

**Heme regulation of human cystathionine β -synthase activity: Insights
from fluorescence and Raman spectroscopy**

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Supporting Information

Full Reference Details

30. Kraus, J. P.; Janosik, M.; Kozich, V.; Mandell, R.; Shih, V.; Sperandeo, M. P.; Sebastio, G.; de Franchis, R.; Andria, G.; Kluijtmans, L. A. J.; Blom, H.; Boers, G. H. J.; Gordon, R. B.; Kamoun, P.; Tsai, M. Y.; Kruger, W. D.; Koch, H. G.; Ohura, T.; Gaustadnes, M., *Hum. Mutat.* **1999**, *13*, 362-375.

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Table S1 Frequencies (cm^{-1}) and assignment of bands in the resonance Raman spectra of PLP-Tris Schiff base, ferric, ferrous, and heat-inactivated ferrous and CO bound forms of hCBS, ferric, ferrous and CO bound forms of R266M hCBS.

Assignment	PLP-Tris Schiff base		hCBS						R266M hCBS		
			Ferric		Ferrous		Heat-inactivated ferrous	CO adduct	Ferric	Ferrous	CO adduct
	pH 8.6	pD 8.6	pH 8.6	pD 8.6	pH 8.6	pD 8.6	pH 8.5	pH 8.6	pH 8.5	pH 8.5	pH 8.5
PLP $\nu_{\text{C}=\text{N}}$	1645	1634	1665	1653	1665	1653			1663		
heme ν_{10}			1634	1633			1627	1632	1631		1631
heme $\nu_{\text{C}_a=\text{C}_b}$			1620		1621	1620	1618	1621	1619	1620	1620
heme ν_{37}			1605	1604	1611	1608	1605	1604		1604	1603
PLP ν_{8a}	1598	1603									
heme ν_2			1579	1581	1583	1584	1582	1584	1587	1582	1583
									1573		
heme ν_{38}			1556	1558	1554	1559	1556	1562	1551	1553	1556
PLP ν_{8b}	1539										
heme ν_{11}			1535		1535		1535				
		1520									
					1515	1518					
heme ν_3			1500	1502	1492	1492	1492	1497	1499	1491	1497
PLP ν_{19a}	1467	1471									
			1466	1467	1469	1470	1471	1469	1468	1470	1465
heme $\delta_s(=\text{CH}_2)$			1437	1437	1433	1430	1429	1431	1434	1430	1431
					1420						
PLP ν_{14}	1396										
heme ν_{12}			1394			1386	1388	1387	1391	1388	1386
heme ν_4			1372	1372	1357	1357	1360	1372	1372	1359	1371
	1364	1369									

PLP ν_{C-O}	1342	1341	1338	1340	1338	1340	1339	1339	1337	1336	
heme $\delta(C_aH)$			1315	1311	1314	1314	1310	1305	1312		1305
PLP $\nu_{C_4-C_4'}$	1309	1310									
			1217	1215			1228	1226	1217	1223	1225
PLP ν_{9a}	1191	1185									
heme ν_{30}			1174	1177			1170	1169	1171	1169	1171
							1128	1131	1119		1120

