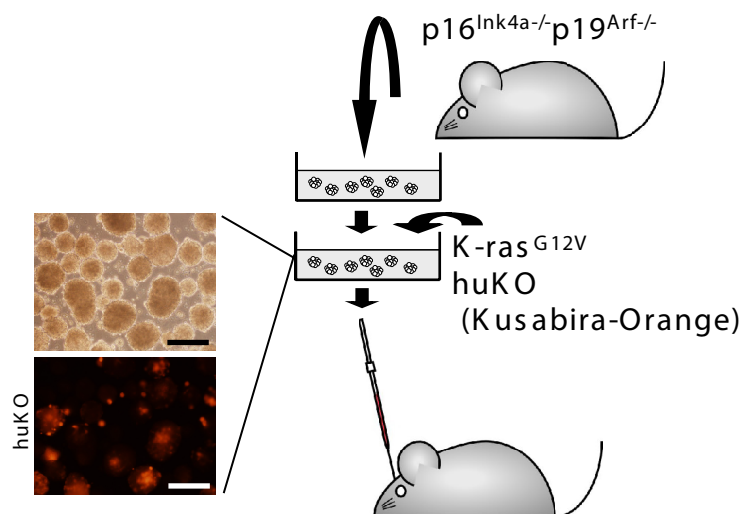
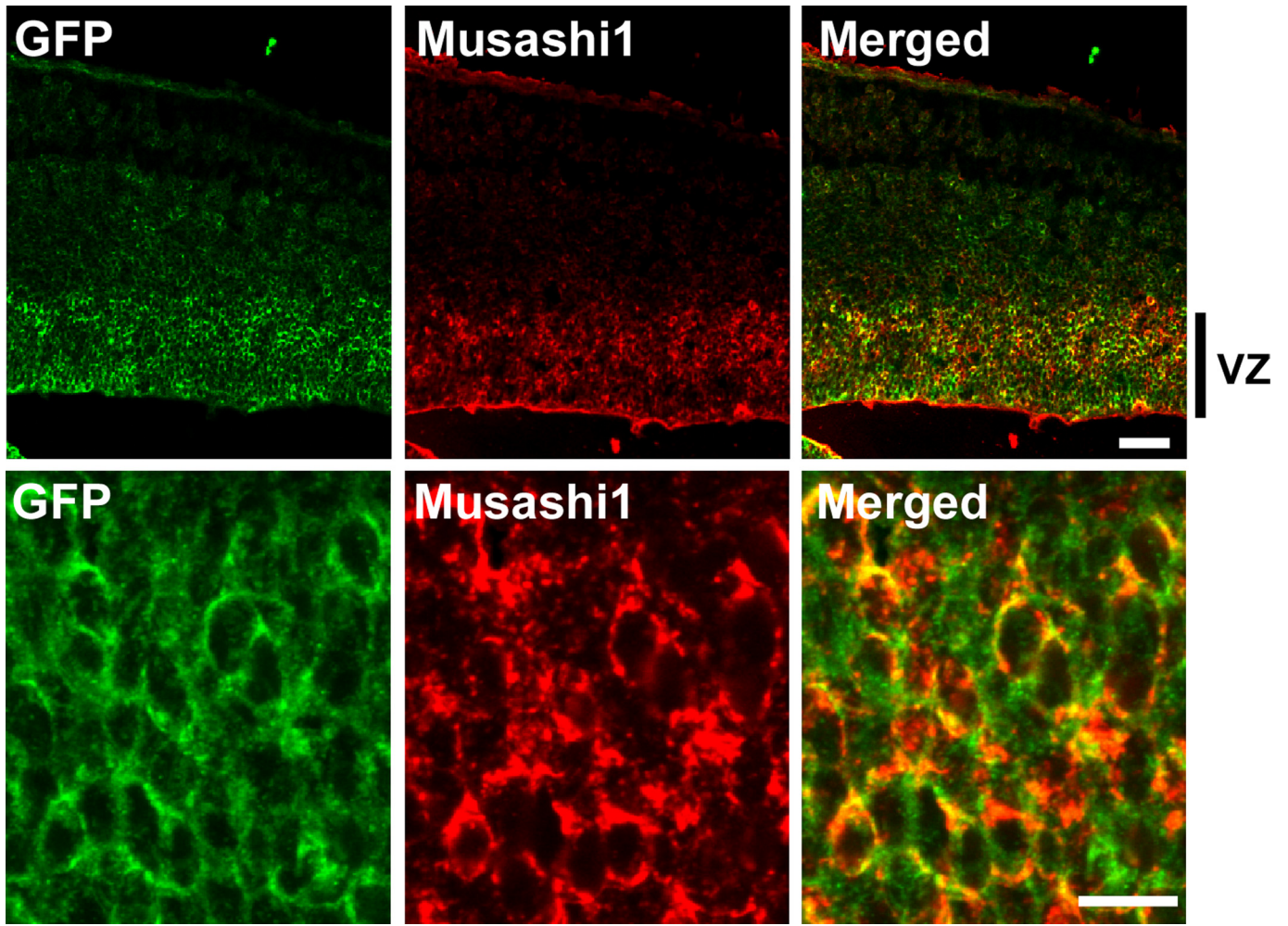


