An Efficient Process for Pd-Catalyzed C–N Cross-Coupling Reactions of Aryl Iodides: Insight Into Controlling Factors

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Supporting Information

General Reagent Information

All reactions were carried out under an argon atmosphere. The 1,4-dioxane, DME, and tert-butanol were purchased from Aldrich Chemical Co. in Sure-Seal bottles and were used as received. Toluene was purchased from J.T. Baker in CYCLE-TAINER® solventdelivery kegs and vigorously purged with argon for 2 h. The solvent was further purified by passing it under argon pressure through two packed columns of neutral alumina and copper (II) oxide. Aryl halides and amines were purchased from Aldrich Chemical Co., Alfa Aesar, Acros Organics or TCI America. All amines, aryl bromides, and aryl iodides that were liquids were distilled from calcium hydride and stored under argon. Amines and aryl halides that were a solid were used as purchased without further purification. Cesium carbonate (fine) was a gift from Chemetall and sodium tert-butoxide was purchased from Aldrich Chemical Co. The bulk of the bases were stored in an N₂ glovebox. Small portions were taken outside the box in glass vials and weighed in the air. Anhydrous sodium iodide was purchased from Aldrich Chemical Co. and stored in an N₂ glovebox. Ligands $1^{1}_{1} 2^{2}_{2}$ and 4^{3} were synthesized using literature procedures. Ligand 3 was purchased from Strem Chemicals. Precatalysts $5^{1}_{,1}$ 6, 7, and $8^{5}_{,1}$ were synthesized using literature procedures. Flash chromatography was performed using a Biotage SP4 instrument with prepacked silica cartridges.

General Analytical Information

All compounds were characterized by ¹H NMR, ¹³C NMR, IR spectroscopy, as well as, in most instances, elemental analysis. Copies of the ¹H and ¹³C spectra can be found at the end of the Supporting Information. Nuclear Magnetic Resonance spectra were recorded on a Varian 300 MHz instrument and Varian 500 MHz instruments. All ¹H NMR

experiments are reported in δ units, parts per million (ppm), and were measured relative to the signals for residual chloroform (7.26 ppm) in the deuterated solvent, unless otherwise stated. All ¹³C NMR spectra are reported in ppm relative to deuterochloroform (77.23 ppm), unless otherwise stated, and all were obtained with ¹H decoupling. All IR spectra were taken on a Perkin – Elmer 2000 FTIR. All GC analyses were performed on a Agilent 6890 gas chromatograph with an FID detector using a J & W DB-1 column (10 m, 0.1 mm I.D.). Elemental analyses were performed by Atlantic Microlabs Inc., Norcross, GA.

General Procedure for Figure 1

An oven-dried test tube, which was equipped with a magnetic stir bar and fitted with a teflon septum, was charged with the precatalyst (1 mol%), 4-iodoanisole (234 mg, 1.0 mmol), and NaOt-Bu (115 mg, 1.2 mmol). The vessel was evacuated and backfilled with argon (this process was repeated a total of 3 times) and then the aniline (110 μ L, 1.2 mmol) and toluene (1 mL) were added via syringe. The solution was stirred at room temperature for 10 min, diluted with Ethyl acetate, and washed with water. Dodecane was then added as an internal standard and the reaction was analyzed by GC.

General Procedure for Table 1

An oven-dried 16 mL vial was equipped with a magnetic stir bar, fitted with a screw-cap Teflon septum, and taken into the glovebox. Once in the glovebox, the vial was charged with 4-haloanisole (1.0 mmol), aniline (55 μ L, 0.6 mmol), NaOt-Bu (58 mg, 0.6 mmol), and solvent (1.7 mL). The reaction was then taken out of the glovebox and placed in an Omnical CRC reaction calorimeter along with a syringe containing a solution of the Pd source in the solvent (300 μ L solvent, 1 mol% Pd precatalyst). The calorimeter was set to 22.4 °C and allowed to thermally equilibrate. After equilibration, the solution of Pd precatalyst was injected and the reaction was then applied to the raw data due to the delay between the moment that the heat is given off of the reaction and the moment that it is detected. The corrected heat flow curve was then converted to fractional conversion by

dividing the area under the curve to any point by the total area under the curve (equation 1). Dodecane was then added as to the reaction as an internal standard and it was analyzed by GC.

$$Equation 1$$
fractional conversion =
$$\frac{\int_{0}^{t} q \cdot dt}{\int_{0}^{t(f)} q \cdot dt}$$

General Procedure for Figure 2

An oven-dried 16 mL vial was equipped with a magnetic stir bar, fitted with a screw-cap Teflon septum, and taken into the glovebox. Once in the glovebox, the vial was charged with 2 mL of a solution of 4-bromoanisole (63 μ L, 0.5 mmol), aniline (55 μ L, 0.6 mmol), NaO*t*-Bu (58 mg, 0.6 mmol), and NaI (0 – 20 mol%) in DME, which was prepared in a 2 mL volumetric flask. The reaction was then taken out of the glovebox and placed in an Omnical CRC reaction calorimeter along with a syringe containing a solution of **8** (4 mg, 1 mol%) in 1,4-dioxane (300 μ L). The calorimeter was set to 22.4 °C and allowed to thermally equilibrate. After equilibration, the solution containing **8** was injected and the reaction was then applied to the raw data due to the delay between the moment that the heat is given off of the reaction and the moment that it is detected. The corrected heat flow curve was then converted to rate by using the equation q = $\Delta H_{rxn} \cdot V \cdot r$, where q is the heat flow, ΔH_{rxn} is the heat of reaction, V is the reaction volume, and r is the reaction rate. The heat of reaction was found by integrating the heat flow vs. time curves.