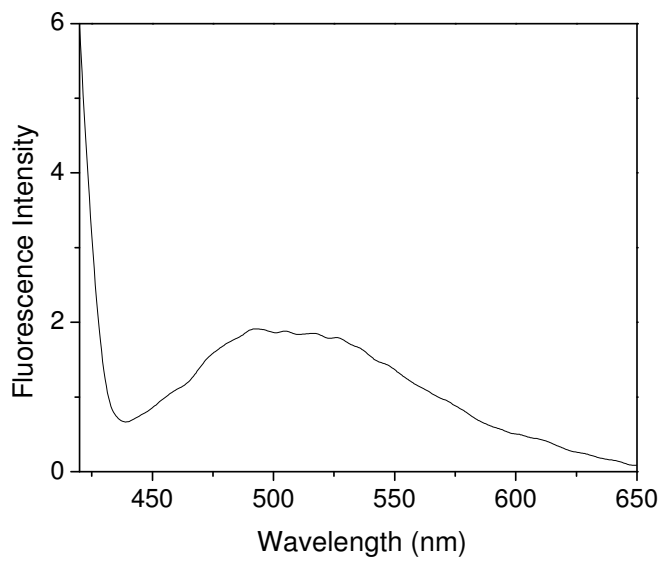


Figure S1. Fluorescence emission spectrum of PLP (106 μM) in Tris buffer (pH 8.5, 100 mM). Excitation wavelength, 410 nm.

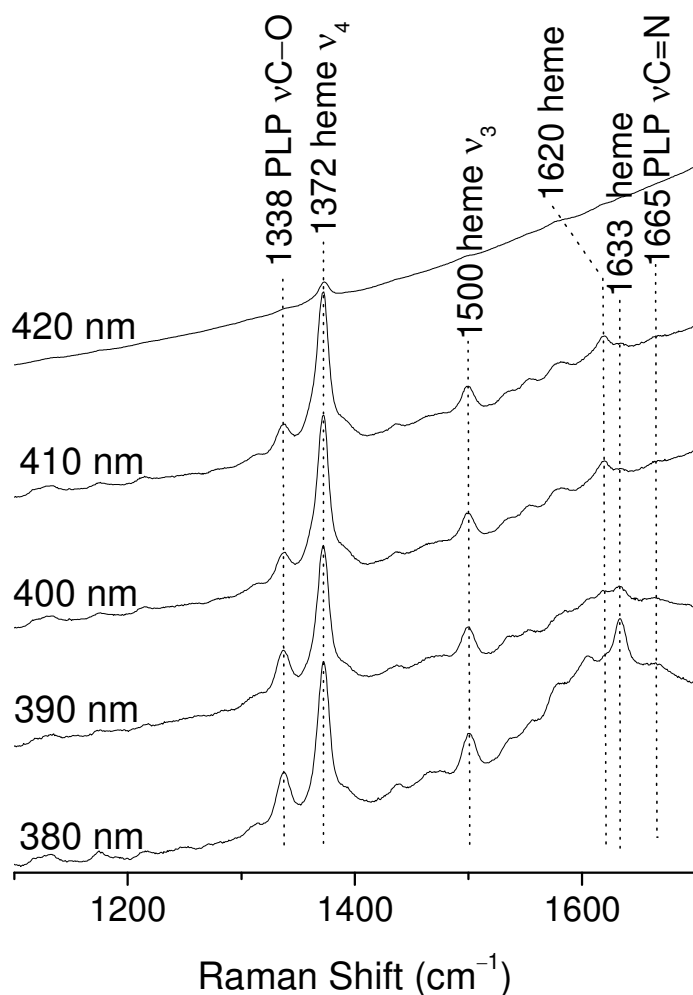


Figure S2. Raman spectra of ferric hCBS in Tris buffer pH 8.6 recorded using excitation wavelengths from 380-420 nm. Accumulation time, 15 min.

Though the heme modes of hCBS were the most intense Raman bands at all excitation wavelengths (380-420 nm), the relative enhancement of the PLP modes increased as the excitation wavelength became shorter, with the additional advantage that the fluorescence background was reduced at shorter excitation wavelengths. This was because the maximum absorption for the PLP Schiff base, 412 nm in the resting state, is at a shorter wavelength than the heme Soret band at 429 nm and 449 nm in the ferric and ferrous states, respectively.

