## **SUPPLEMENTAL MATERIAL**

**Generation of hyper-immune bovine sera**. Hyperimmune sera were obtained from cattle experimentally inoculated with *E. coli* O157 strains of both cattle and human origin (1), which included the strain EDL933, one of the sequenced O157 strains (2). The rationale was that exposure of these cattle to diverse O157 strains of both lineage I and II (3) might engender antibody responses against a wider variety of protein antigens expressed in the gastrointestinal tract (GIT) of cattle rendering the hyperimmune sera optimal for profiling the immunoproteome of this organism in the bovine reservoir. Cattle were first confirmed to be negative for O157 colonization using highly sensitive 10 recto-anal swab culture techniques (1), and then inoculated once orally with  $10^{10}$  CFU of each strain (1). Serum samples, collected from nine cattle that had remained culture positive for up to two months (1) following experimental inoculation, demonstrated high titers of anti-O157 lipopolysaccharide (LPS) antibodies (1). Preimmune sera were also collected from the same animals prior to inoculation.

**Preliminary evaluation of reactivity of pooled hyperimmune sera.** We assessed the quality of pooled hyperimmune sera, by reacting them with previously identified, secreted O157 proteins encoded in the locus of enterocyte effacement (LEE) (4). Cattle immunized with such proteins, including those expressed from the LEE, reportedly show decreased shedding of O157 (4). We first confirmed reactivity of hyper-immune cattle sera with O157 lipopolysaccharide (LPS) purified in our laboratory using a protocol described previously (5). We then amplified genes encoding full-length EspB from O157 strain EDL933 (test), and PilA from *Vibrio cholerae* El Tor N16961 (control), and

cloned them into the expression vector, pET-30b (Novagen, Inc., Madison, WI.) under 2 control of the phage T7 promoter. Following confirmation of in-frame cloning via gene sequencing, we transformed plasmids into *E. coli* BL21(DE3). Recombinant clones were induced with isopropyl-β-D-thiogalactopyranoside (IPTG) and probed in a colony immunoblot assay with pooled, hyper-immune cattle sera adsorbed against purified O157 LPS, as described previously by our group (6). None of the above recombinant clones reacted with pooled, pre-immune cattle sera (data not shown).

**Affinity-purification of PAbs from hyperimmune sera**. Prior to affinity purification, we pooled hyperimmune sera from nine cattle to compensate for variations in individual immune responses and identify a wider complement of O157 proteins expressed within the GIT of bovine reservoirs. PAbs from 1 ml of pooled hyperimmune cattle sera were purified using HiTrap Protein G HP (5 ml) columns (Amersham Biosciences, Piscataway, NJ), as recommended by the manufacturer with a few modifications. Briefly, pooled sera were diluted 1:4 in binding buffer (0.02 M Sodium phosphate buffer, pH 7.0), and then loaded using a syringe onto a protein G column equilibrated with ten volumes of binding buffer. Protein G binds all IgG subclasses but not other immunoglobulin isotypes (7). Following a wash with ten volumes of binding buffer, bound IgG PAbs were eluted with 4 ml of elution buffer (0.1 M Glycine, pH 2.7), directly into five tubes each containing 200 μl of 1M Tris-HCl, pH 9.0. Affinity-purified IgG PAbs ("Bait" PAbs) were quantified using a nomograph (8) and prepared for coupling. 