General Procedure for Figure 3

An oven-dried 16 mL vial was equipped with a magnetic stir bar, fitted with a screw-cap Teflon septum, and taken into the glovebox. Once in the glovebox, the vial was charged with 4-iodoanisole (117 mg, 0.5 mmol), aniline (55 μ L, 0.6 mmol), NaOt-Bu (58 mg, 0.6

mmol), NaI or tetrahexylammonium iodide (0 – 20 mol%), and toluene (1.7 mL). The reaction was then taken out of the glovebox and placed in an Omnical CRC reaction calorimeter along with a syringe containing a solution of **8** (4 mg, 1 mol%) in toluene (300 μ L). The calorimeter was set to 22.4 °C and allowed to thermally equilibrate. After equilibration, the solution containing **8** was injected and the reaction was stirred until the heat flow on the calorimeter returned to the baseline. A correction was then applied to the raw data due to the delay between the moment that the heat is given off of the reaction and the moment that it is detected. The corrected heat flow curve was then converted to rate by using the equation $q = \Delta H_{rxn} \cdot V \cdot r$, where q is the heat flow, ΔH_{rxn} is the heat of reaction, V is the reaction volume, and r is the reaction rate. The heat of reaction was found by integrating the heat flow vs. time curves.

General Procedure for Figure 4

An oven-dried test tube, which was equipped with a magnetic stir bar and fitted with a teflon septum, was charged with the precatalyst (1 mol%), 4-iodoanisole (234 mg, 1.0 mmol), and NaOt-Bu (115 mg, 1.2 mmol). The vessel was evacuated and backfilled with argon (this process was repeated a total of 3 times) and then the morpholine (105 μ L, 1.2 mmol) and toluene (1 mL) were added via syringe. The solution was heated to 80 °C for 3 min, then cooled to room temperature, diluted with Ethyl acetate, and washed with water. Dodecane was then added as an internal standard and the reaction was analyzed by GC.

General Procedures for Tables 2, 3, and 4

General Procedure A: An oven-dried test tube, which was equipped with a magnetic stir bar and fitted with a teflon septum, was charged with NaO*t*-Bu (1.2 equiv). The vessel was evacuated and backfilled with argon (this process was repeated a total of 3 times) and then the aryl iodide (1 equiv), amine (1.2 equiv), and toluene (0.3 - 1.0 mL/mmol) were added via syringe (aryl iodides or amines that were solids at room temperature were added with the and base). A solution of the ligand and precatalyst in toluene (0.002 M, 0.005 - 0.2 mol% **1**, 0.005 - 0.2 mol% **5**) was added and the reaction was heated to 110

°C until the starting material was completely consumed as monitored by GC. The reaction mixture was then cooled to room temperature, diluted with ethyl acetate, washed with water, concentrated in vacuo, and purified via the Biotage SP4 (silica-packed 50 or100 g snap cartridge).

General Procedure B: An oven-dried test tube, which was equipped with a magnetic stir bar and fitted with a teflon septum, was charged with the ligand (0.05 - 0.2 mol%), the precatalyst (0.05 - 0.2 mol%), and NaOt-Bu (1.2 - 1.4 equiv). The vessel was evacuated and backfilled with argon (this process was repeated a total of 3 times) and then the aryl iodide (1 equiv), amine (1.2 - 1.4 equiv), and toluene (0.3 mL/mmol) were added via syringe (aryl iodides or amines that were solids at room temperature were added with the precatalyst and base). The solution was heated to 85 °C until the starting material was completely consumed as monitored by GC. The reaction mixture was then cooled to room temperature, diluted with ethyl acetate, washed with water, concentrated in vacuo, and purified via the Biotage SP4 (silica-packed 50 or100 g snap cartridge).

General Procedure C: General procedure A was used with the following modification: 1.4 equiv of Cs_2CO_3 was used as the base.

General Procedure D: An oven-dried test tube, which was equipped with a magnetic stir bar and fitted with a teflon septum, was charged with the ligand (0.5 - 1 mol%), the precatalyst (0.5 - 1 mol%), and K_2CO_3 (1.4 equiv). The vessel was evacuated and backfilled with argon (this process was repeated a total of 3 times) and then the aryl iodide (1 equiv), amine (1.4 equiv), and *t*-BuOH (2 mL/mmol) were added via syringe (aryl iodides or amines that were solids at room temperature were added with the precatalyst and base). The solution was heated to 110 °C until the starting material was completely consumed as monitored by GC. The reaction mixture was then cooled to room temperature, diluted with ethyl acetate, washed with water, concentrated in vacuo, and purified via the Biotage SP4 (silica-packed 50 g snap cartridge).