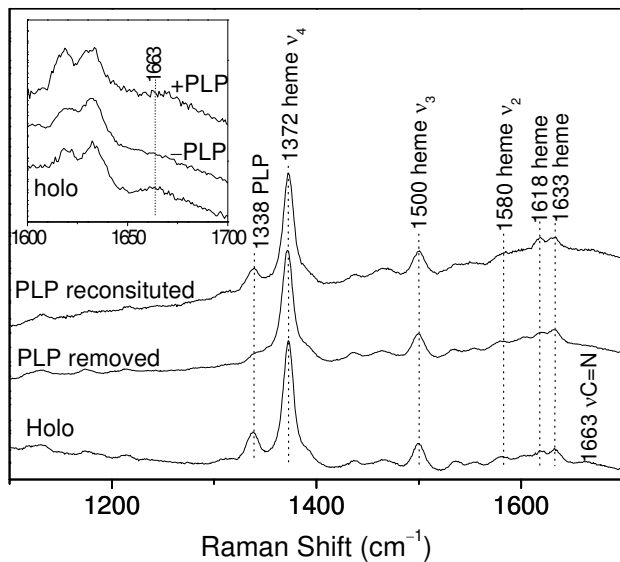


Figure S3. Raman spectra of Fe(III)hCBS before and after PLP removal by washing with serine and apo-Fe(III)hCBS reconstituted with PLP in 50 mM Tris buffer pH 8.6. Excitation wavelength, 390 nm. Accumulation time, 30 min.

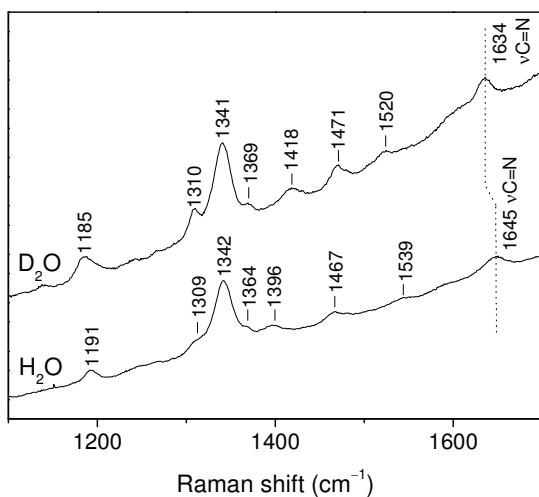


Figure S4. Raman spectra of the PLP-Tris Schiff base at pH (pD) 8.6 in H₂O and D₂O. Excitation wavelength, 390 nm. Accumulation time, 15 min.

The aldehyde group of PLP reacts with the amine of Tris [tris(hydroxymethyl)aminomethane] to form a Schiff base adduct in aqueous solution. Raman spectra were recorded of the PLP-Tris Schiff base as a control to check that signals from PLP in the Raman spectra of hCBS in Tris buffer solutions were coming from PLP bound to hCBS and not PLP that had been released from the protein and reacted with the buffer. The D₂O experiment was also done to demonstrate the shift in the vC=N band upon deuteration of the PLP-Schiff base.

