

Table W1. List of Antibodies.

Antibody	Supplier
$\alpha_6\beta_4$	Clone B5.5 [1]
Actin	BD, Heidelberg, G.
Annexin V	Santa Cruz, Heidelberg, G.
β -Catenin	BD, Heidelberg, G.
bFGF	Oncogene, Boston, MA
C3	MP Biomed., Eschwege, G.
C4.4A	Clone C4.4 [1]
Caveolin	BD, Heidelberg, G.
CD44s	Clone Ox50 (EAACC)
CD44v6	Clone A2.6 [1]
CD104	BD, Heidelberg, G.
CD106	Biozol, Eching, G.
CD11a	BD, Heidelberg, G.
CD11b	Clone Ox42 (EAACC)
CD11c	Clone Ox41 (EAACC)
CD13	[2]
CD133	Abcam, Cambridge, UK
CD151	[3]
CD18	BD, Heidelberg, G.
CD24	Santa Cruz, Heidelberg, G.
CD29	BD, Heidelberg, G.
CD31	BD, Heidelberg, G.
CD49a	BD, Heidelberg, G.
CD49b	BD, Heidelberg, G.
CD49c	BD, Heidelberg, G.
CD49d	BD, Heidelberg, G.
CD49e	BD, Heidelberg, G.
CD49f	Abcam, Cambridge, UK
CD53	BD, Heidelberg, G.
CD54	Biozol, Eching, G.
CD61	Biozol, Eching, G.
CD81	Santa Cruz, Heidelberg, G.
CD9	BD, Heidelberg, G.
Claudin-7	[4]
c-Met	New England Biolabs, Frankfurt, G.
Collagen I	Rockland, Gilbertsville, PA
Collagen II	LabVision, Fremont, CA
Collagen III	ARB, Golden, CO
Collagen IV	Rockland, Gilbertsville, PA
D6.1A	Clone D6.1 [1]
EpCAM	Clone D5.7 [1]
EWI-F	[5]
EGFR	Santa Cruz, Heidelberg, G.
FN	BD, Heidelberg, G.
Galectin	Santa Cruz, Heidelberg, G.
HAS3	Abcam, Cambridge, UK
HSP-1	Santa Cruz, Heidelberg, G.
Hyaluronan	Rockland, Gilbertsville, PA
INS-2	Santa Cruz, Heidelberg, G.
Laminin 1	Rockland, Gilbertsville, PA
Laminin 5	BD, Heidelberg, G.
MMP13	Dianova, Hamburg, G.
MMP2	Dianova, Hamburg, G.
MMP9	Dianova, Hamburg, G.
Osteopontin	Santa Cruz, Heidelberg, G.
PDGF	BD, Heidelberg, G.
PDGFR	BD, Heidelberg, G.
PGK-1	Santa Cruz, Heidelberg, G.
S100A4	Abcam, Cambridge, UK
SDF-1	Abcam, Cambridge, UK
Tenascin	LabVision, Fremont, CA
TGF β	Santa Cruz, Heidelberg, G.
Thrombospondin	Santa Cruz, Heidelberg, G.
TNF α	BD, Heidelberg, G.
uPA	Calbiochem, Darmstadt, G.
uPAR	Calbiochem, Darmstadt, G.
VEGF	Biotrend, Köln, G.
VEGFR1	Biotrend, Köln, G.
VEGFR2	Biotrend, Köln, G.
VN	Biotrend, Köln, G.
vWF	Abcam, Cambridge, UK

CD44s indicates CD44 standard isoform; EAACC, European Association of Animal Cell Cultures, Porton Down, UK; G., Germany.

