAAGGA	57°C	164	Promoter	HC
CpG186_89_F1_B CpG186_89_R1 CpG186_89_S1	/Biotin/TGTAGTTTGGATATTTTGATGGG AACCAATCCTACCTTACAACCT TCCTACCTTACAACCTTT	57°C	220	Promoter	HC
NDP_F1 NDP_R1 NDP_S1	AGAGAGAGAATGTAAATGGAAAAGTGTA /Biotin/ATTAACCTCTTATAATCCATAATACCA AGAGAATGTAAATGGAAA	57°C	255	Promoter	IC
CHM_F1 CHM_R1 CHM_S1	GTGGGAGATTTGGATATTTTTGAT /Biotin/AAATAAAAATCTCCTTATTCACAAAAC GATAATATTGAAGTAAAATTGTTAG	57°C	111	Promoter	IC
PHEX_F1 PHEX_R1 PHEX_S1	AGTTTTTAAAGTGTGGGATTATAGG /Biotin/ACTTCAACAAATCCCCAAAATAAA AAAGTGTGGGATTATAGG	57°C	93	Promoter	LC

II1RAPL1_F1 II1RAPL1_R1 II1RAPL1_S1	/Biotin/TTGGGGAGATAGTGATGGG CACACTCTTAATAACCTCCTTTTCATC ATCTCTTCTCTTTAAAACAAAT	55°C	91	Promoter	LC
ODZ1_F1 ODZ1_R1 ODZ1_S1	GTATTAAGGATTAAGTTGGAGGTTGTAGT /Biotin/TTATACTCCTCACCCTTTCAAATCTAAT ATAGTTTTTAAAAATATTGTATTG	57°C	193	Promoter	LC
ARHGAP6_F1 ARHGAP6_R41 ARHGAP6_S1	/Biotin/ATTTGATTGAAGGTTGAATGAG CCAACCCTAAATTCAATATTCTT CAATATTCTTTACCCCA	64.5°C	149	Within Gene	HC
MXA5_E_F1 MXA5_E_R1 MXA5_E_S1	TTTTTTGATGGAAAGGGTT /Biotin/TCTTCCCTAACAAAAAATATAACAAACT TTTTTTGATGGAAAGG	57°C	90	Within Gene	HC
ShROOM2_F1 SHROOM2_R1 SHROOM2_S1	GGTGGAGAATGTTTTAATAATTG /Biotin/CCCCCATTCCAAATCAA GGTGGAGAATGTTTTA	53°C	86	Within Gene	IC
BHLHB9_F2 BHLHB9_R2 BHLHB9_S2	/Biotin/GGGGTTTTTTGAGGTAGTTGGTGT CCCCTCTCAAACCCACCTTAAT TCTCAAACCCACCTTAAT	57°C	102	Within Gene	IC
IRCH2_int_F1 IRCH2_int_R1 IRCH2_int_S1	GAGTAGGAGGTTATTATGAGGAGAA /Biotin/ACTAAACTACTATAACCCCCACTATAAAT GGAGGTTATTATGAGGAGA	57°C	101	Within Gene	IC
POLA_F3 POLA_R2	/Biotin/GGGGGGTAGTGTTTTATGTATATTAAT ACCACATAAAACCCACACATATAAT	57°C	115	Within Gene	LC