# Supporting Information

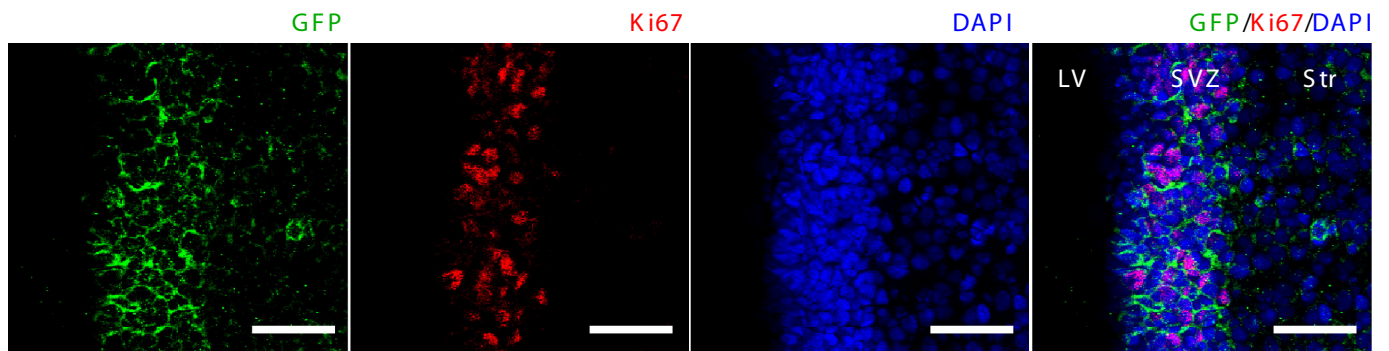
Tamase et al. 10.1073/pnas.0905016106



**Fig. S1.** Generation of the brain tumor model. Neurosphere cells derived from  $p16^{Ink4a^{-/-}}p19^{Arf^{-/-}}$  neonates were infected with retrovirus carrying the mutant K-ras ( $K-ras^{G12V}$ ) and  $huKO$  genes. These cells were then inoculated into the brains of WT recipient mice. Neurosphere cells (Upper, bright field; Lower,  $huKO$  fluorescence) are shown. (Scale bars: 200  $\mu m$ .)