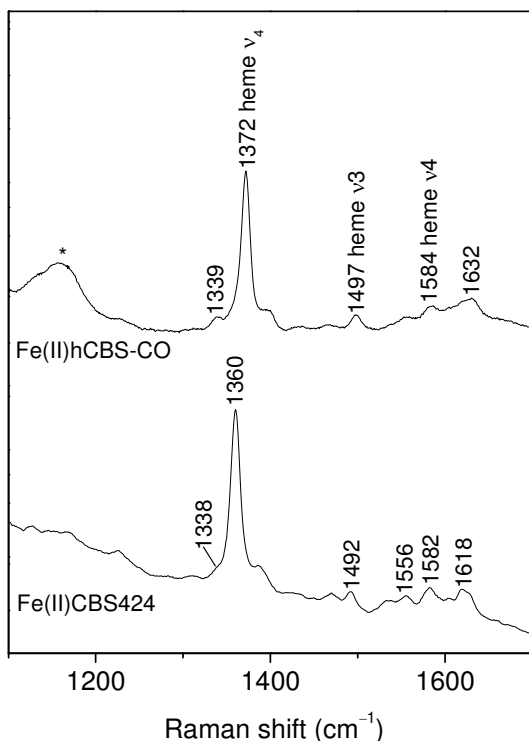


Figure S5. Raman spectra of Fe(II)CBS424 and Fe(II)hCBS-CO formed from Fe(II)hCBS 20 min after the initiation of CO binding ($\lambda_{\text{ex}} = 390 \text{ nm}$) in 100 mM Tris buffer pH 8.6. Accumulation time, 10 min. *Peak arising from the reductant, dithionite.

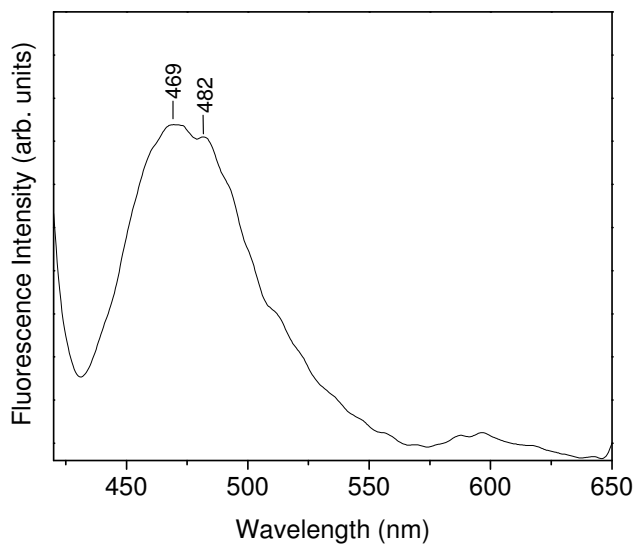


Figure S6. Fluorescence spectrum 23 h after initiation of CO binding to Fe(II)CBS (36 μM) in 100 mM phosphate buffer pH 8.0. 410 nm excitation spectrum.

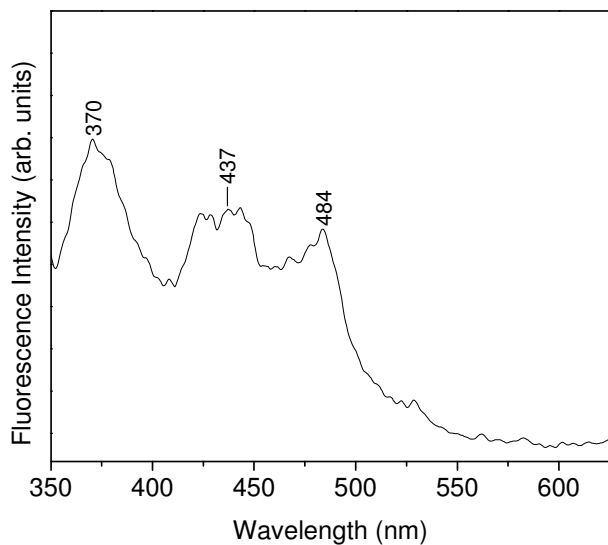


Figure S7. Fluorescence emission spectrum of PLP (106 μM) in Tris buffer (pH 8.5, 100 mM). Excitation wavelength, 330 nm.