POLA_S5	ATAAACTAACTTTTCCTATC				
GLRA2_Int_F3	/Biotin/GAATTTTTGATGGATTGGATATGG				
GLRA2_Int_R4	CCTTCTATTA ACTCCACACTCCTATATCA	57°C	183	Within Gene	LC
GLRA2_Int_S2	ATCTCATAACTATCTACATTAACC				
GLRA2_E4_F3	TGTAAATAGAATTTTTGTGTTAGGGTAAT				
GLRA2_E4_R1	/Biotin/ATAGAATTTTTGTGTTAGGG	57°C	135	Within Gene	LC
GLRA2_E4_S3	ATAGAATTTTTGTGTTAGGG				
WNK5_Int_F1	/Biotin/TAAAAATTAGTTGGGAGTGGTGGTAGG				
WNK5_Int_R1	CTCATTTACATTTTCCTCCCTCATCA	57°C	217	Within Gene	LC
WNK5_Int_S1	CCAAATTAATAACAATAACACA				
TBL1X_int_F1	TGTGTTAAGTTTGATTGTAGAAATGAAT				
TBL1X_int_R1	/Biotin/CCCTAAATAATAATCTCAATTTTCCTCATA	55°C	147	Within Gene	LC
TBL1X_int_S1	GTAGAAATGAATTTGAAGAAG				
CpG145_89_F1	TTGGATTTGTTTGTAGGATTG				
CpG145_89_R1_B	/Biotin/CAAACCCA ACTCAATAACCT	57°C	182	Between Genes	HC
CpG145_89_S1	GGATTTGTTTGTAGGAT				
CpG46_F1	/Biotin/GGTTTTAGTGGTTTTGATTTATAGAGT				
CpG46_R1	CTCCTCTACTAAAAACAACCTACC	57°C	108	Between Genes	HC
CpG46_S1	TCCTCTACTAAAAACAACCT				
CpG36_F1	GGAAAGGAAAAGGGAGAATT				
CpG36_R20	/Biotin/CCCTCACC ACTAAACAATTAA	57°C	80	Between Genes	HC
CpG36_S18	GGAAAGGAAAAGGGAGAATT				

rs5960421_1_F1	GGTTGTAGAGTGTGGTAGAGG	57°C	143	Between Genes	IC
rs5960421_1_R1	/Biotin/CCCTCCCACCAAATCAAAT				
rs5960421_1_S1	AGTGTGGTAGAGGTGTT				
CpG70_F1	GTTGAAGTAGGAGGTTGGATGTA	55°C	169	Between Genes	IC
CpG70_R1	/Biotin/CTAAACTCCTATTCTCCAATTATAACAAC				
CpG70_S1	GAAGTAGGAGGTTGGAT				
rs36021843_F1	/Biotin/ATGGTTGGTTATATGGTTATTAGAGTT	57°C	185	Between Genes	LC
rs36021843_R1	CCCTAAAAAATAACCTCCTACTTAACTAT				
rs36021843_S1	AATAATATTCCACCTCCC				
rs1212068_1_F1	/Biotin/TGAGAGATGAGTGTTATGGAGAAA	64.5°C	183	Between Genes	LC
rs1212068_1_R1	CAAAAACAAACTCTCCAAATTCA				
rs1212068_1_S1	TCTCCAAATTCAAATCAAT				
rs34350719_F1	GTTTTGGGTTGGAAAAATTAGAGT	57°C	79	Between Genes	LC
rs34350719_R1	/Biotin/CCCATAAAATTCAAAAAACTTCTACCT				
rs34350719_S1	TGGGTTTGGAAAAATTAG				
rs17308229_F1	GGTTTTTTATTTTTGAGATTGTTAG	64°C	214	Between Genes	LC
rs17308229_R1	/Biotin/AACCACTCAAATCTACAAACAATA				
rs17308229_S1	TATTATAAGTTATTGTATTAGGG				
rs4825396_F1	/Biotin/TTTTTGATGGGGGAGAAGGGT	57°C	113	Between Genes	LC
rs4825396_R1	CCCATCCTAATCTCCTATTTCTTATCC				
rs4825396_S1	TCCTATTTCTTATCCACA				

a) F: Forward primers, R: Reverse primer, S: Sequencing primers

Supplementary Table 2: Methylation assays for island (HC and IC) promoters failing to show MeXiP or containing features believed to interfere with MeXiP or showing discordant methylation results.