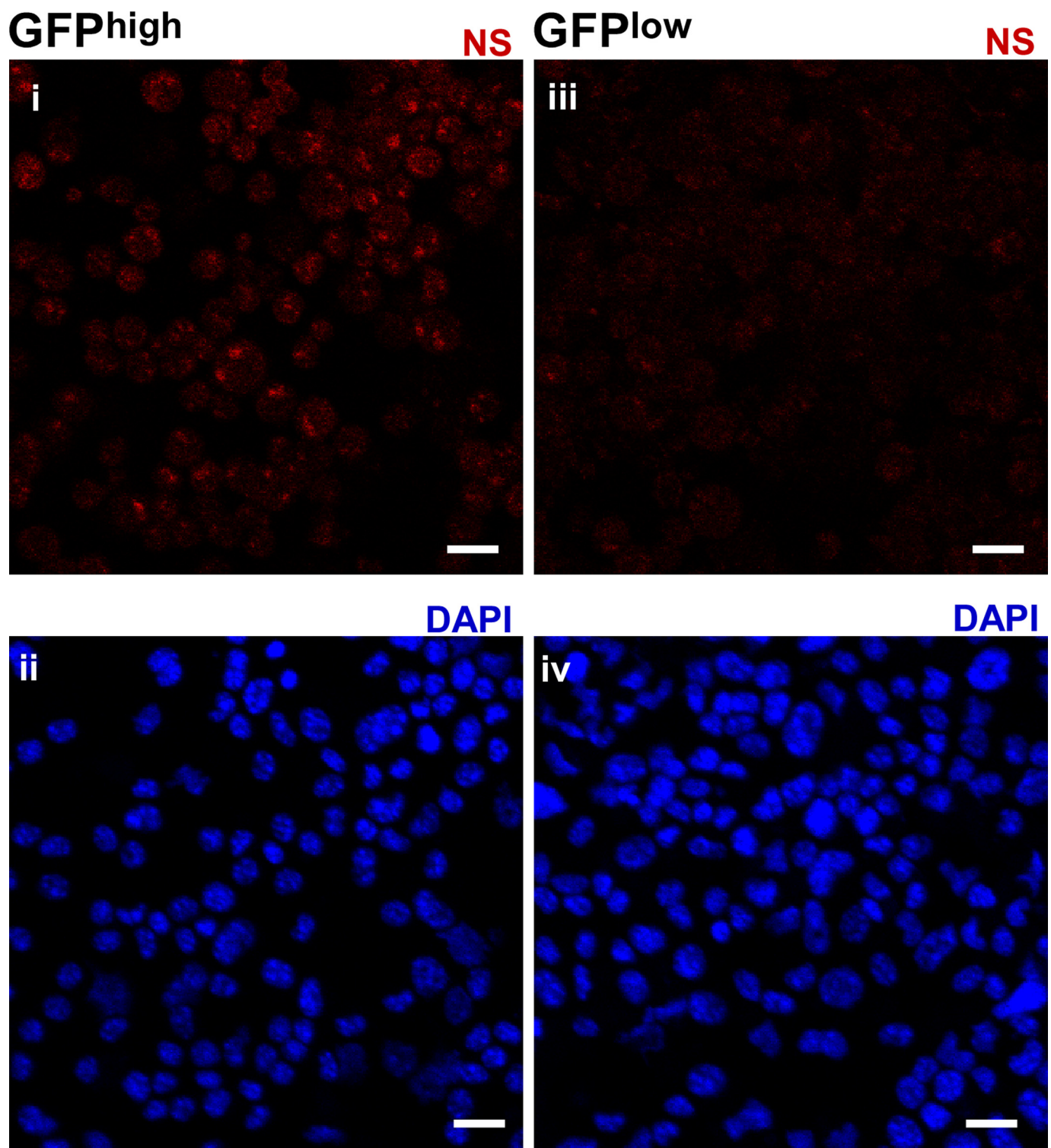


**Fig. S2.** NSC/NPCs in E14.5 NS-GFP-Tg mice. Coronal sections of the forebrains of E14.5 NS-GFP-Tg mice were subjected to immunofluorescence analysis with anti-GFP (green) and anti-musashi-1 (14H-1, red) [Kaneko, Y., et.al. (2000) *Dev Neurosci* 22:139–153]. VZ, ventricular zone. (Bottom) Higher magnification views of the images in Upper. (Scale bars: Upper, 100  $\mu\text{m}$ ; Lower, 10  $\mu\text{m}$ ). For immunostaining with the biotinylated anti-musashi-1 antibody, the tyramide signal amplification (TSA) system was used according to the manufacturer’s protocol (Perkin Elmer).