4-methoxy-*N***-phenylaniline**⁴ (Table 2) Following general procedure A, a mixture of 4-iodoanisole (468 mg, 2.0 mmol), aniline (256 μ L, 2.8 mmol), NaO*t*-Bu (269 mg, 2.8 mmol), **5** and **1** (50 μ L, 0.002 M in toluene, 0.005 mol%), and toluene (2 mL) was heated to 110 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 50 g snap column; 0-45% EtOAc/hexanes) to provide the title compound as a white solid (390 mg, 98%), mp 104 – 105 °C (lit. 104 – 106 °C). ¹H NMR (300 MHz, CDCl₃) δ : 7.31 (t, *J* = 7.5 Hz, 2H), 7.15 (d, *J* = 9.0 Hz, 2H), 6.97 (m, 5H), 5.58 (bs, 1H), 3.87 (s, 3H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ : 155.5, 145.5, 136.1, 129.7, 122.5, 119.9, 115.9, 115.0, 55.9 ppm. IR (neat, cm⁻¹): 3384, 2837, 1597, 1513, 1298, 1250, 1182, 1034, 752, 696. Anal. Calcd. for C₁₃H₁₃NO: C, 78.36; H, 6.58. Found: C, 78.07; H, 6.62.



4-methoxy-*N***-phenylaniline**⁴ (Table 2) Following general procedure A, a mixture of 4-iodoanisole (468 mg, 2.0 mmol), aniline (256 μL, 2.8 mmol), NaO*t*-Bu (269 mg, 2.8 mmol), **5** and **1** (100 μL, 0.002 M in toluene, 0.01 mol%), and toluene (900 μL) was heated to 110 °C for 5 min. The crude product was purified via the Biotage SP4 (silica-packed 50 g snap column; 0-45% EtOAc/hexanes) to provide the title compound as a white solid (388 mg, 97%), mp 104 – 105 °C (lit. 104 – 106 °C). ¹H NMR (300 MHz, CDCl₃) δ: 7.31 (t, *J* = 7.5 Hz, 2H), 7.15 (d, *J* = 9.0 Hz, 2H), 6.97 (m, 5H), 5.58 (bs, 1H), 3.87 (s, 3H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ: 155.5, 145.5, 136.1, 129.7, 122.5, 119.9, 115.9, 115.0, 55.9 ppm. IR (neat, cm⁻¹): 3384, 2837, 1597, 1513, 1298, 1250, 1182, 1034, 752, 696. Anal. Calcd. for C₁₃H₁₃NO: C, 78.36; H, 6.58. Found: C, 78.38; H, 6.68.



2-methyl-*N***-(3-(trifluoromethyl)phenyl)aniline** (Table 2) Following general procedure A, a mixture of 2-iodotoluene (350 μ L, 2.0 mmol), 3-aminobenzotrifluoride (350 μ L, 2.8 mmol), NaO*t*-Bu (269 mg, 2.8 mmol), **5** and **1** (100 μ L, 0.002 M in toluene, 0.01 mol%), and toluene (900 μ L) was heated to 110 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 50 g snap column; 0-45% EtOAc/hexanes) to provide the title compound as a yellow oil (438 mg, 87%). ¹H NMR (300 MHz, CDCl₃) δ : 7.42 – 7.28 (m, 4H), 7.23 – 7.10 (m, 4H), 5.58 (bs, 1H), 2.35 (s, 3H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ : 145.4, 140.1, 132.1, 131.9, 131.6, 130.5, 130.1, 127.3, 125.7, 124.0, 123.5, 121.2, 119.4, 116.6, 116.5, 116.5, 113.0, 112.9, 112.9, 18.1 ppm (observed complexity due to C–F splitting). IR (neat, cm⁻¹): 3398, 3034, 2926, 1585, 1496, 1338, 1165, 1124, 1069, 699. Anal. Calcd. for C₁₄H₁₂F₃N: C, 66.93; H, 4.81. Found: C, 66.98; H, 4.80.



4-chloro-*N***-phenylaniline**⁵ (Table 2) Following general procedure A, a mixture of 4chloroiodobenzene (477 mg, 2.0 mmol), aniline (182 µL, 2.0 mmol), NaO*t*-Bu (269 mg, 2.8 mmol), **5** and **1** (500 µL, 0.002 M in toluene, 0.05 mol%), and toluene (500 µL) was heated to 80 °C for 2 h. The crude product was purified via the Biotage SP4 (silicapacked 50 g snap column; 0-25% EtOAc/hexanes) to provide the title compound as a white solid (397 mg, 97%), mp 67 – 68 °C (lit. 72 °C). ¹H NMR (500 MHz, CDCl₃) δ: 7.38 (t, *J* = 7.5 Hz, 2H), 7.29 (d, *J* = 6.5 Hz, 2H), 7.12 (d, *J* = 7.5 Hz, 2H), 7.05 (m, 3H), 5.70 (bs, 1H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ: 143.0, 142.2, 129.8, 129.6, 125.7, 121.8, 119.1, 118.4 ppm. IR (neat, cm⁻¹): 3403, 1590, 1505, 1485, 1384, 1312, 1090, 750, 692, 504. Anal. Calcd. for C₁₂H₁₀ClN: C, 70.77; H, 4.95. Found: C, 70.57; H, 5.03.