References

- [1] Matzku S, Wenzel A, Liu S, and Zöller M (1989). Antigenic differences between metastatic and nonmetastatic rat tumor variants characterized by monoclonal antibodies. *Cancer Res* **49**, 1294–1299.
- [2] Chang YW, Chen SC, Cheng EC, Ko YP, Lin YC, Kao YR, Tsay YG, Yang PC, Wu CW, and Roffler SR (2005). CD13 (aminopeptidase N) can associate with tumor-associated antigen L6 and enhance the motility of human lung cancer cells. *Int J Cancer* **116**, 243–252.
- [3] Claas C, Wahl J, Orlicky D, Karaduman H, Schnölzer M, Kempf T, and Zöller M (2005). The tetraspanin D6.1A and its molecular partners on rat carcinoma cells. *Biochem J* **389**, 99–110.
- [4] Langbein L, Pape UF, Grund C, Kuhn C, Prätzel S, Moll I, Moll R, and Franke WW (2003). Tight junction-related structures in the absence of a lumen: occludin, claudins and tight junction plaque proteins in densely packed cell formations of stratified epithelia and squamous cell carcinomas. *Eur J Cell Biol* **82**, 385–400.
- [5] Orlicky DJ, Lieber JG, Morin CL, and Evans RM (1998). Synthesis and accumulation of a receptor regulatory protein associated with lipid droplet accumulation in 3T3-L1 cells. *J Lipid Res* **39**, 1152–1161.

Table W2. List of Primers.

The following primers were used		
CD133	Sense	5'-AGCCAAGACACCTTCAATGC-3'
	Antisense	5'-ACGGTGTGAGTTCCTGTC-3'
CD166	Sense	5'-AACCTGGAGAGTCAGAGCA-3'
	Antisense	5'-TGCGAGCTGTGATTTGTTTC
CD24	Sense	5'-ACATCGGTTGCACCAATTTTC-3'
	Antisense	5'-GAGAGAGAGGCCAGGAGAC-3'
GADPH	Sense	5'-GACCCCTTCATTGACCTCAAC-3'
	Antisense	5'-CTTCTCCATGGTGGTGAAGAC-3'

Amplification was performed for 30 cycles starting with 1 μ g of complementary DNA. Annealing temperatures have been as follows: CD133, 57°C; CD24 and CD166, 60°C; and GAPDH, 55°C. Polymerase chain reaction products were separated by electrophoresis in a 1.5% agarose gel.