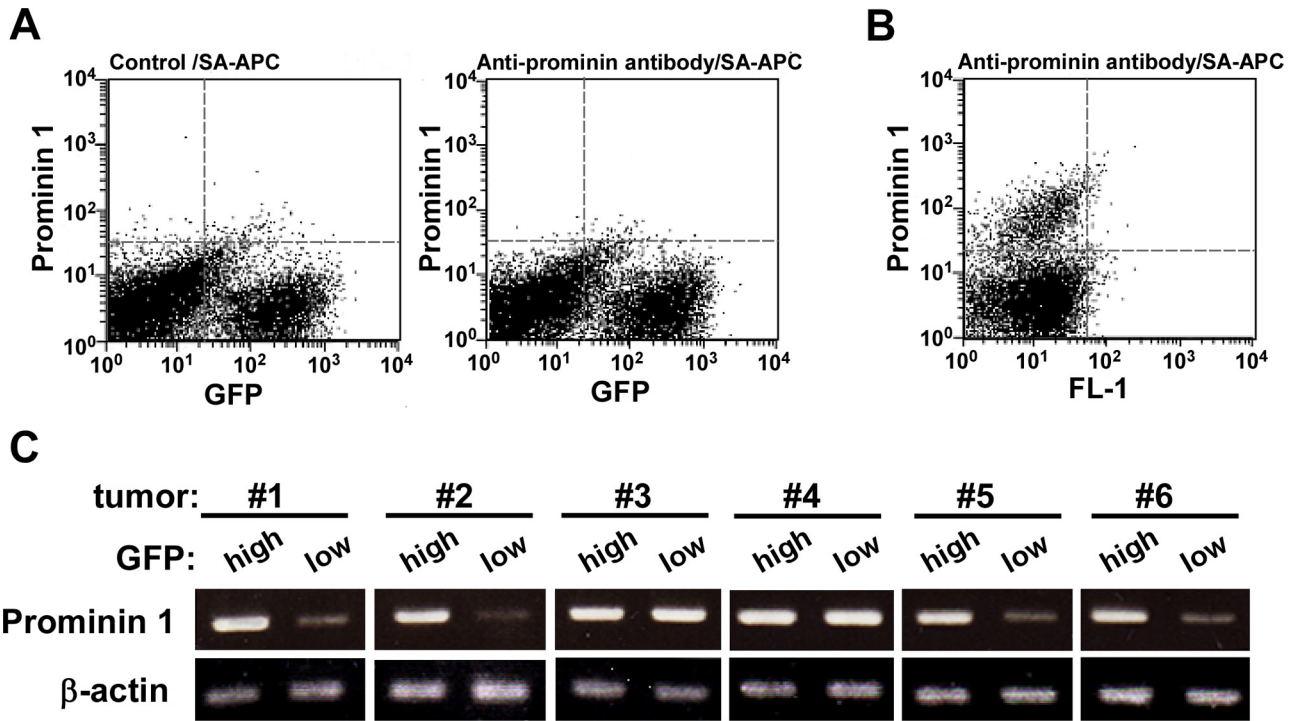


**Fig. S3.** NSC/NPCs in postnatal NS-GFP-Tg mice. Coronal sections of forebrains of P3 NS-GFP-Tg mice were subjected to immunofluorescence analysis with anti-GFP (green), anti-Ki67 (red) and DAPI (nuclear staining, blue). SVZ, subventricular zone; Str, striatum; LV, Lateral ventricle. (Scale bars: 50  $\mu\text{m}$ .)