3-chloro-*N***-phenylaniline** (Table 2) Following general procedure A, a mixture of 3chloroiodobenzene (247 μ L, 2.0 mmol), aniline (182 μ L, 2.0 mmol), NaO*t*-Bu (269 mg, 2.8 mmol), **5** and **1** (500 μ L, 0.002 M in toluene, 0.05 mol%), and toluene (500 μ L) was heated to 80 °C for 2 h. The crude product was purified via the Biotage SP4 (silicapacked 50 g snap column; 0-25% EtOAc/hexanes) to provide the title compound as a yellow oil (395 mg, 96%). ¹H NMR (500 MHz, CDCl₃) δ : 7.38 (t, *J* = 7.5 Hz, 2H), 7.22 (t, *J* = 8.0 Hz, 1H), 7.15 (d, *J* = 7.5 Hz, 2H), 7.09 (m, 2H), 6.95 (dd, *J* = 8.0 Hz, *J* = 2.0 Hz, 2H), 5.73 (bs, 1H) ppm. ¹³C NMR (125 MHz, CDCl₃) δ : 145.1, 142.2, 135.3, 130.7, 129.8, 122.4, 120.8, 119.3, 117.0, 115.4 ppm. IR (neat, cm⁻¹): 3403, 3059, 1589, 1496, 1480, 1314, 993, 916, 752, 681. Anal. Calcd. for C₁₂H₁₀ClN: C, 70.77; H, 4.95. Found: C, 71.05; H, 4.91.



N-benzyl-4-methoxyaniline¹ (Table 2) Following general procedure B, a mixture of 4iodoanisole (2.33 g, 10 mmol), benzylamine (1.31 mL, 12 mmol), NaOt-Bu (1.34 g, 14 mmol), **5** (4 mg, 0.05 mol%) and **1** (2.5 mg, 0.05 mol%), and toluene (3 mL) was heated to 85 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 100 g snap column; 0-40% EtOAc/hexanes) to provide the title compound as a yellow oil (1.761 g, 83%). ¹H NMR (300 MHz, CDCl₃) δ : 7.46 (m, 5H), 6.91 (d, *J* = 9.0 Hz, 2H), 6.71 (d, *J* = 9.0 Hz, 2H), 4.37 (s, 2H), 3.90 (bs, 1H), 3.84 (s, 3H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ : 152.5, 142.9, 140.2, 129.0, 127.9, 127.5, 115.3, 114.5, 56.1, 49.5 ppm. IR (neat, cm⁻¹): 3415, 3028, 2831, 1511, 1452, 1234, 1036, 819, 742, 697.



N-cyclohexyl-4-methoxyaniline⁶ (Table 2) Following general procedure B, a mixture of 4-iodoanisole (2.33 g, 10 mmol), cyclohexylamine (1.603 mL, 14 mmol), NaO*t*-Bu (1.34 g, 14 mmol), **5** (8 mg, 0.1 mol%) and **1** (5 mg, 0.1 mol%), and toluene (3 mL) was heated to 85 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 100 g snap column; 0-50% EtOAc/hexanes) to provide the title compound as a white solid (1.875 g, 92%), mp 44 – 46 °C. ¹H NMR (300 MHz, CDCl₃) δ : 6.80 (d, *J* = 9.0 Hz, 2H), 6.60 (*J* = 9.0 Hz, 2H), 3.76 (s, 3H), 3.26 (bs, 1H), 3.20 (m, 1H), 2.07 (m, 2H), 1.78 (m, 2H), 1.68 (m, 1H), 1.48 – 1.05 (m, 5H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ : 152.1, 141.9, 115.1, 115.0, 56.0, 53.0, 33.9, 26.3, 25.4 ppm. IR (neat, cm⁻¹): 3388, 2929, 2852, 1511, 1450, 1240, 1040, 819, 756, 515.



2-(phenylamino)benzonitrile (Table 2) Following general procedure C, a mixture of iodobenzene (112 µL, 1.0 mmol), 2-aminobenzonitrile (165 mg, 1.2 mmol), Cs₂CO₃ (456 mg, 1.4 mmol), and **5** and **1** (1 mL, 0.002 M in toluene, 0.2 mol%) was heated to 110 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 50 g snap column; 0-45% EtOAc/hexanes) to provide the title compound as a white solid (175 mg, 90%), mp 50 – 51 °C. ¹H NMR (300 MHz, CDCl₃) δ : 7.51 (d, *J* = 7.8 Hz, 1H), 7.39 (m, 3H), 7.21 (m, 3H), 7.15 (t, *J* = 7.2 Hz, 1H), 6.85 (t, *J* = 7.5 Hz, 1H), 6.54 (bs, 1H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ : 147.6, 140.3, 134.2, 133.4, 129.9, 124.4, 121.9, 119.5, 118.0, 114.5, 98.8 ppm. IR (neat, cm⁻¹): 3338, 2217, 1592, 1575, 1517, 1457, 1319, 1293, 745, 695. Anal. Calcd. for C₁₃H₁₀N₂: C, 80.39; H, 5.19. Found: C, 80.09; H, 5.12.



