

Supplementary Figure Legends

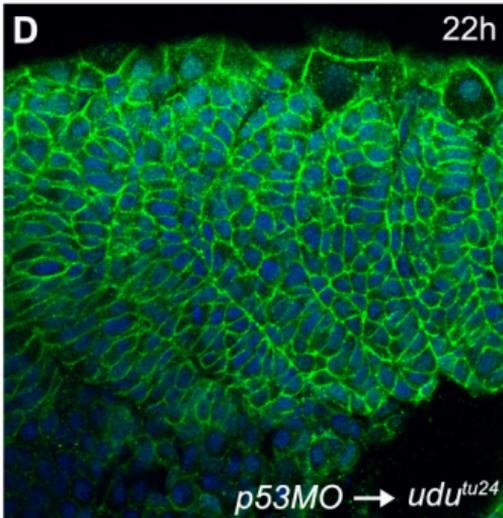
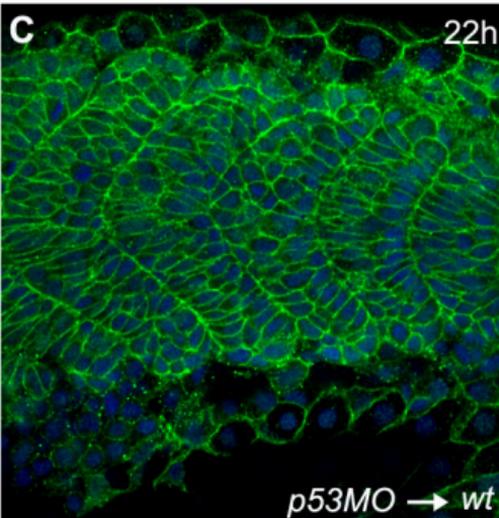
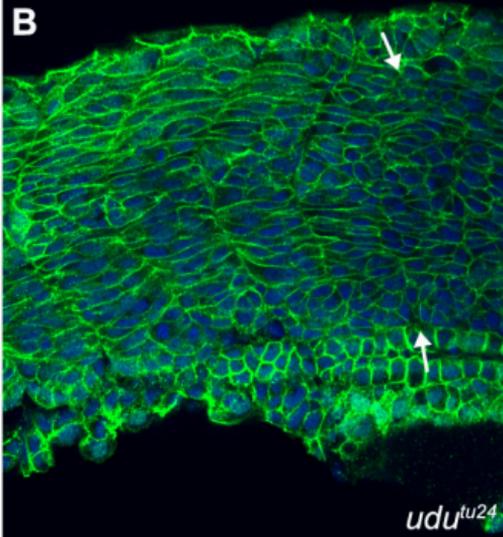
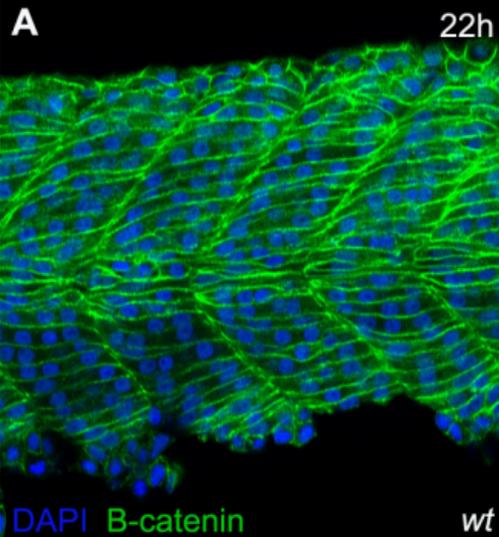
Supplementary Figure S1. Genotyping experiments confirm that embryos selected for subsequent experiments were homozygous *udu^{tu24}* mutants. (A–E) Somite structures of wild-type embryos and *udu^{tu24}* embryos injected with *p53*-MO. (A–B) Wild-type embryos showed somite structures with characteristic chevron-shape and well-defined somite boundaries. (C–D) *udu^{tu24}* embryos have somite structures that were not V-shaped and posterior somites that were not clearly defined by boundaries. (E) In *udu^{tu24}* embryos injected with *p53*-MO, V-shaped somite structures were restored but not with characteristic chevron-shape. (F) Representative DNA sequence analyses showed nonsense mutation in homozygous *udu^{tu24}* mutants in which Tyr has been converted to a STOP codon.

Supplementary Figure S2. Somite boundaries highlighted by membrane-localized β -catenin antibody. (A–B) High resolution confocal images of somite boundaries in wild-type and *udu^{tu24}* embryos, approximately the 12th to 16th somite region. Somites of wild-type embryos were chevron-shaped with clear boundaries; however, mutant somite boundaries were not clearly defined as indicated by arrows. (C–D) High resolution confocal images of posterior somite boundaries in wild-type and *udu^{tu24}* embryos injected with *p53*-MO.

Supplementary Figure S3. Segmentation clock is functional in *udu^{tu24}* mutants. Expression of *deltaC* in homozygous *udu^{tu24}* embryos at 15 hpf (12 ss). *deltaC* expression patterns were arranged in a sequence corresponding to a temporal oscillation in the PSM linked to somitogenesis. The different phases of the oscillation cycle were numbered I, II and III and intermediate phases were designated I+, II+ and III+. All embryos were in dorsal view with anterior to the top.

Supplementary Table 1. Gene accession number and primer sequence used in quantitative real-time PCR.

Gene	Accession number	5' sequence	3' sequence
<i>p53</i>	NM_131327	ACCACAGCTTGGTGCTGAAT	GCCTGGACATGCACACACA
<i>mdm2</i>	AF010255	TGGAGTCCATCAACTCTGAT	CGAGGAAGAGGAGGGTTGAAC
<i>cyclin D1</i>	NM_131025	GCAGCTTTTAGGAGCAACTT	CTGAGGCCAGATCCCACCTC
<i>p21^{WAF/CIP1}</i>	AL912410	GACTGAGGAATGGATCTTTC	CTTCATCTGTCTGGAGCTGCAT
<i>caspase 8</i>	NM_131510	CCAGACAATCTGGATGAACT	CATTGTTTCAGATACAGGGTTGTTG
<i>bax</i>	NM_131562	GCTGCACTTCTCAACAACCTT	CGGGCCACTCTGATGAAGAC
<i>gadd45a1</i>	AY714218	AAGGATGGACTCGGTGATTA	CCTTCACATCGTCGTCATCAGT
<i>gadd45β1</i>	AB180735	AGGAGGTTGTTGGATGCAA	TCTGGATCCACATTCATGAGTTG
<i>gadd45β2</i>	AB180736	GAGGATCTGGACGACATT	TGAGAGTTAGTGACGAGGATGCA
<i>atm</i>	AJ605775	ACGAGCTAATCTGTCAGTCA	TTCTGCCCCTCCAAGTC
<i>atr</i>	CT623247	ATGGTCACTGACAACAAGCT	GCGGCCAGCTTTTTCAAAT
<i>chk1</i>	CB363820	TGCGTTAACAACCGCTTCA	TTTTGCCATGTCCACAACCTT
<i>chk2</i>	AF265346	AGCGTGCTCAATTGCACAGA	TCGTCTGCCACCCATGTG
<i>elf1a</i>	NM_131263	TGGAAATTCGAGACCAGCAA	TCACGGGTCTGTCCGTTCTT



15h, *deltaC/udu*^{tu24}

