



#### Additional file 7.

**Design of a homologous IPC for use in the qPCR/MCA or qPCR assays.** This DNA control can be used for coextraction and/or coamplification with the clinical samples. The Mfold web server [1] was used for prediction of its secondary structure ( $\Delta G = -0.15$  kcal/mol).

The 93 bp sequence (TCAAGCAAAGCAAAGGCTCGCCGCCAGCACGCCGCCGCACGT  
**GGACTTTCGGCCCTTCTTGGCCTTTGCGGGCATCCATCGCGAGCACGGA**) is compatible with the qPCR/MCA and the TaqMan qPCR assays developed in this work. The target sites of the primers used in the qPCR/MCA and the qPCR assays for *A. astaci*-detection are underlined. Note that the 16 bp-reverse primer used in qPCR/MCA is shortened by a base at its 5' end compared to the reverse primer of TaqMan qPCR. Bold letters denote the site targeted by the hydrolysis probe DYE-AAAGGCCAAGAAGGGCGGAAAGTCC-BHQ to monitor the IPC in the TaqMan qPCR assay designed for *A. astaci*-quantification.

In the dye-based qPCR/MCA assay the IPC can be differentiated by a 3 °C higher melting temperature from the *A. astaci*-specific chitinase peak. This was predicted by the Nearest Neighbour method implemented in the online oligonucleotide properties calculator OligoCalc [2].

#### References:

1. Zuker M: **Mfold web server for nucleic acid folding and hybridization prediction.** *Nucleic Acids Res* 2003, **31**(13):3406-3415.
2. Kibbe WA: **OligoCalc: an online oligonucleotide properties calculator.** *Nucleic Acids Res* 2007, **35**(Web Server issue):W43-46.