Ethyl 4-(3,5-dimethylphenylamino)benzoate⁷ (Table 2) Following general procedure C, a mixture of 3,5-dimethyliodobenzene (144 μL, 1.0 mmol), ethyl 4-aminobenzoate (231 mg, 1.4 mmol), Cs₂CO₃ (456 mg, 1.4 mmol), **5** and **1** (500 μL, 0.002 M in toluene, 0.1 mol%), and toluene (0.5 mL) was heated to 110 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 50 g snap column; 0-50% EtOAc/hexanes) to provide the title compound as a white solid (244 mg, 91%), mp 117 – 119 °C (lit. 119 °C). ¹H NMR (300 MHz, CDCl₃) δ: 7.94 (d, *J* = 8.4 Hz, 2H), 6.99 (d, *J* = 8.4, 2H), 6.81 (s, 2H), 6.73 (s, 1H), 6.12 (bs, 1H), 4.36 (q, *J* = 7.2 Hz, 2H), 2.32 (s, 6H), 1.40 (t, *J* = 7.2 Hz, 3H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ: 166.9, 148.5, 141.1, 139.4, 131.7, 125.0, 121.3, 118.3, 114.9, 60.7, 21.6, 14.7 ppm. IR (neat, cm⁻¹): 3343, 1698, 1595, 1510, 1352, 1282, 1170, 1109, 831, 769. Anal. Calcd. for C₁₇H₁₉NO₂: C, 75.81; H, 7.11. Found: C, 75.54; H, 7.13.



4-methyl-*N***-(4-nitrophenyl)aniline** (Table 2) Following general procedure C, a mixture of 4-iodotoluene (218 mg, 1.0 mmol), 4-nitroaniline (193 mg, 1.4 mmol), Cs₂CO₃ (456 mg, 1.4 mmol), **5** and **1** (500 µL, 0.002 M in toluene, 0.1 mol%), and toluene (0.5 mL) was heated to 110 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 50 g snap column; 0-40% EtOAc/hexanes) to provide the title compound as an orange solid (220 mg, 96%), mp 138 – 139 °C. ¹H NMR (300 MHz, CDCl₃) δ : 8.09 (d, *J* = 9.0 Hz, 2H), 7.20 (d, *J* = 7.8 Hz, 2H), 7.12 (d, *J* = 7.8 Hz, 2H), 6.88 (d, *J* = 9.0 Hz, 2H), 6.38 (bs, 1H), 2.37 (s, 3H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ : 151.2, 139.4, 136.9, 135.0, 130.5, 126.6, 122.9, 113.4, 21.2 ppm. IR (neat, cm⁻¹): 3342, 1595, 1524, 1481, 1298, 1187, 1111, 829, 749, 499. Anal. Calcd. for C₁₃H₁₂N₂O₂: C, 68.41; H, 5.30. Found: C, 68.35; H, 5.22.

Experimental Procedures for Examples Described in Table 3



4-phenylmorpholine⁸ (Table 3) Following general procedure A, a mixture of iodobenzene (1.119 mL, 10 mmol), morpholine (1.223 mL, 14 mmol), NaO*t*-Bu (1.34 g, 14 mmol), **4** and **8** (1.25 mL, 0.002 M in toluene, 0.025 mol%), and toluene (1.75 mL) was heated to 85 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 100 g snap column; 0-50% EtOAc/hexanes) to provide the title compound as a white solid (1.632 g, 99%), mp 53 – 55 °C. ¹H NMR (500 MHz, CDCl₃) δ : 7.39 (t, *J* = 7.0 Hz, 2H), 7.00 (m, 3H), 3.91 (t, *J* = 5.0 Hz, 4H), 3.19 (t, *J* = 5.0 Hz, 4H) ppm. ¹³C NMR (125 MHz, CDCl₃) δ : 151.7, 129.6, 120.3, 116.0, 67.2, 49.6 ppm. IR (neat, cm⁻¹): 2888, 2856, 2826, 1599, 1496, 1449, 1231, 1120, 926, 773. Anal. Calcd. for C₁₀H₁₃NO: C, 73.59; H, 8.03. Found: C, 73.53; H, 8.19.



1-(4-methoxyphenyl)piperidine⁸ (Table 3) Following general procedure B, a mixture of 4-iodoanisole (2.33 g, 10 mmol), piperidine (1.381 mL, 14 mmol), NaOt-Bu (1.34 g, 14 mmol), **8** (3.5 mg, 0.05 mol%) and **4** (2.5 mg, 0.05 mol%), and toluene (3 mL) was heated to 85 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 100 g snap column; 0-40% EtOAc/hexanes) to provide the title compound as a

yellow oil (1.633 g, 85%). ¹H NMR (500 MHz, CDCl₃) δ : 6.96 (d, J = 9.0 Hz, 2H), 6.88 (d, J = 9.0 Hz, 2H), 3.79 (s, 3H), 3.07 (t, J = 5.5 Hz, 4H), 1.77 (m, 4H), 1.59 (m, 2H) ppm. ¹³C NMR (125 MHz, CDCl₃) δ : 153.8, 147.2, 119.0, 114.6, 55.7, 52.6, 26.5, 24.5 ppm. IR (neat, cm⁻¹): 2934, 2792, 1511, 1453, 1244, 1041, 919, 823, 700, 539.



1-(4-methoxyphenyl)piperidine⁸ (Table 3) Following general procedure B, a mixture of 4-iodoanisole (2.33 g, 10 mmol), piperidine (1.381 mL, 14 mmol), NaO*t*-Bu (1.34 g, 14 mmol), **8** (3.5 mg, 0.05 mol%) and **4** (2.5 mg, 0.05 mol%), and dioxane (3 mL) was heated to 85 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 100 g snap column; 0-40% EtOAc/hexanes) to provide the title compound as a yellow oil (1.711 g, 89%). ¹H NMR (500 MHz, CDCl₃) δ : 6.96 (d, *J* = 9.0 Hz, 2H), 6.88 (d, *J* = 9.0 Hz, 2H), 3.79 (s, 3H), 3.07 (t, *J* = 5.5 Hz, 4H), 1.77 (m, 4H), 1.59 (m, 2H) ppm. ¹³C NMR (125 MHz, CDCl₃) δ : 153.8, 147.2, 119.0, 114.6, 55.7, 52.6, 26.5, 24.5 ppm. IR (neat, cm⁻¹): 2934, 2792, 1511, 1453, 1244, 1041, 919, 823, 700, 539.



