P42615	MQIPRMSLRQLAWSGA	AVLLLVGTLLLAWSAVRQQESTLAIRAVHQGTTMPDGF	53	YQJB_ECOLI
Q 3 Y X D 0	MQIPRMSLRQLAWSGT	TVLLLVGTLLLAWSAVRQQESTLAIRAVHQGTTMPDGF	53	Q3YXD0_SHISS
A8APZ0	MVKPHITLRQLAWTTS	SFLVLMSVLFLVWSTIRQQESTLAIRAIHQGVSMPDGF	53	A8APZ0_CITK8
Q 8 X F 9 2	MLKPRITARQLIWISA	AFLLMLTILMMTWSTLRQQESTLAIRAVNQGASMPDGF	53	Q8XF92_SALTI
A 6 T E D 5	MMVMKRPSLRQFSWLLG	GGSLLLGALFWLWLAVQQQEATLAIRPVGQGIGMPDGF	54	A6TED5_KLEP7
A4WES8	MVISPLALRRLSYGLI	IALVLLSALILVWTALQRQESTLAIRPVSQGASVPDGF	53	A4WES8_ENT38
A7MIS1	MLS	SAGALLAAMLLCWAALQKHETTLEIRAVSQGATIPDGF	4 0	A7MIS1_ENTS8
Q7CGC4	MINFRGRFGRPLWHYLVLF	PVVLLLLAVILLTPMIVQTESTLKIRPNQQGLSLPDGF	56	Q7CGC4_YERPE
Q6D9H4	MSIRWLSPTLSPRKIGRILLI	LLALPIIALTQSQPPRHSQDDAMLHIKPYDGAALPDGF	58	Q6D9H4_ERWCT
		: : : : : : : * : * * * *		
P42615	SIWHHLDAHGIPFKSITPKNDTI	LLITFDSSDQSAAAKAVLDRTLPHGYIIAQQDNNSQAM	113	YQJB_ECOLI
Q3YXD0	SIWHHLDAHGIPFKSITPKNDTI	LLITFDSSDQSAAAKAVLDRTLPHGYIIAQQDNNSQAM	113	Q3YXD0_SHISS
A 8 A P Z 0	SIWHHLDANGIRFKSITPQNDAI	LLITFDSSAQSAAAKAVLDRTLPHGYIIAQQNGDSQAV	113	A8APZ0_CITK8
Q8XF92	SVLHHLDANGIHFKSITPKNDMI	LLITFDSPAQSAAAKTVLDQTLPHGYVVAQQDDDNETV	113	Q8XF92_SALTI
A 6 T E D 5	SVWHHLDANGIRFKSITPQKDGI	LLIKFDSTAQGAAAKEVLGRALPHGYIIALLEDDNSPT	114	A6TED5_KLEP7
A4WES8	YIWHHLDANGIQFKSITPQDDVI	LLIKFDSSAQSAAAKVVLDRSLPRGYIIALQDDQSQTA	113	A4WES8_ENT38
A7MIS1	SVWHHLDANGIRFKSITPQDDII	LLIKFDTSDQSEAARKVLYRSLPRGFVIAQQEEKSLTP	100	A7MIS1_ENTS8
Q7CGC4	YLYQHLNGRGIHIKSIIPENDSI	L V V S L E F P E Q Q M Q A I E V L Q D V L P A G Y A I V A S E S K K R - H	115	Q7CGC4_YERPE
Q6D9H4	YVYQRLNEKGIAIKSITPEQDSI	LIVRLASPEQSIAARDVLRLSLPK-VTITAQQATSPTP	117	Q6D9H4_ERWCT
	: ::*: .** :*** *:.* *	*::::.******:.:.		
P42615	QWLTRLRDNSHRFG 127 Y	YQJB_ECOLI		
Q3YXD0	QWLTRLRDNSHRFG 127 Q	Q 3 Y X D 0 _ S H I S S		
A 8 A P Z 0	QWLTRLRDNPHRFG 127 A	A 8 A P Z 0 _ C I T K 8		
Q 8 X F 9 2	QWLSRLRESSHRFG 127 Q	Q8XF92_SALTI		
A 6 T E D 5	AWLSRLRDAPHRLG 128 A	A 6 T E D 5 _ K L E P 7		
A4WES8	VWLTRLRDTSHRFG 127 A	A4WES8_ENT38		
A7MIS1	AWLTRLRHDIQRLG 114 A	A7MIS1_ENTS8		
Q 7 C G C 4	RLLPVFRSNQQNLG 129 Q	Q7CGC4_YERPE		
Q6D9H4	FWQQKLTQKQSKLG 131 Q	Q 6 D 9 H 4 _ E R W C T		
	: :*			

Figure S1. Multiple sequence alignment of MzrA (YqjB) sequences from *Enterobacteriaceae*. Amino acid sequences were aligned using the CLUSTAL W (1.83) program

(http://ch.EMBnet.org/). Amino acids that are either identical (*), have similar side chains (:) or similar overall characteristics (.) are shown in the last line.