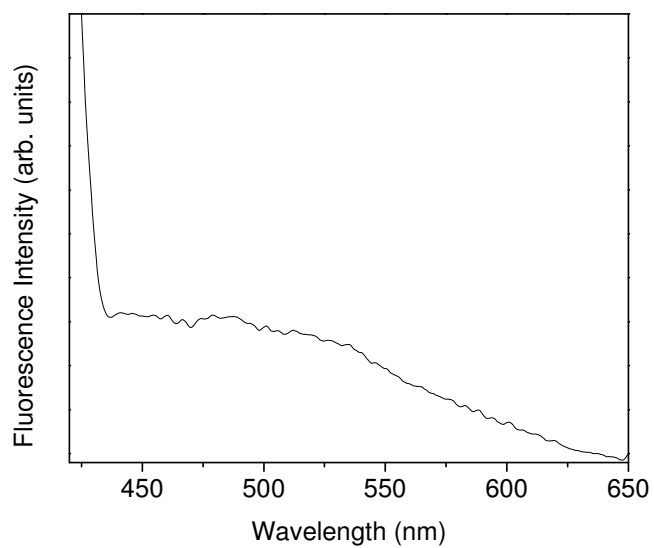


Figure S8. Fluorescence emission spectrum of PLP (50 μM) in phosphate buffer (pH 8.0, 100 mM). Excitation wavelength, 410 nm.

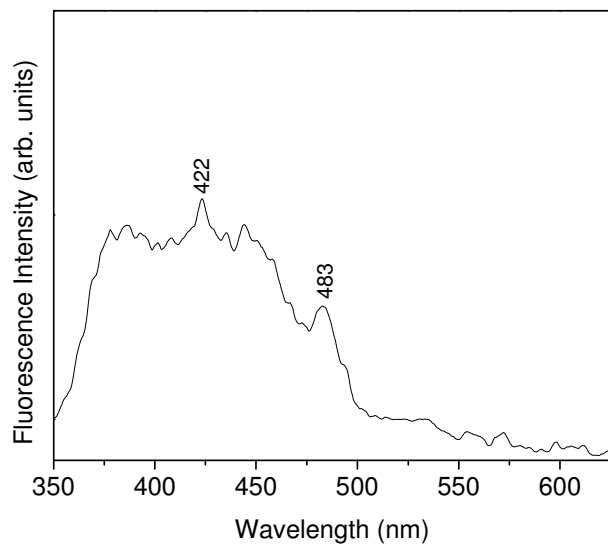


Figure S9. Fluorescence emission spectrum of PLP (50 μM) in phosphate buffer (pH 8.0, 100 mM). Excitation wavelength, 330 nm.

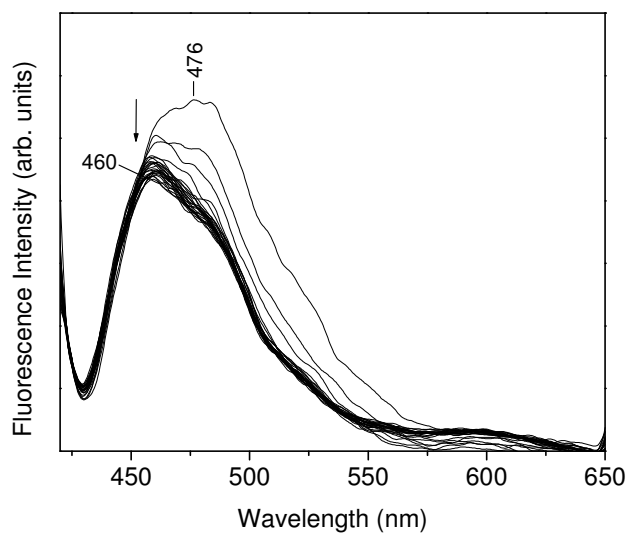


Figure S10. Time dependence of fluorescence spectra after CO binding to Fe(II)hCBS (30 μM) in 5.0 mM phosphate buffer pH 8.0. 410 nm excitation spectra were collected every 10 min for 4 h.