N,*N*-dibutyl-3,5-dimethylaniline (Table 3) Following general procedure A, a mixture of 3,5-dimethyliodobenzene (1.44 mL, 10 mmol), dibutylamine (2.35 mL, 14 mmol), NaO*t*-Bu (1.34 g, 14 mmol), **4** and **8** (2.50 mL, 0.002 M in toluene, 0.05 mol%), and toluene (0.5 mL) was heated to 85 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 100 g snap column; 0-50% EtOAc/hexanes) to provide the title compound as a yellow oil (2.127 g, 91%). ¹H NMR (300 MHz, CDCl₃) δ : 6.66 (s, 3H), 3.60 (t, *J* = 7.5 Hz, 4H), 2.64 (s, 6H), 1.93 (pentet, *J* = 8.1 Hz, 4H), 1.72 (sextet, *J* = 7.5 Hz, 4H), 1.33 (t, *J* = 7.5 Hz, 6H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ : 148.9, 139.1, 117.9, 110.3, 51.4, 30.1, 22.5, 21.0, 14.6 ppm. IR (neat, cm⁻¹): 2957, 2872, 1597, 1486, 1367, 1304, 1196, 1064, 814, 691. Anal. Calcd. for C₁₆H₂₇N: C, 82.34; H, 11.66. Found: C, 82.17; H, 11.91.



1-(3,5-dimethylphenyl)-4-methylpiperazine (Table 3) Following general procedure B, a mixture of 3.5-dimethyliodobenzene (1.443 mL, 10 mmol), *N*-methylpiperazine (1.550 mL, 14 mmol), NaOt-Bu (1.34 g, 14 mmol), **8** (7 mg, 0.1 mol%) and **4** (4.5 mg, 0.1 mol%), and toluene (3 mL) was heated to 85 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 100 g snap column; 75-100% EtOAc/hexanes) to provide the title compound as a yellow oil (1.808 g, 89%). ¹H NMR (300 MHz, CDCl₃) δ : 6.61 (s, 2H), 6.56 (s, 1H), 3.22 (t, *J* = 5.1 Hz, 4H), 2.59 (t, *J* = 5.1 Hz, 4H), 2.38 (s, 3H), 2.32 (s, 6H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ : 151.7, 138.8, 121.9, 114.3, 55.5, 49.5, 46.5, 22.0 ppm. IR (neat, cm⁻¹): 2937, 2794, 1597, 1452, 1376, 1291, 1265, 1143, 1009, 828.



4-methoxy-*N***-methyl-***N***-phenylaniline** (Table 3) Following general procedure A, a mixture of 4-iodoanisole (468 mg, 2.0 mmol), *N*-methylaniline (303 µL, 2.8 mmol), NaO*t*-Bu (269 mg, 2.8 mmol), **4** and **8** (100 µL, 0.002 M in toluene, 0.01 mol%), and toluene (0.9 mL) was heated to 85 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 50 g snap column; 0-40% EtOAc/hexanes) to provide the title compound as a yellow oil (342 mg, 80%). ¹H NMR (500 MHz, CDCl₃) δ : 7.25 (t, *J* = 9.0 Hz, 2H), 7.15 (d, *J* = 9.0 Hz, 2H), 6.95 (d, *J* = 9.5 Hz, 2H), 6.85 (m, 3H), 3.86 (s, 3H), 3.31 (s, 3H) ppm. ¹³C NMR (125 MHz, CDCl₃) δ : 156.6, 150.0, 142.5, 129.2, 126.6, 118.6, 116.0, 115.0, 55.8, 40.8, 39.8 ppm. IR (neat, cm⁻¹): 2951, 2834, 1597, 1508, 1344, 1244, 1035, 835, 751, 695. Anal. Calcd. for C₁₄H₁₅NO: C, 78.84; H, 7.09. Found: C, 79.07; H, 7.24.



3,5-dimethyl-N,N-diphenylaniline (Table 3) Following general procedure A, a mixture

of 3.5-dimethyliodobenzene (289 µL, 2.0 mmol), diphenylamine (473 mg, 2.8 mmol), NaOt-Bu (269 mg, 2.8 mmol), **4** and **8** (500 µL, 0.002 M in toluene, 0.05 mol%), and toluene (0.5 mL) was heated to 110 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 50 g snap column; 0-40% EtOAc/hexanes) to provide the title compound as a white solid (489 mg, 90%), mp 135 – 136 °C. ¹H NMR (300 MHz, CDCl₃) δ : 7.34 (t, *J* = 7.2 Hz, 4H), 7.20 (*J* = 7.5 Hz, 4H), 7.09 (t, *J* = 7.2 Hz, 2H), 6.86 (s, 2H), 6.80 (s, 1H), 2.34 (s, 6H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ : 148.4, 148.1, 139.2, 129.5, 125.2, 125.2, 124.4, 122.7, 21.7 ppm. IR (neat, cm⁻¹): 3035, 1589, 1492, 1338, 1294, 1273, 1230, 1029, 753, 693. Anal. Calcd. for C₂₀H₁₉N: C, 87.87; H, 7.01. Found: C, 87.62; H, 7.11.