Strain or	Description	Reference or Source
Plasmid	Description	
MC4100	F- araD139 Δ(argF-lac)U139 rspL150 relA1 fibB5301 ptsF25	(Casadahan 1076)
	deoCl thi-1 rbsR	(Casadaball, 1970)
JM109	e14-(McrA-) recA1 endA1 gyrA96 thi-1 hsdR17 (r _K m _K ⁺) supE44 relA1	Promega
	Δ (<i>lac-proAB</i>) [F' <i>traD36 proAB lacI</i> ₄ Z Δ M15].	Tomega
RAM1292	MC4100 $\Delta ara714$	(Werner and Misra, 2005)
BTH101	F- cya99 araD139 galE15 galK16 rpsL1 (Str ') hsdR2 mcrA1	FUROMEDEX
	mcrB1	LOKOMEDEX
BL21(DE3)	F–, $ompT$, $hsdS_B(r_B-, m_B-)$, dcm , gal , $\lambda(DE3)$	Novagen
RAM1318	MC4100 Δ <i>bamB</i> ::scar <i>degP</i> ::Tn10	(Charlson <i>et al.</i> , 2006)
RAM1503	RAM1292 / pBAD24	This Study
RAM1504	RAM1292 / pBAD24- <i>mzrA</i>	This Study
RAM1505	RAM1292 / pBAD24-mzrA(FLAG)	This Study
RAM1506	RAM1292 / pBAD33	This Study
RAM1507	RAM1292 / pBAD33-mzrA(FLAG)	This Study
RAM1508	RAM1292 $\Delta mzrA$::scar	This Study
RAM1509	RAM1508/ pBAD33	This Study
RAM1510	RAM1508/ pBAD33-mzrA(FLAG)	This Study
RAM1511	RAM1508 $\Delta omr::lacZ$	This Study
RAM1512	RAM1292 $\Delta yqj::lacZ$	This Study
RAM1513	RAM1512 zii::Tn10	This Study
RAM1514	RAM1512 <i>cpxA</i> * <i>zii</i> ::Tn <i>10</i>	This Study
RAM1515	RAM1512 $\Delta cpxR$::Cm ^r zii::Tn10	This Study
RAM1516	RAM1292 zii::Tn10	This Study
RAM1517	RAM1292 <i>cpxA</i> * <i>zii</i> ::Tn10	This Study
RAM1518	RAM1516 $\Delta mzrA$::scar	This Study
RAM1519	RAM1517 $\Delta mzrA$. scar	This Study
RAM1520	RAM1516 AenvZ··Cm ^r	This Study
RAM1521	$RAM1517 \Lambda envZ^{}Cm^{r}$	This Study
RAM1522	RAM1292 Aomr. lacZ	This Study
RAM1523	RAM1522 / nBAD24	This Study
RAM1524	RAM1522 / pBAD24-mzrA	This Study
RAM1525	RAM1522 = 7 pBHD2 + m2m $RAM1522 = 7 iii:Tn 10$	This Study
RAM1526	RAM1522 cmr4* zijTn10	This Study
RAM1527	$RAM1522 \Delta cmr R^{T} Cm^{T} zii Tn 10$	This Study
RAM1528	$R \Delta M 1525 \Delta m zr A \cdots scar$	This Study
RAM1520	RAM1525 AmzrAsear	This Study
DAM1520	$\frac{P M M 1202}{P M M 202} M M M M M M M M M M M M M M M M M M M$	This Study
DAM1521	$\frac{RAW1272}{m} \frac{M}{2} \frac{M}{m} \frac{M}{m$	This Study
DAM1532	RAM1530 / pTro00A mar $A/ELAG$	This Study
DAM1522	$\frac{RAM1530}{PMC37R-m2rA(FLAG)}$	This Study
RAM1533	$RAM1330 \Delta 0 mr ucz$	This Study
RAM1534	$\frac{\text{RAW1292 } OmpR4/2}{\text{DAM1524 } / \text{pTro00A}}$	This Study
RAMI533	RAW1534 / p11099A $RAW1524 / rTre00A menA/ELAC)$	This Study
RAMI530	RAM1534 / p1fc99A-m2fA(FLAG)	This Study
RAMI53/	$RAM1292 \Delta envZ::scar$	This Study
RAM1538	RAM1537 / p1rc99A	I his Study
RAM1539	RAM1537/p1rc99A-mzrA(FLAG)	This Study
KAM1540	$KAM153 / \Delta omr::lacZ$	This Study
KAMI541	KAM1292 envZ[K39/L]	This Study
KAM1542	KAM1541 $\Delta omr::lacZ$	This Study
KAM1543	B1H101 / pKT25 pUT18C	This Study
KAM1544	BTH101 / pKT25mzrA and pUT18C	This Study
RAM1545	BTH101 / pKT25mzrA and pUT18CenvZ	This Study
RAM1546	BTH101 / pKT25 <i>mzrA</i> and pUT18 <i>envZ</i>	This Study