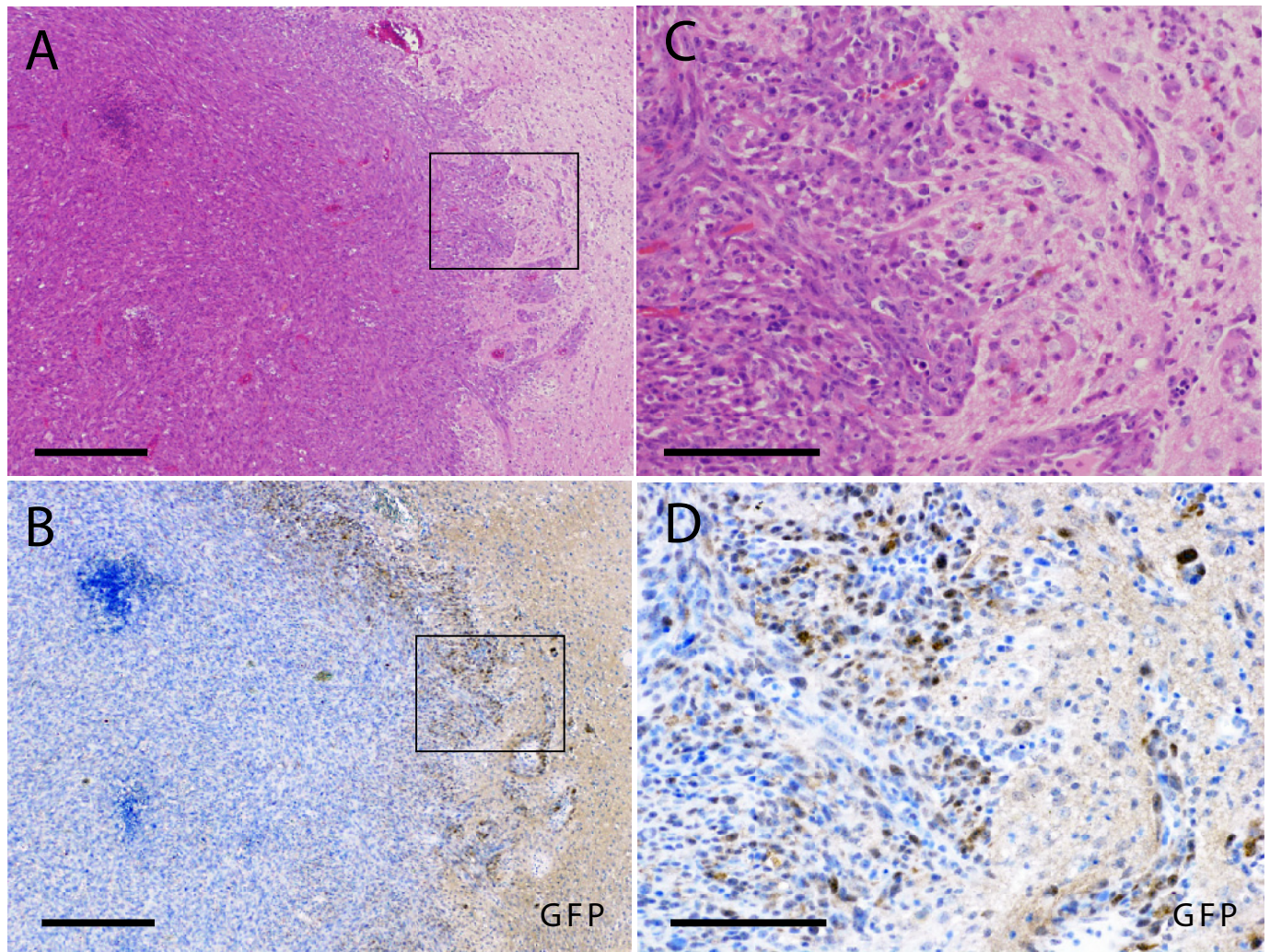


**Fig. S4.** Endogenous NS expression by GFP<sup>high</sup> or GFP<sup>low</sup> tumor cells. Cytospin smears of sorted GFP<sup>high</sup> or GFP<sup>low</sup> cells were fixed and stained with (*i* and *iii*) anti-NS antibody (red) or (*ii* and *iv*) DAPI (blue, nuclear staining). (Scale bars: 20  $\mu\text{m}$ .)