3,5-dimethyl-*N*,*N***-diphenylaniline** (Table 3) Following general procedure C, a mixture of ethyl 4-iodobenzoate (152 μ L, 1.0 mmol), 4-cyano-*N*-methylaniline (185 mg, 1.4 mmol), Cs₂CO₃ (456 mg, 1.4 mmol), **4** and **8** (500 μ L, 0.002 M in toluene, 0.1 mol%), and toluene (0.5 mL) was heated to 110 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 50 g snap column; 0-30% EtOAc/hexanes) to provide the title compound as a clear oil (266 mg, 95%). ¹H NMR (300 MHz, CDCl₃) δ : 7.99 (d, *J* = 8.4 Hz, 2H), 7.45 (d, *J* = 9.0 Hz, 2H), 7.15 (d, *J* = 8.7 Hz, 2H), 6.96 (d, *J* = 8.7 Hz, 2H), 4.33 (q, *J* = 6.9 Hz, 2H), 3.38 (s, 3H), 1.36 (t, *J* = 7.2 Hz, 3H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ : 166.2, 151.4, 151.0, 133.6, 131.5, 125.9, 122.6, 119.9, 118.0, 102.7, 61.1, 40.2, 14.6 ppm. IR (neat, cm⁻¹): 2982, 2218, 1710, 1595, 1508, 1347, 1278, 1179, 1107, 773. Anal. Calcd. for C₁₇H₁₆N₂O₂: C, 72.84; H, 5.75. Found: C, 72.90; H, 5.76.

Experimental Procedures for Examples Described in Table 4

Note: All heteroaryliodides that were not commercially available were synthesized using literature procedures.⁹



N-p-tolylpyrazin-2-amine (Table 4) Following general procedure D, a mixture of 4-

iodotoluene (218 mg, 1.0 mmol), aminopyrazine (133 mg, 1.4 mmol), K₂CO₃ (197 mg, 1.4 mmol), **5** (4 mg, 0.5 mol%) and **1** (2.5 mg, 0.5 mol%), and *t*-BuOH (2 mL) was heated to 110 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 50 g snap column; 0-50% EtOAc/hexanes) to provide the title compound as a white solid (167 mg, 95%), mp 112 – 113 °C. ¹H NMR (300 MHz, CDCl₃) δ : 8.19 (s, 1H), 8.06 (s, 1H), 7.92 (d, *J* = 2.7 Hz, 1H), 7.62 (bs, 1H), 7.30 (d, *J* = 8.4 Hz, 2H), 7.14 (*J* = 8.1 Hz, 2H), 2.33 (s, 3H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ : 153.2, 142.2, 136.9, 134.3, 133.6, 133.0, 130.2, 121.2, 21.1 ppm. IR (neat, cm⁻¹): 3289, 3100, 1626, 1524, 1385, 1142, 1004, 814, 506, 413. Anal. Calcd. for C₁₁H₁₁N₃: C, 71.33; H, 5.99. Found: C 71.06; H, 6.03.



N-(3,5-dimethylphenyl)pyrimidin-2-amine (Table 4) Following general procedure D, a mixture of 3,5-dimethyliodobenzene (144 μ L, 1.0 mmol), 2-aminopyrimidine (133 mg, 1.4 mmol), K₂CO₃ (197 mg, 1.4 mmol), **5** (8 mg, 1 mol%) and **1** (5 mg, 1 mol%), and *t*-BuOH (2 mL) was heated to 110 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 50 g snap column; 0-40% EtOAc/hexanes) to provide the title compound as a yellow oil (161 mg, 81%). ¹H NMR (300 MHz, CDCl₃) δ : 8.44 (d, *J* = 4.8 Hz, 2H), 8.26 (s, 1H), 7.28 (s, 2H), 6.74 (s, 1H), 6.69 (t, *J* = 4.8 Hz, 1H), 2.53 (s, 6H) ppm. ¹³C NMR (75 MHz, CDCl₃) δ : 160.8, 158.3, 139.6, 138.8, 125.0, 118.0, 112.3, 21.8 ppm. IR (neat, cm⁻¹): 3274, 3014, 1580, 1539, 1449, 1404, 1246, 838, 797, 631. Anal. Calcd. for C₁₂H₁₃N₃: C, 72.33; H, 6.58. Found: C 72.23; H, 6.65.



N-(**pyridin-3-yl**)**pyrazin-2-amine** (Table 4) Following general procedure D, a mixture of 3-iodopyridine (205 mg, 1.0 mmol), aminopyrazine (133 mg, 1.4 mmol), K_2CO_3 (197 mg, 1.4 mmol), **5** (8 mg, 1 mol%) and **1** (5 mg, 1 mol%), and *t*-BuOH (2 mL) was heated to 110 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 50 g snap column; 0-10% MeOH/EtOAc) to provide the title compound as a white solid

(160 mg, 93%), mp 181 – 182 °C. ¹H NMR (300 MHz, DMSO) δ : 9.70 (s, 1H), 8.61 (s, 1H), 8.25 (s, 1H), 8.15 (m, 3H), 7.95 (s, 1H), 7.30 (m, 1H) ppm. ¹³C NMR (75 MHz, DMSO) δ : 152.7, 142.8, 141.7, 140.8, 138.0, 135.8, 134.9, 125.3, 124.2 ppm. IR (neat, cm⁻¹): 3305, 2317, 1643, 1585, 1528, 1485, 1462, 1359, 1006, 702. Anal. Calcd. for C₉H₈N₄: C, 62.78; H, 4.68. Found: C 62.54; H, 4.72.