RAM1547	BTH101 / pKT25 <i>mzrA</i> and pUT18C <i>ompR</i>	This Study
RAM1548	BTH101 / pKT25mzrA and pUT18ompR	This Study
RAM1549	BTH101 / pKT25envZ and pUT18envZ	This Study
RAM1551	RAM1292 $\Delta ppiA::lacZ$	This Study
RAM1552	RAM1551/pBAD24	This Study
RAM1553	RAM1551/pBAD24-mzrA	This Study
RAM1554	RAM1292 $\Delta ompF::lacZ$	This Study
RAM1555	RAM1554 / pBAD24	This Study
RAM1556	RAM1554 / pBAD24-mzrA	This Study
RAM1557	RAM1292 mgrB::\placMu55	This Study
RAM1558	RAM1557 / pBAD24	This Study
RAM1559	RAM1557 / pBAD24-mzrA	This Study
RAM1560	RAM1292 mgtA::λplacMu55	This Study
RAM1561	RAM1560 / pBAD24	This Study
RAM1562	RAM1560 / pBAD24-mzrA	This Study
RAM1563	RAM1292 / pBAD24-mzrA::phoA	This Study
RAM1564	RAM1292 / pBAD24-mzrA[Δ13-28]::phoA	This Study
RAM1565	RAM1292 dsbA ⁻ ::Kan ^r / pBAD24-mzrA::phoA	This Study
RAM1566	RAM1292 / pBAD24-mzrA::lacZα	This Study
RAM1567	RAM1292 / pBAD24- <i>mzrA</i> [Δ13-28]:: <i>lacZα</i>	This Study
RAM1568	JM109 pBAD24	This Study
RAM1569	JM109 pBAD24-mzrA	This Study
RAM1570	JM109 pBAD24-mzrA::lacZα	This Study
RAM1571	JM109 pBAD24- <i>mzrA</i> [13-28]:: <i>lacZα</i>	This Study
RAM1572	RAM1540 / pBAD24	This Study
RAM1573	RAM1540 / pBAD24-mzrA	This Study
RAM1574	RAM1292 $\Delta rseA$::scar	This Study
RAM1575	RAM1574 $\Delta yqj::lacZ$	This Study
RAM1576	RAM1292/ pTrc99A	This Study
RAM1577	RAM1292/ pTrc99A-mzrA(FLAG)	This Study
RAM1578	RAM1318/ pBR322	This Study
RAM1579	RAM1318/ pBR322 exuT'exuRyqjABCDE'	This Study
RAM1580	RAM1292 λ RS88 <i>cpxP</i> '- <i>lacZ nadA</i> ::Tn10	This Study
RAM1581	RAM1580/ pBAD24	This Study
RAM1582	RAM1580/ pBAD24-mzrA(FLAG)	This Study
RAM1583	RAM1530 $\Delta yqj::lacZ$	This Study
RAM1584	RAM1537 $\Delta yqj::lacZ$	This Study
RAM1585	RAM1541 $\Delta yqj::lacZ$	This Study
RAM1586	RAM1292 envZ11	This Study
RAM1587	RAM1586 $\Delta yqj::lacZ$	This Study