Assay name	Average Methylation				Features interfering with MeXiP ^a				Discordant Assay Methylation Patterns ^d	Previous methylation data ^e	Evaluation
	Female Blood	Female Placenta	Male Blood	Male Placenta	CpG Density	Repetitive Element ^b	Escapes X inactivation	Distance to TSS (bp) ^c			
AR_P54_R	0.0 9	0.0 4	0.0 0	0.0 1	HC	-	0/6	-54	Yes	(1)	The 2 Illumina assays for AR are discordant and P189 does not match previous methylation results.
AR_P189_R	0.4 5	0.3 3	0.0 7	0.1 0	HC	-	0/6	-189			
BCAP31_P1072_F	0.9 9	0.9 9	0.9 9	0.9 9	HC	-	3/9	-1074	-	(2)	Also near a duplication on chr 16 ^f .
BCAP31_P1131_F	0.9 9	0.9 9	0.9 9	0.9 9	HC	-	3/9	-1133			
BGN_E282_R	0.7 2	0.5 9	0.2 0	0.1 3	IC	-	0/5	282	Yes	-	The 2 Illumina assays for BGN are discordant however the other assay is an LC.
CTAG1B_P4_R	1.0 0	0.9 0	0.9 9	0.9 2	IC	-	-	-4	-	(3,4)	CT gene family highly methylated, additionally many other CTs on the array were LCs ^g .
CTAG1B_P77_F	1.0 0	0.9 9	1.0 0	0.9 9	IC	-	-	-77			
DLG3_E340_F	0.8 4	0.6 7	0.0 2	0.2 7	IC	-	-	340	-	-	Shows MeXiP despite distance to TSS.
FHL1_E229_R	0.4 6	0.3 4	0.0 0	0.0 5	HC	-	1/9	229	-	-	Shows MeXiP despite distance to TSS and/or presence of a LINE.
FHL1_P768_F	0.5 0	0.3 9	0.0 4	0.2 6	IC	LINE	1/9	-768			

FMR1_P484_R	0.9 7	0.6 0	0.6 8	0.5 7	IC	-	1/9	-484	Yes	(5,6,7)	The 2 Illumina assays for FMR1 are discordant and P484 does not match previous methylation results.
FMR1_P62_R	0.6 9	0.0 8	0.0 1	0.0 1	HC	-	1/9	-62			
G6PD_E190_F	0.7 4	0.1 5	0.0 1	0.0 4	HC	-	0/5	-783	-	(8, 9)	Shows MeXiP despite distance to TSS and/or presence of a LINE.
G6PD_P1065_R	0.8 7	0.3 7	0.1 3	0.0 7	IC	LINE	0/5	472			
L1CAM_P19_F	0.6 2	0.5 6	0.3 0	0.1 8	IC	-	-	-19	Yes	-	The 2 Illumina assays for L1CAM are discordant however the other assay is an LC.
MKRN4_E249_R	0.9 9	0.3 1	0.9 9	0.1 5	HC	-	-	249	Yes	-	Pseudogene
MKRN4_P1320_R	0.9 9	0.9 0	0.9 9	0.9 8	HC	-	-	-1320	Yes		
NDP	13%	16%	6%	9%	IC	-	-	-59	-	-	undetermined
PCTK1_E77_R	0.7 2	0.5 4	0.8 1	0.6 6	HC	-	6/6	77	-	(10)	Assay does not match previous methylation results.
PLS3_E70_F ⁱ	0.2 9	0.6 1	0.1 1	0.1 1	IC	-	5/9	70	-	(11)	Previous methylation results only examined methylation in males.
PLS3_P94_R	0.3 0	0.4 9	0.2 6	0.2 0	IC	-	5/9	-94			
SLC6A8_P193_R	0.8 6	0.1 7	0.8 4	0.1 0	IC	-	-	-193	Yes	(2)	The 3 Illumina assays for SLC6A8 are discordant however the other assay is an LC. Previous methylation results only examined methylation in males and there is also has a pseudogene on chr 16 ^f .
SLC6A8_seq_28_S227_F	0.7 3	0.6 3	0.0 0	0.0 9	HC	-	-	-1681			
STK23_E182_R	0.9 9	0.7 2	0.8 7	0.2 6	HC	-	-	182	Yes	(2)	The 2 Illumina assays for STK are discordant and Previous methylation results only examined methylation in males.
STK23_P24_F	0.8 7	0.6 3	0.4 5	0.2 1	HC	-	-	-24			