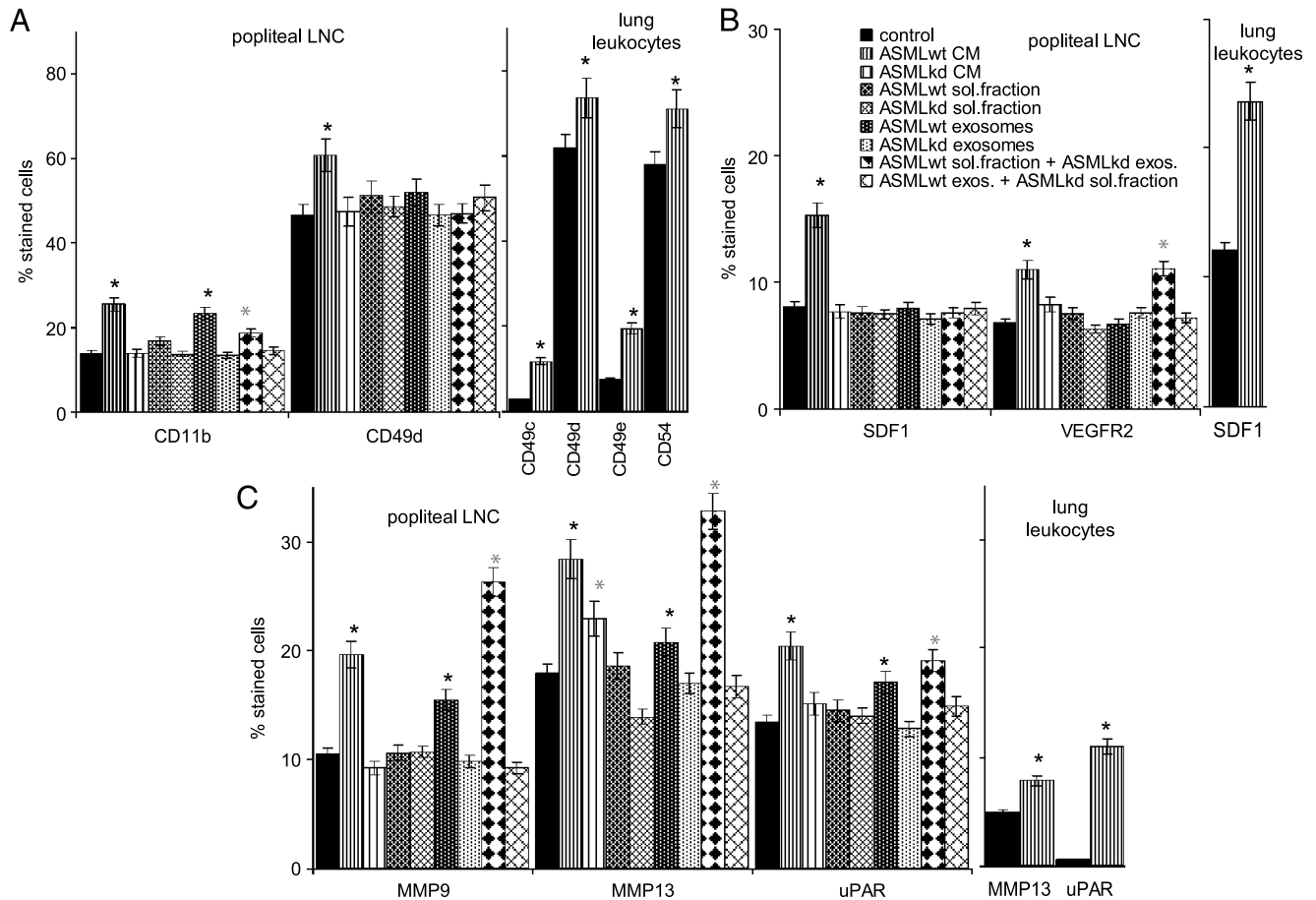


Figure W1. The impact of ASML^{wt}-CM on gene expression in target cells. BDX rats received ASML^{wt}- or ASML^{kd}-CM or fractions thereof and thereafter ASML^{kd} cells as described in Figure 1. LN and lung leukocytes were isolated. Expression of (A) integrins, (B) chemokines, chemokine receptors, and (C) matrix degrading enzymes was evaluated by flow cytometry. The percentage of stained cells (mean \pm SD of three experiments) is shown. Significant differences between control medium *versus* CM and fractions thereof are indicated by \star ; significant differences between ASML^{wt}- *versus* ASML^{kd}-CM and fractions thereof are indicated by \star .

Table W3. ASML^{wt} Versus ASML^{kd} Cells: Protein Expression and Recovery in the Soluble Matrix and in Exosomes.