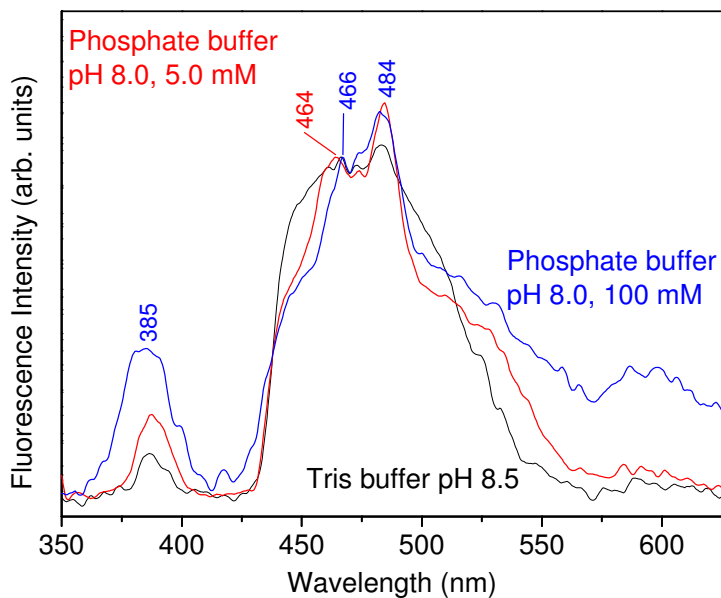


Figure S11. Fluorescence spectra measured 5-6 h after CO binding to Fe(II)hCBS in Tris buffer 100 mM, pH 8.5; phosphate buffer 100 mM, pH 8.0; and phosphate buffer 5.0 mM, pH 8.0. 330 nm excitation.

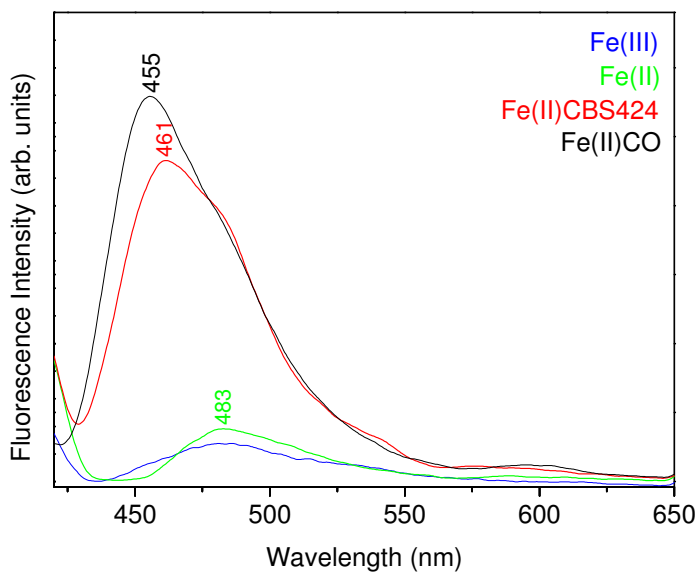


Figure S12. Fluorescence emission spectra of the indicated forms of hCBS in Tris buffer 100mM, pH 8.5. [hCBS] = 48 μ M. 410 nm excitation spectra.

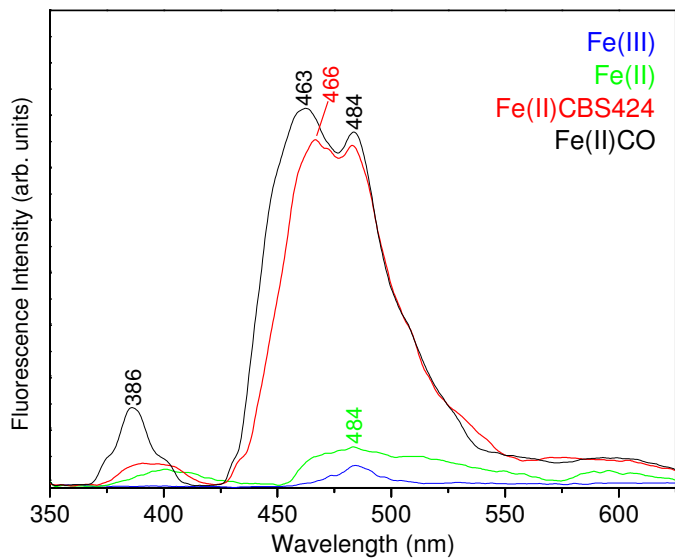


Figure S13. Fluorescence emission spectra of the indicated forms of hCBS in Tris buffer pH 8.5, [hCBS] = 48 μ M. 330 nm excitation spectra.

