Supplemental methods:

Subcellular fractionation of 3T3-L1 adipocyte and western blot. Subcellular fractionation of adipocytes was performed as follows: In brief, we washed D10 mature 3T3-L1 adipocytes stably overexpressing N-terminal c-Myc tagged murine Bscl2 twice with PBS and harvested the cells in TES buffer (20 mM Tris pH 7.4, 1 mM EDTA, 250 mM sucrose) in the presence of protease inhibitor cocktail (Sigma), left them on ice for 30 min before homogenizing them by 20 strokes up and down using a Potter-Elvehjem homogenizer with a Teflon pestle. The total homogenate was cleared by centrifugation at 1,500 *g* for 10 min. The pellet contained unlysed cells and released nucleus. The supernatant was centrifuged again at 16,900 *g* for 20 min. The resulting supernatant was collected and subjected to differential centrifugation at 212,000 *g* for 60 min to obtain supernatant as cytosol, and pellet as microsomes. All fractions except cytosol were re-suspended in TES buffer plus 1% Triton X-100, and protein concentrations were determined using a BCA kit (Biorad). For western blot, we loaded equal amounts of proteins on SDS-polyacrylamide gels and transferred them to an Immobilon-P membrane (Millipore). The blots were probed with specific antibodies for visualization by enhanced chemiluminescence (SuperSignal kit; Pierce).