Marker	Cells	Soluble Fraction	Exosomes
	wt/kd	wt/kd	wt/kd
Metastasis markers			
CD44	+/+	nt/nt	nt/nt
CD44v6	+++/-	+++/-	-/-
EpCAM	+++/>+++	-/-	+++/>+++
CD133	±/±	nt/nt	nt/nt
CD166	±/±	nt/nt	nt/nt
C4.4A	+++/>+++	-/-	+++/>+++
α ₆ β ₄	+++/>+++	+/-	+++/>+++
CD24	+/+	-/-	+/+
S100A4	+++/>+++	-/-	+++/>+++
Matrix proteins			
Coll I	+++/>+++	+++/>+++	nt/nt
Coll II	+++/>+++	+++/>+++	nt/nt
Coll III	+/+	+/+	nt/nt
Coll IV	+++/>+++	+++/>+++	nt/nt
FN	+++/>+++	+++/>+++	±/±
Galectin 3	+++/>+++	+++/>+++	nt/nt
HA	+++/>+++	+++/>+++	nt/nt
LN1	+++/>+++	+++/>+++	nt/nt
LN5	+++/>+++	+++/>+++	+/+
Tenascin	+++/>+++	+++/>+++	nt/nt
VN	+++/>+++	+++/>+++	nt/nt
Adhesion molecules			
CD11a	+/+	±/±	+/+
CD11b	+/+	-/-	+/+
CD11c	+/+	-/-	+/+
CD18	+/+	±/±	+/+
CD29	+++/>+++	+/+	+++/>+++
CD49b	+/+	-/-	+/+
CD49c	±/±	-/-	+++/>+++
CD49d	±/±	-/-	±/±
CD49e	+/+	±/±	+/+
CD49f	+++/>+++	-/-	+++/>+++
CD61	±/±	-/-	nt/nt
CD104	+++/>+++	+/-	+++/>+++
Enzymes and receptors			
uPA	+++/>+++	nt/nt	nt/nt
uPAR	+++/>+++	+++/>+++	+++/>+++
MMP2	+++/>+++	+/+	+++/>+++
MMP9	+/+	+/+	±/±
MMP13	+/+	-/-	+/+
CD13	+++/>+++	+++/>+++	+++/>+++
HAS3	+++/>+++	+++/>+++	-/-
PGK-1	+++/>+++	+/+	+++/>+++
Tetraspanins			
CD9	+++/>+++	+++/>+++	+++/>+++
CD81	+++/>+++	-/-	+++/>+++
CD151	+++/>+++	+++/>+++	+++/>+++
D6.1A	+++/>+++	-/-	+++/>+++
Cytokines and receptors			
HGF	+++/>+++	-/-	+++/>+++
PDGF	+/+	nt/nt	nt/nt
SDF-1	+/+	+/+	nt/nt
VEGF	+/+	+/+	nt/nt
OPN	+/+	-/-	-/-
TSP	+++/>+++	±/±	+++/>+++
c-Met	+++/>+++	+++/>+++	-/-
VEGFR1	+/+	nt/nt	nt/nt
VEGFR2	+/+	nt/nt	nt/nt
Others			
C3	+++/>+++	+++/>+++	+++/>+++
Annexin II	+++/>+++	-/-	+++/>+++
Annexin V	+++/>+++	-/-	+++/>+++
HSP-1	+++/>+++	-/-	+++/>+++
Caveolin	+++/>+++	-/-	+++/>+++
EWI-F	+++/>+++	±/±	+++/>+++
INS-2	+++/>+++	+/+	+++/>+++
Claudin-7	+++/>+++	-/-	+++/>+++

Coll indicates collagen; *kd*, knock down; *wt*, wild type; --, negative; ±, weak; +, distinct; ++, strong; +++, very strong; *nt*, not tested.

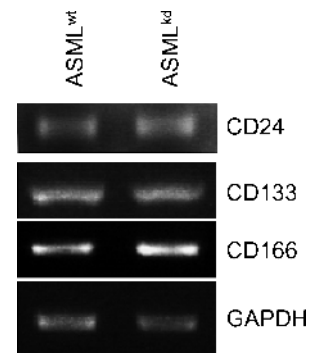


Figure W2. CIC marker expression in ASML^{wt} and ASML^{kd} cells. Expression of the CIC markers CD24, CD133, and CD166 in ASML^{wt} and ASML^{kd} cells was evaluated by reverse transcription-polymerase chain reaction. These markers are expressed by both ASML^{wt} and ASML^{kd} cells.