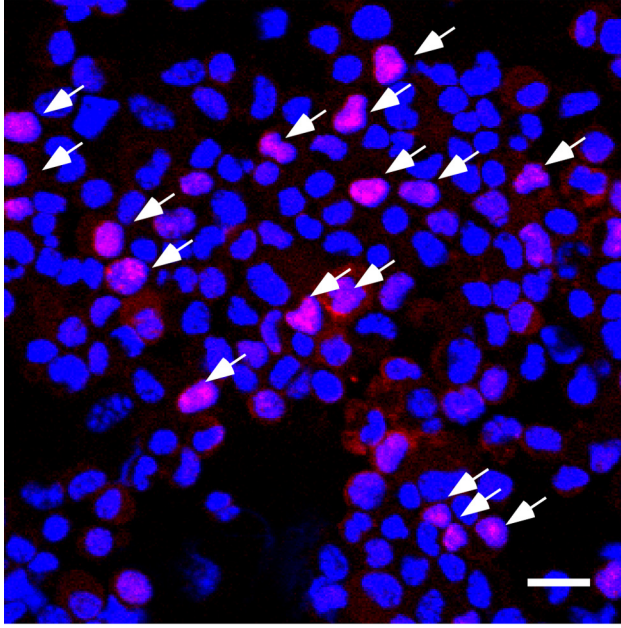
**Fig. S5.** Prominin 1 expression in GFP<sup>high</sup> or GFP<sup>low</sup> tumor cells. (A) Flow cytometry analysis of brain tumor cells with anti-prominin 1 antibody. Dissociated brain tumor cells were stained with biotin-conjugated anti-prominin 1 antibody (13A4, eBioscience), followed by staining with streptavidin (SA)-allophycocyanin (APC) (Right). Simultaneously, tumor cells were incubated with only SA-APC as control (Left). (B) SVZ cells from P3 C57BL/6 mice were stained with anti-prominin 1 antibody as shown in A. Expression of prominin protein was not detected in tumor cells, despite the fact that the antibody recognized prominin protein in normal brain tissue. Note: We assume that the epitope for recognition by antibody in brain tumors may differ or be masked compared with normal tissue. Alternatively, prominin 1 protein expression may be extremely low in tumors. (C) Expression of prominin 1 mRNA in brain tumor cells. Total RNA was purified from GFP<sup>high</sup> and GFP<sup>low</sup> cells isolated from three independent original tumors and prominin 1 mRNA levels were evaluated by RT-PCR.  $\beta$ -actin, control. Prominin 1 mRNA levels in GFP<sup>high</sup> tumor cells were higher than that seen in GFP<sup>low</sup> cells in tumors 1, 2, 5, and 6, but a similar correlation was not observed in tumors 3 and 4.





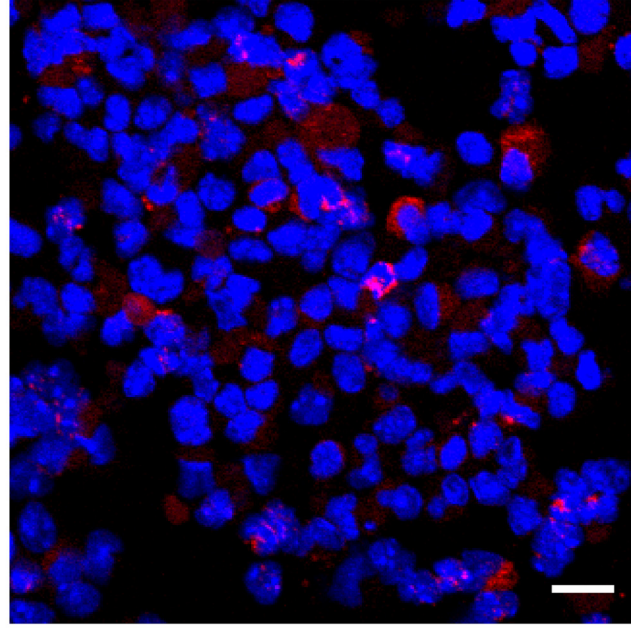
**Fig. S6.** Localization of T-ICs in the original brain tumor. Serial sections of one of the original tumors were subjected to: (A and C) H&E staining and (B and D) anti-GFP staining. Magnified views of the areas indicated by the squares in A and B are shown in C and D, respectively. (Scale bars: A and B, 1 mm; C and D, 200  $\mu\text{m}$ ).

## GFP<sup>high</sup> cells



**Ki67/DAPI**

## GFP<sup>low</sup> cells



**Ki67/DAPI**

**Fig. S7.** Cell cycle status of GFP<sup>high</sup> and GFP<sup>low</sup> tumor cells. Cytospin smears of sorted GFP<sup>high</sup> and GFP<sup>low</sup> cells were fixed and immunostained to detect Ki67 (red). Blue, nuclear marker DAPI. Representative data of 3 independent experiments are shown. (Scale bars: 20  $\mu\text{m}$ .) Arrows, Ki67<sup>+</sup> cells.