Plasmids

pBAD24	Amp ^r ; Expression vector; arabinose inducible	(Guzman et al., 1995)
pBAD33	Cm ^r ; Expression vector; arabinose inducible	(Guzman et al., 1995)
pTrc99A	Amp ^r ; Expression vector; IPTG inducible	GE Healthcare
pBAD33 <i>mzrA-</i> FLAG	Cm ^r ; pBAD33 containing the <i>mzrA</i> (FLAG) gene	This Study
pBAD24mzrA	Amp ^r ; pBAD24 containing the <i>mzrA</i> gene	This Study
pBAD24 <i>mzrA-</i> FLAG	Amp ^r ; pBAD24 containing the <i>mzrA</i> (FLAG) gene	This Study
pBAD24 <i>mzrA-</i> phoA	Amp ^r ; pBAD24- <i>mzrA</i> :: <i>phoA</i>	This Study
pBAD24 <i>mzrA</i> [∆ 13-28]-phoA	Amp ^r ; pBAD24- <i>mzrA</i> [Δ13-28]:: <i>phoA</i>	This Study
pBAD24 <i>mzrA-</i> lacZα	Amp ^r ; pBAD24- <i>mzrA</i> :: <i>lacZ</i> α	This Study

pBAD24 <i>mzrA</i> [Δ 13-28]- <i>lacZα</i>	Amp ^r ; pBAD24- <i>mzrA</i> [Δ 13-28]:: <i>lacZ</i> α	This Study
pTrc99A <i>mzrA</i> - FLAG	Amp ^r ; pTrc99A containing the <i>mzrA</i> (FLAG) gene	This Study
pKD3	Cm ^r ; Plasmid carrying Cm ^r gene cassette	(Datsenko and Wanner, 2000)
pKD4	Kan ^r ; Plasmid carrying Kan ^r gene cassette	(Datsenko and Wanner, 2000)
pKD46	Amp ^r ; Red recombinase expression plasmid	(Datsenko and Wanner, 2000)
pCP20	Cm ^r and Amp ^r ; FLP recombinase expression plasmid, temperature sensitive	(Datsenko and Wanner, 2000)
pKG136	<i>ahp</i> FRT <i>lacZY</i> ⁺ t _{his} oriR6K	(Ellermeier et al., 2002)
pKT25	Kan ^r ; Harbors the T25 fragment of adenylate cyclase from <i>B. pertussis</i>	EUROMEDEX
pUT18	Amp ^r ; Harbors the T18 fragment of adenylate cyclase from <i>B.</i>	EUROMEDEX
pUT18C	Amp ^r ; Harbors the T18 fragment of adenylate cyclase from <i>B.</i>	EUROMEDEX
pKT25 <i>mzrA</i>	Kan ^r : pKT25 with the <i>mzrA</i> gene	This Study
pKT25envZ	Kan ^r ; pKT25 with the <i>envZ</i> gene	This Study
pKT25 <i>ompR</i>	Kan ^r ; pKT25 with the <i>ompR</i> gene	This Study
pUT18CmzrA	Amp ^r ; pUT18C with the <i>mzrA</i> gene	This Study
pUT18CenvZ	Amp ^r ; pUT18C with the <i>envZ</i> gene	This Study
pUT18CompR	Amp ^r ; pUT18C with the <i>ompR</i> gene	This Study
pUT18CcpxA	Amp ^r ; pUT18C with the <i>cpxA</i> gene	This Study
PUT18envZ	Amp ^r ; pUT18 with the <i>envZ</i> gene	This Study
pUT18 <i>ompR</i>	Amp ^r ; pUT18 with the <i>ompR</i> gene	This Study
pUT18 <i>cpxA</i>	Amp ^r ; pUT18 with the <i>cpxA</i> gene	This Study
pBR322	Amp ^r ;Tet ^r cloning vector	(Bolivar, et al. 1977)
pBR322-exuT' exuRyqjABCDE'	Amp ^r ;pBR322 with <i>exuT'exuRyqjABCDE'</i> genes	This Study

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2. Casadaban M J (1976) Transposition and fusion of the lac genes to select promoters in *Escherichia coli* using bacteriophage lambda and Mu. *J Mol Biol* **141:**541-555.

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4. Datsenko K A, Wanner B L (2000) One-step inactivation of chromosomal genes in *Escherichia coli* K-12 using PCR products. *Proc Natl Acad Sci USA* **97:**6640-6645.

5. Ellermeier C D, Janakiraman A, Slauch J M (2002) Construction of targeted single copy *lac* fusions using λ Red and FLP-mediated site-specific recombination in bacteria. *Gene* **290**:153-161.

6. Guzman L M, Belin D, Carson M J, Beckwith J (1995) Tight regulation, modulation, and high-level expression by vectors containing the arabinose P_{BAD} promoter. *J Bacteriol* **177:**4121-4130.