SYBL1_E23_R	0.7 5	0.2 1	0.7 2	0.0 7	HC	-	0/5	23	-	(12,8)	Silent on both X and Y chromosomes. P349 shows MeXiP despite presence of a LINE.
SYBL1_P349_F	0.5 0	0.1 1	0.5 7	0.0 1	IC	LINE	0/5	-349	-		
Xist_seq_80_S47_R	0.8 5	0.8 3	0.9 7	0.9 7	IC	-	-	-31	Yes	(13,14 ,15)	The 2 Illumina assays for XIST are discordant however the other assay is an LC. Is also expressed only from the Xi ^h .

- a) Grey shading represents possible features which may interfere with MeXiP.
- b) Three LC assays (CTAG2_P1426_F, MAGEC3_P903_F, TIMP1_P615_R) were also located within repetitive elements
- c) Seven LC assays (CDM_seq_21_S260_R, CTAG2_P1426_F, MAGEA1_P926_F, MAGEC3_E307_F, MAGEC3_P903_F, MCF2_P1024_R, TIMP1_E254_R) were also beyond 700 bp upstream or 200 bp downstream
- d) Grey shading indicates a gene with multiple Illumina assays which show different methylation patterns. Three genes (BTK, MCF2, TIMP1) had only LC assays and also were discordant between assays within the same gene.
- e) Grey shading indicates that Illumina methylation results conflict with previous methylation results.
- f) Recent genome-wide studies suggest a hypermethylation of pseudogenes and duplicated regions thus it is possible that the presence of a tandem duplication or pseudogene may predispose genes to hypermethylation which may explain the high methylation seen for *BCAP* and *SLC6A8* ^{(2),(16)}.
- g) Members of cancer-testis (CT) antigen family of genes are often found in palindromic repeats as multicopy genes and pseudogenes and have typically been shown to be highly methylated in all tissues except the germline – a pattern generally found for genes with germline-specific expression ^{(3),(2),(17)}. Consistent with high levels of methylation in all tissues other than testis, all MAGEs and CTAGs showed hypermethylation in blood and placenta regardless of CpG density emphasizing that gene function as well as CpG density is important in determining methylation status ^{(3),(2)}.
- h) Both XIST assays on the Illumina GoldenGate panel showed nearly 100% methylation in males however females showed methylation levels up to 95%. While the trend of these methylation levels was as expected the level of methylation in females appears to have been substantially overestimated by the Illumina assay.