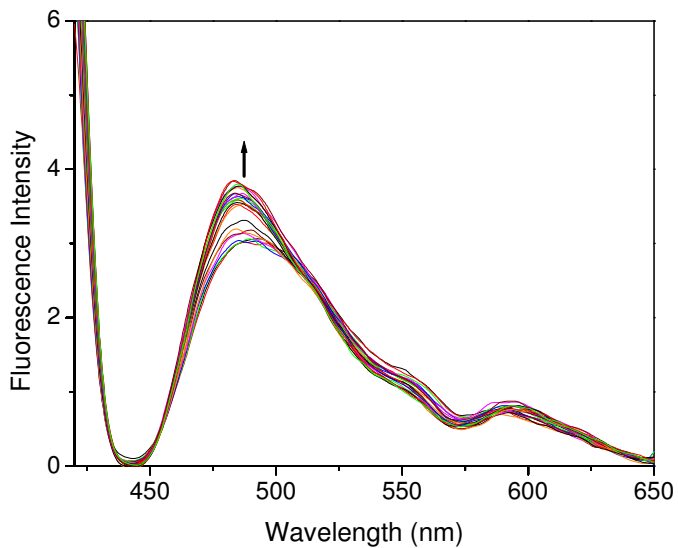


Figure S14. Time dependence of fluorescence emission spectra of Fe(II)hCBS (31 μ M) in Tris buffer pH 8.5. Excitation wavelength: 410 nm. Spectra were collected at 10 min intervals for 4 h.

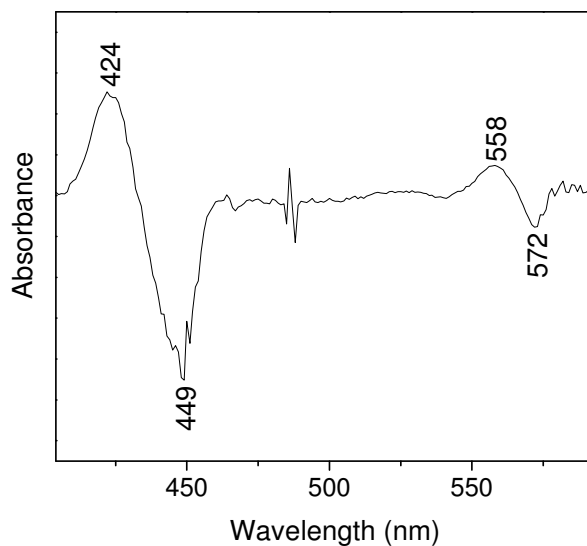


Figure S15. UV-vis absorption difference spectrum of the ferrous hCBS in Tris buffer pH 8.5 of sample used to record the time-dependent fluorescence measurements. The spectrum recorded prior to the fluorescence measurements was subtracted from the spectrum recorded after completion of the fluorescence measurements.