N-(pyridin-4-yl)pyrazin-2-amine (Table 4) Following general procedure D, a mixture of iodopyrazine (98 μL, 1.0 mmol), 4-amnopyridine (132 mg, 1.4 mmol), K₂CO₃ (197 mg, 1.4 mmol), **5** (8 mg, 1 mol%) and **1** (5 mg, 1 mol%), and *t*-BuOH (2 mL) was heated to 110 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 50 g snap column; 0-10% MeOH/EtOAc) to provide the title compound as a brown solid (155 mg, 90%), decomposed above 200 °C. ¹H NMR (500 MHz, DMSO) δ: 9.97 (s, 1H), 8.33 (m, 3H), 8.23 (s, 1H), 8.06 (s, 1H), 7.66 (m, 2H) ppm. ¹³C NMR (125 MHz, DMSO) δ: 152.2, 150.7, 147.8, 141.8, 136.3, 136.0, 112.7 ppm. IR (neat, cm⁻¹): 3444, 1617, 1522, 1425, 1400, 1356, 1145, 1008, 815, 517.



N-(**pyrazin-2-yl**)**isoquinolin-4-amine** (Table 4) Following general procedure D, a mixture of 4-iodoisoquinoline (128 mg, 0.5 mmol), aminopyrazine (67 mg, 0.7 mmol), K₂CO₃ (99 mg, 0.7 mmol), **5** (4 mg, 1 mol%) and **1** (2.5 mg, 1 mol%), and *t*-BuOH (1 mL) was heated to 110 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 50 g snap column; 0-10% MeOH/EtOAc) to provide the title compound as a white solid (88 mg, 80%), mp 214 – 216 °C. ¹H NMR (300 MHz, DMSO) δ : 9.42 (s, 1H), 9.07 (s, 1H), 8.97 (s, 1H), 8.41 (s, 1H), 8.14 (t, *J* = 8.4 Hz, 2H), 8.05 (s, 1H), 7.95 (s, 1H), 7.77 (t, *J* = 7.2 Hz, 1H), 7.69 (t, *J* = 7.2 Hz, 1H) ppm. ¹³C NMR (75 MHz, DMSO) δ : 153.9, 148.3, 141.9, 137.0, 135.3, 135.0, 131.1, 130.7, 130.6, 129.3, 128.5, 128.1, 122.6 ppm. IR (neat, cm⁻¹): 3391, 1628, 1580, 1521, 1477, 1384, 784, 762, 446, 436.



N-(**pyridin-2-yl)quinolin-3-amine** (Table 4) Following general procedure D, a mixture of 3-iodoquinoline (128 mg, 0.5 mmol), 2-aminopyridine (66 mg, 0.7 mmol), K_2CO_3 (99 mg, 0.7 mmol), **5** (4 mg, 1 mol%) and **1** (2.5 mg, 1 mol%), and *t*-BuOH (1 mL) was heated to 110 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 50 g snap column; 0-10% MeOH/EtOAc) to provide the title compound as a white solid (80 mg, 73%), mp 224 – 226 °C. ¹H NMR (300 MHz, DMSO) δ : 9.60 (bs, 1H), 8.92 (s, 2H), 8.27 (s, 1H), 7.86 (m, 1H), 7.64 (t, *J* = 7.2 Hz, 1H), 7.50 (m, 2H), 6.96 (d, *J* = 8.1 Hz, 1H), 6.84 (t, *J* = 6.0 Hz, 1H) ppm. ¹³C NMR (75 MHz, DMSO) δ : 156.2, 148.0, 145.6, 143.4, 138.2, 136.2, 129.2, 127.8, 127.5, 126.8, 118.4, 115.9, 112.1 ppm. IR (neat, cm⁻¹): 3285, 2919, 1633, 1580, 1483, 1431, 1384, 1349, 1156, 768.



N-(**pyrazin-2-yl**)**pyrimidin-5-amine** (Table 4) Following general procedure D, a mixture of 5-iodopyrimidine (103 mg, 0.5 mmol), aminopyrazine (67 mg, 0.7 mmol), K_2CO_3 (99 mg, 0.7 mmol), **5** (4 mg, 1 mol%) and **1** (2.5 mg, 1 mol%), and *t*-BuOH (1 mL) was heated to 110 °C for 24 h. The crude product was purified via the Biotage SP4 (silica-packed 50 g snap column; 0-10% MeOH/EtOAc) to provide the title compound as a white solid (76 mg, 88%), sublimed above 200 °C. ¹H NMR (300 MHz, DMSO) δ : 9.88 (s, 1H), 9.11 (s, 2H), 8.76 (s, 1H), 8.27 (s, 1H), 8.18 (s, 1H), 8.02 (s, 1H) ppm. ¹³C NMR (75 MHz, DMSO) δ : 152.1, 151.8, 146.5, 141.7, 136.7, 135.8, 135.6 ppm. IR (neat, cm⁻¹): 3388, 2918, 1579, 1529, 1443, 1384, 1055, 1005, 833, 716.

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