References

1. Kubota, T., Nonoyama, S., Tonoki, H., Masuno, M., Imaizumi, K., Kojima, M., Wakui, K., Shimadzu, M. and Fukushima, Y. (1999) A new assay for the analysis of X-chromosome inactivation based on methylation-specific PCR. *Human Genetics*, **104**, 49-55.
2. Grunau, C., Hindermann, W. and Rosenthal, A. (2000) Large-scale methylation analysis of human genomic DNA reveals tissue-specific differences between the methylation profiles of genes and pseudogenes. *Hum Mol Genet.* , **9**, 2651-63.
3. De Smet, C., Lurquin, C., Lethé, B., Martelange, V. and Boon, T. (1999) DNA Methylation Is the Primary Silencing Mechanism for a Set of Germ Line- and Tumor-Specific Genes with a CpG-Rich Promoter. *Molecular and Cellular Biology*, **19**, 7327–7335.
4. Warburton, P.E., Giordano, J., Cheung, F., Gelfand, Y. and Benson, G. (2004) Inverted repeat structure of the human genome: the X-chromosome contains a preponderance of large, highly homologous inverted repeats that contain testes genes. *Genome Res*, **14**, 1861-9.
5. Panagopoulos, I., Lassen, C., Kristofferson, U. and Aman, P. (1999) A methylation PCR approach for detection of fragile X syndrome. *Hum Mutat.*, **14**, 71-9.
6. Carrel, L. and Willard, H.F. (1996) An assay for X inactivation based on differential methylation at the fragile X locus, FMR1. *Am. J. Med. Genet.*, **64**, 27-30.
7. Hansen, R., Gartler, S., Scott, C., Chen, S.-H. and Laird, C. (1992) Methylation analysis of CGG sites in the CpG island of the human FMR1 gene. *Hum. Mol. Genet.*, **1**, 571-578.
8. Huppke, P., Bohlander, S., Krämer, N., Laccone, F. and Hanefeld, F. (2002) Altered methylation pattern of the G6 PD promoter in Rett syndrome. *Neuropediatrics*, **33**, 105-8.
9. Wolf, S.F., Dintzis, S., Toniolo, D., Persico, G., Lunnen, K.D., Axelman, J. and Migeon, B.R. (1984) Complete concordance between glucose -6- phosphate dehydrogenase activity and hypomethylation of 3' CpG clusters: implications for X chromosome dosage compensation. *Nucl. Acids Res.*, **12**, 9333-9348.
10. Carrel, L., Clemson, C.M., Dunn, J.M., Miller, A.P., Hunt, P.A., Lawrence, J.B. and Willard, H.F. (1996) X inactivation analysis and DNA methylation studies of the ubiquitin activating enzyme E1 and PCTAIRE-1 genes in human and mouse. *Hum. Mol. Genet.*, **5**, 391-402.
11. Oprea, G.E., Kröber, S., McWhorter, M.L., Rossoll, W., Müller, S., Krawczak, M., Bassell, G.J., Beattie, C.E. and Wirth, B. (2008) Plastin 3 is a protective modifier of autosomal recessive spinal muscular atrophy. *Science*, **320**, 524-7.

12. Huber, R., Hansen, R.S., Strazzullo, M., Pengue, G., Mazzarella, R., D'Urso, M., Schlessinger, D., Pilia, G., Gartler, S.M. and D'Esposito, M. (1999) DNA methylation in transcriptional repression of two differentially expressed X-linked genes, GPC3 and SYBL1. *Proc Natl Acad Sci U S A*, **96**, 616-21.
13. Vasques, L.R., Stabellini, R., Xue, F., Tian, X.C., Soukoyan, M. and Pereira, L.V. (2005) XIST repression in the absence of DNMT1 and DNMT3B. *DNA Res*, **12**, 373-8.
14. Song, M.A., Park, J.H., Jeong, K.S., Park, D.S., Kang, M.S. and Lee, S. (2007) Quantification of CpG methylation at the 5'-region of XIST by pyrosequencing from human serum. *Electrophoresis*, **28**, 2379-84.
15. Hendrich, B.D., Brown, C.J. and Willard, H.F. (1993) Evolutionary conservation of possible functional domains of the human and murine XIST genes. *Hum. Mol. Genet.*, **2**, 663-672.
16. Rauch, T.A., Wu, X., Zhong, X., Riggs, A.D. and Pfeifer, G.P. (2009) A human B cell methylome at 100-base pair resolution. *PNAS*, **106**, 671:8.
17. Weber, M., Hellmann, I., Stadler, M.B., Ramos, L., Paabo, S., Rebhan, M. and Schubeler, D. (2007) Distribution, silencing potential and evolutionary impact of promoter DNA methylation in the human genome. *Nat Genet*, **39**, 457-66.