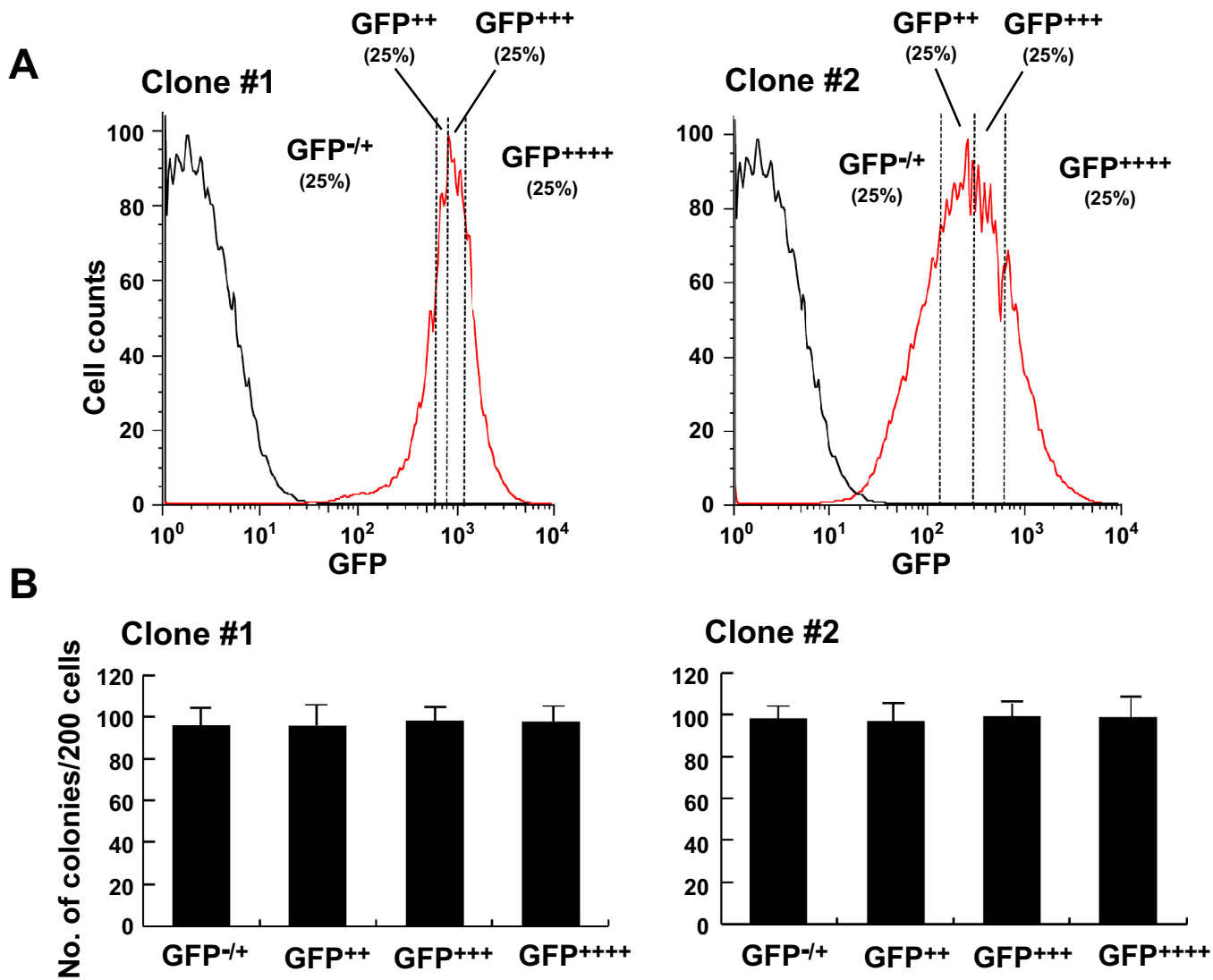


Fig. S8. GFP expression in NS-GFP C6. The NS-GFP construct was introduced into the C6 glioma cell line and single clone-derived lines were established by limiting dilution. (A) Fractionation of GFP-expressing cells from NS-GFP C6 cells. Cells were fractionated by flow cytometry into 4 subpopulations, GFP<sup>-/+</sup>, GFP<sup>++</sup>, GFP<sup>+++</sup> and GFP<sup>++++</sup>, based on GFP fluorescence intensity as indicated. Black peak, control C6; red peak, NS-GFP C6. Two representative clones are shown. (B) Colony forming capacity of fractionated NS-GFP C6 cells. Fractionated cell subpopulations from (A) were cultured. Data shown are the mean number  $\pm$  SD. of colonies generated per 200 cells ( $n = 3$ ).