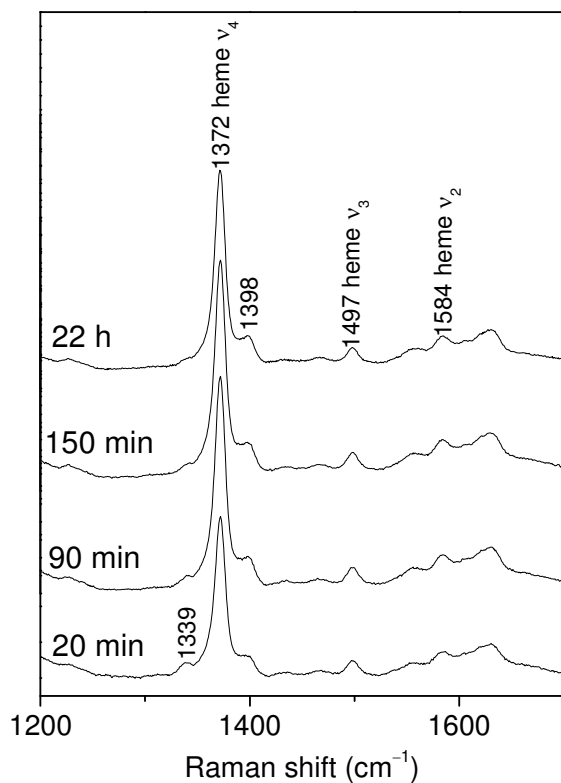


Figure S16. Raman spectra of hCBS-CO formed from Fe(II)hCBS in Tris buffer pH 8.6 at the indicated time points after the start of CO binding. $\lambda_{\text{ex}}=390$ nm. Accumulation time, 10 min.

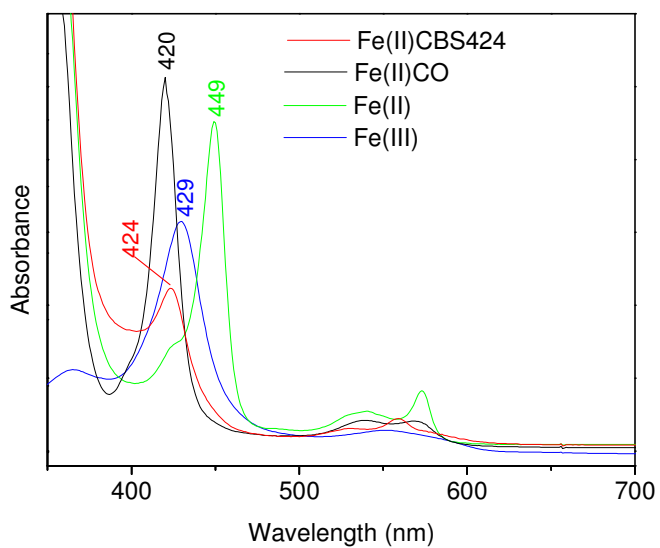


Figure S17. Electronic absorption spectra of the indicated forms of hCBS in Tris buffer pH 8.5.

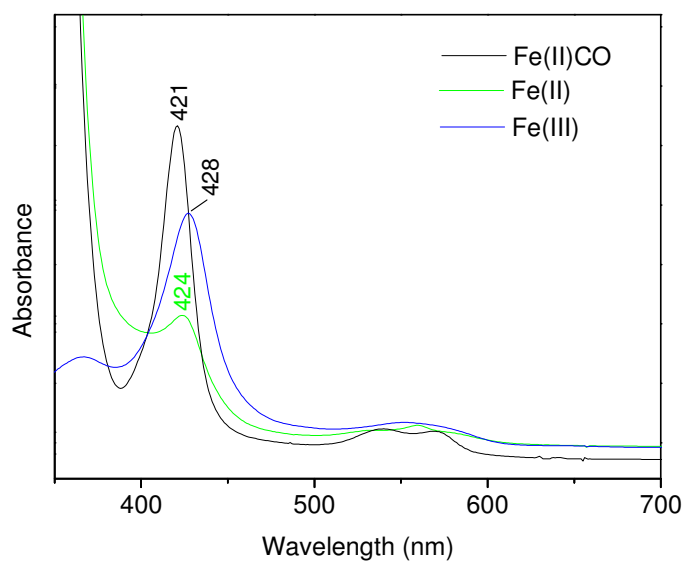


Figure S18. Electronic absorption spectra of the indicated forms of R266M hCBS in phosphate buffer pH 8.0.