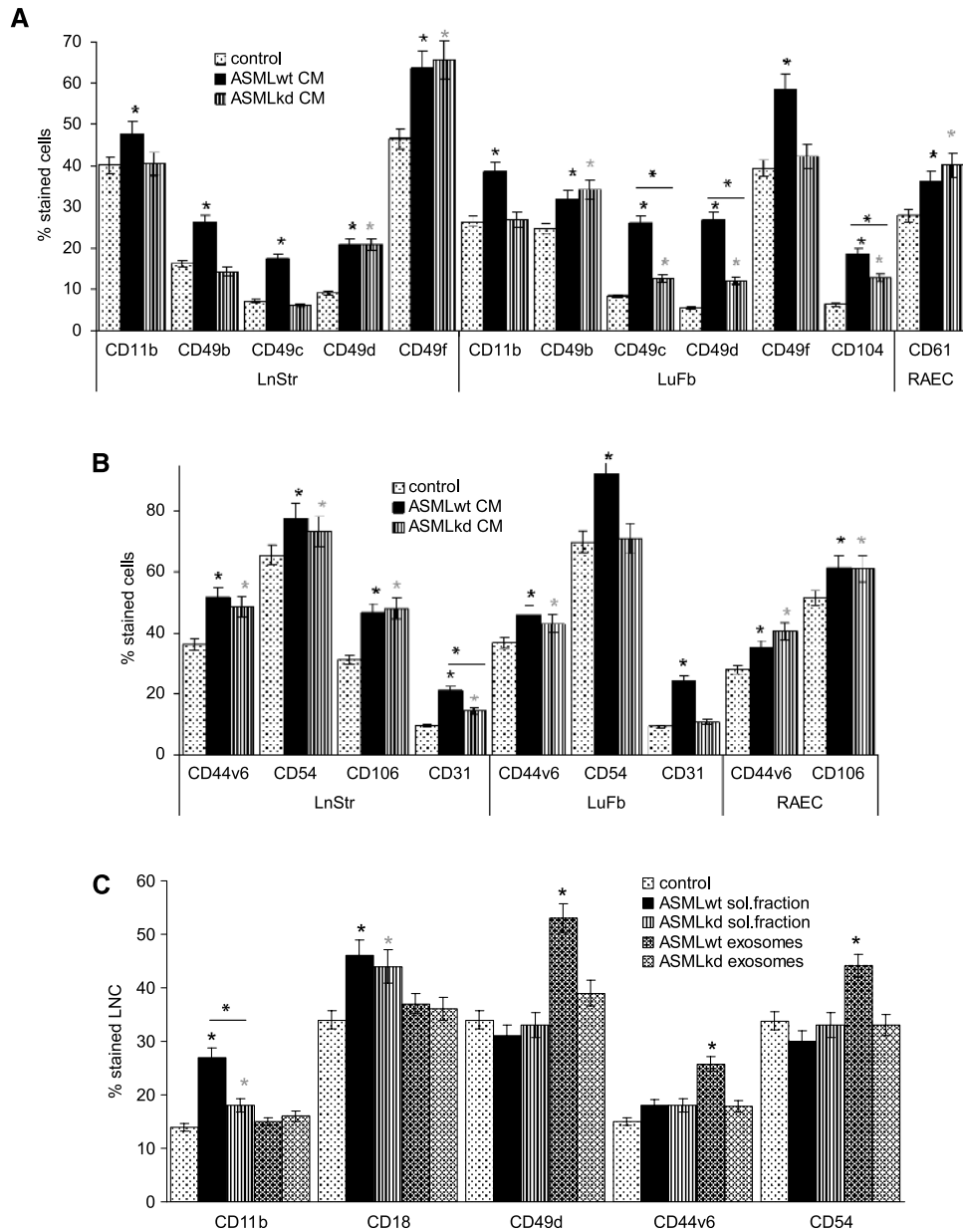


Figure W3. Modulated gene expression by ASML^{wt}-conditioned medium. (A, B, D, E, and F) LnStr, LuFb, and RAEC were cultured for 48 hours in the presence of ASML^{wt}- and ASML^{kd}-CM. (C) LNC and (G) LnStr, LuFb, and RAEC were cultured for 48 hours in the presence of ASML^{wt}- and ASML^{kd}-soluble fraction or exosomes. Cells were harvested, and the expression of (A–C) adhesion molecules and (D and E) cytokines/chemokines and receptors and (F and G) matrix degrading enzymes was evaluated by flow cytometry. Mean values \pm SD of three experiments are shown. Significant differences in comparison the control medium are indicated by \star ; significant differences between ASML^{wt}- versus ASML^{kd}-CM and fractions thereof are indicated by \ast .