## **REFERENCES**

- 1. Rice, D.H., Sheng, H.Q., Wynia, S.A., and Hovde, C.J. (2003) Rectoanal Mucosal Swab Culture Is More Sensitive Than Fecal Culture and Distinguishes *Escherichia coli* O157:H7-Colonized Cattle and Those Transiently Shedding the Same Organism. *J. Clin. Microbiol.* **41**, 4924-4929.
- 2. Perna, N.T., Plunkett, G., Burland, V., Mau, B., Glasner, J.D., Rose, D.J., Mayhew, G.F., Evans, P.S., Gregor, J., Kirkpatrick, H.A., Posfai, G., Hackett, J., Klink, S., Boutin, A., Shao, Y., Miller, L., Grotbeck, E.J., Davis, N.W., Lim, A., Dimalanta, E.T., Potamousis, K.D., Apodaca, J., Anantharaman, T.S., Lin, J., Yen, G., Schwartz, D.C., Welch, R.A., and Blattner,F.R. (2001) Genome sequence of enterohaemorrhagic *Escherichia coli* O157:H7. *Nature* **409**, 529- 533.
- 3. Kim, J., Nietfeldt, J., and Benson, A.K. (1999) Octamer-based genome scanning distinguishes a unique subpopulation of *Escherichia coli* O157:H7 strains in cattle. *Proc. Natl. Acad. Sci.* **96**, 13288-13293.
- 4. Potter, A.A., Klashinsky, S., Li, Y., Frey, E., Townsend, H., Rogan, D., Erickson, G., Hinkley, S., Klopfenstein, T., Moxley, R.A., Smith, D.R., and Finlay, B.B. (2004) Decreased shedding of *Escherichia coli* O157:H7 by cattle following vaccination with type III secreted proteins. *Vaccine* **22**, 362-369.
- 5. Inzana,T.J. (1983) Electrophoretic heterogeneity and interstrain variation of the lipopolysaccharide of *Haemophilus influenzae*. *J Infect Dis* **148**, 492-499.
- 6. John, M., Kudva, I.T., Griffin, R.W., Dodson, A.W., McManus, B., Krastins, B., Sarracino, D., Progulske-Fox, A., Hillman, J.D., Handfield, M., Tarr, P.I., and Calderwood, S.B. (2005) Use of In Vivo-Induced Antigen Technology for Identification of *Escherichia coli* O157:H7 Proteins Expressed during Human Infection. *Infect. Immun.* **73**, 2665-2679.
- 7. Harlow, E.and Lane, D. (1998) Antibodies-A Laboratory Manual. (Cold Spring Harbor Laboratory, New York).
- 8. Bollag, D.M., Rozycki, M.D., and Edelstein, J.S. (1996) Protein Methods . (Wiley-Liss, Inc., New York).
- 9. Dziva, F., van Diemen, P.M., Stevens, M.P., Smith, A.J., and Wallis, T.S. (2004) Identification of *Escherichia coli* O157 : H7 genes influencing colonization of the bovine gastrointestinal tract using signature-tagged mutagenesis. *Microbiology* **150**, 3631-3645.
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## **Supplementary Table 1.** *In vivo***-expressed O157 proteins identified by PELS in cattle.**











Periplasm 4 10% - -

Periplasm 1 6% - -

Periplasm 1 3% - -

Periplasm 1 9% - -

















![](_page_17_Picture_86.jpeg)

![](_page_18_Picture_109.jpeg)

![](_page_19_Picture_100.jpeg)

![](_page_20_Picture_109.jpeg)

![](_page_21_Picture_111.jpeg)

glucosidase

![](_page_22_Picture_112.jpeg)

![](_page_23_Picture_123.jpeg)

![](_page_24_Picture_113.jpeg)

![](_page_25_Picture_114.jpeg)

![](_page_26_Picture_140.jpeg)

![](_page_27_Picture_112.jpeg)

![](_page_28_Picture_93.jpeg)

![](_page_29_Picture_101.jpeg)

![](_page_30_Picture_109.jpeg)

![](_page_31_Picture_104.jpeg)

![](_page_32_Picture_121.jpeg)

![](_page_33_Picture_114.jpeg)

![](_page_34_Picture_114.jpeg)

![](_page_35_Picture_117.jpeg)

![](_page_36_Picture_116.jpeg)

![](_page_37_Picture_127.jpeg)

![](_page_38_Picture_90.jpeg)

<sup>1</sup> Based on homology to the sequenced, *E. coli* O157 strain EDL933 genome, and plasmid, pO157.

<sup>2</sup> Putative functions assigned to hypothetical proteins using the Conserved Domain Database (CDD).

<sup>3</sup>Bacterial cell localization of proteins determined by PSORTb v.2.0/PSLpred/ PSORTdb, and SignalP 3.0 prediction program.

<sup>4</sup>STM, signature tagged mutagenesis<sup>9</sup>; IVIAT, *in vivo* induced antigen technology<sup>6</sup>.