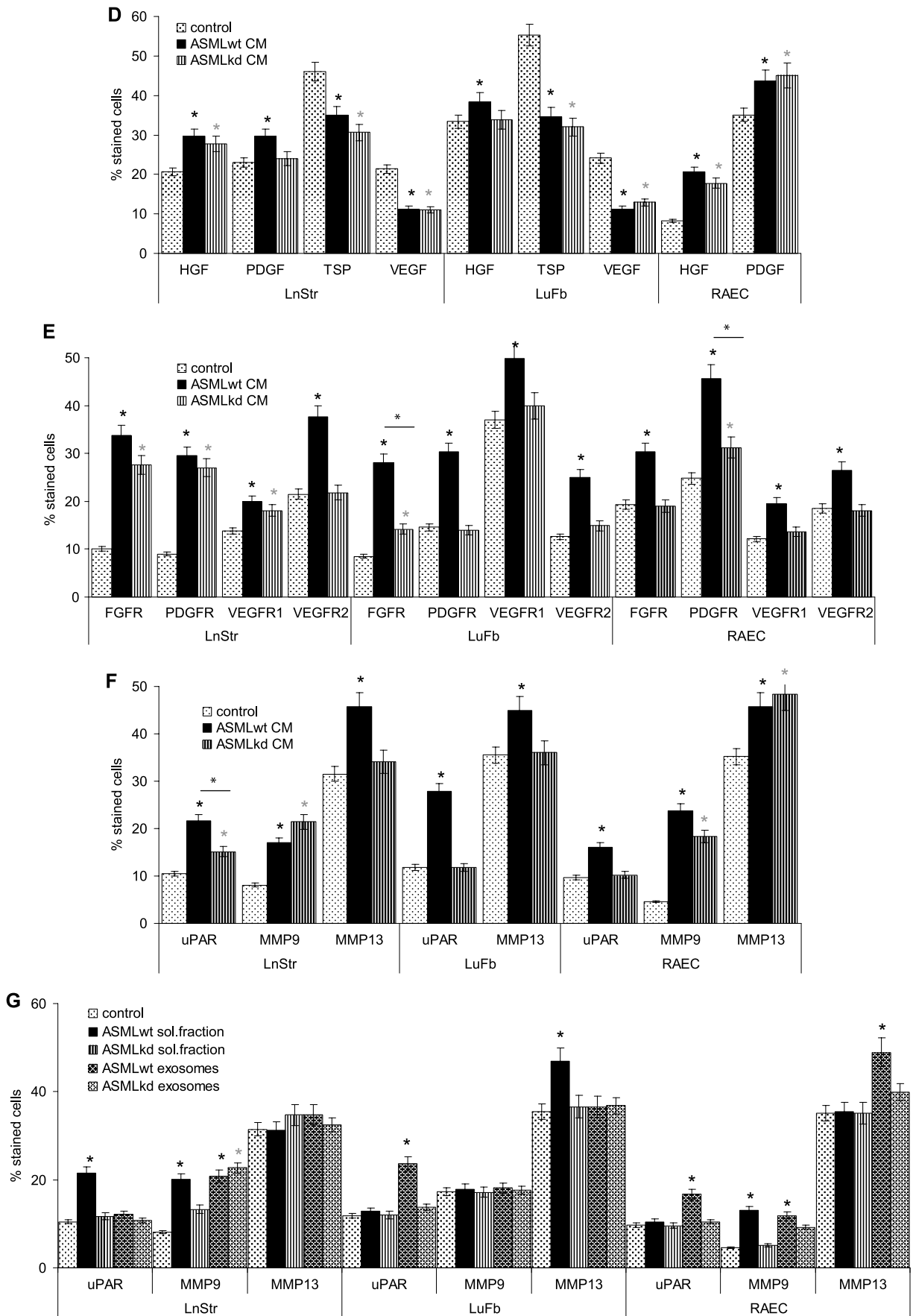


Figure W3. (continued).