7. Werner J, Misra R (2005) YaeT (Omp85) affects the assembly of lipid-dependent and lipid-independent outer membrane proteins of *Escherichia coli*. *Mol Microbiol* **57**:1450-1459.

Table S2: Primers used for cloning and deletion

Deletion Primers				
Primer Name	Sequence (5'-3')			
vai 4 Fwd	GTTAACCGATTCACCAGGAATAATGAATGGAACTTTTGTGTAGGCTGGA			
<i>yqjA</i> Pwu	GCTGCTTCG			
mar A Fund	GAAATATGGAAATCGGGGGGTAAGGGATGCAAATATG	GTAGGCTGGAGCT		
<i>mzrA</i> Fwd	GCTTCG			
man (Dou	GTGAGTGATATGTTTCGGATAACAGGAAGTTACATATGAATATCCTCCTT			
m2rA Kev	AG			
anuZ Euro	CTTTGTACCGGACGGCTCTAAAGCATGAGGCGATTG	CGCTTCTGTAGGCT		
envz rwu	GGAGCTGCTTCG			
amuZ Dou	ACCTTCGCCTCCCGTTTATTTACCCTTCTTTTGTCGTC	JCCCATATGAATAT		
envz Rev	CCTCCTTAG			
omn P Fund	CCGCTGGTGGCGTTTGGCTTCAGGTTGCTAAAGTGGTGATTGTAGGCTGG			
om B I wa	AGCTGCTTCG			
omu (Dou	ACACGAGATAAAGAACGCGAGCGACAGTAAATTAGGTGCGCATATGAA			
omra Rev	TATCCTCCTTAG			
enr P Fund	TTTCTGCCTCGGAGGTATTTAAACAATGAATAAAATCCTGTGTAGGCTGG			
<i>cpxK</i> Fwd	AGCTGCTTCG			
on P Day	AGATGGCGAAGATGCGCGCGGTTAAGCTGCCTATCATGAAGCCATCATA			
<i>cpxk</i> kev	TGAATATCCTCCTTAG			
rso 1 Fund	GGATACTGGATAAGGGTATTAGGCATGCAGAAATGT	GTAGGCTGGAGCT		
rseA I'wa	GCTTCG			
rso 1 Pay	CCAAAGTTGCTTCATTACTGCGATTGCGTTCCTAACA	TATGAATATCCTC		
rsea kev	CTTAG			
Eunnagian Vastan Dui	mong (reatriction sites shown in lower acco)			
Expression vector Prin	Seguence(5 ² , 2 ²)	Clauina Vastan		
Primer Name	Sequence(5 - 5)	Cloning vector		
<i>mzrA Eco</i> RI Fwd		pBAD24		
<i>mzrA Xba</i> I Fwd		pBAD24 and		
We A US DILL DOLL DOLL		pBAD33		
<i>mzrA HinD</i> III Kev		pBAD24		
<i>mzra</i> -FLAG <i>HinD</i> III		pBAD24 and		
Kev		рВАD33 «DAD34		
<i>mzrA Xba</i> l Kev		pBAD24		
phoA Xbal Fwd	A LATCTAGAA LOULIGIIUIGUAAAAUUUU	pBAD24 <i>mzrA</i>		
phoA Sall Rev	ATAIgtegaeTTATTTCAGCCCCAGAGCG	pBAD24 <i>mzrA</i>		
Two Hybrid System P	rimers (restriction sites shown in lower case)			
Primer Name	Sequence $(5^{2}-3^{2})$	Cloning Vector		
<i>mzrA</i> E2HS <i>Xba</i> I Fwd	AGCTGtctagaGCAAATACCTCGCATGTCGC	pUT18C and pKT25		
<i>mzrA</i> E2HS <i>Knn</i> I Rev	GCAGTggtaccCCGAAGCGATGAGAGTTATCCCG	pUT18C and pKT25		
		pUT18, pUT18C and		
<i>envZ</i> E2HS <i>Xba</i> l Fwd	AGCTGtctagacAGGAGGTTGCGCTTCTCGCCACG	pKT25		
		pUT18 pUT18C and		
<i>envZ</i> E2HS <i>Kpn</i> I Rev	GCAGTggtaccCCTTCTTTTGTCGTGCCCTGC	p6116, p61160 und pKT25		
ompR E2HS XhaI Fwd	AGCTGtctagaGCAAGAGAACTACAAGATTCTGG	pUT18 pUT18C		
ompic E2115 Abui 1 wa	noerolouguoonnonnennennonnieroo	pUT18 pUT18C and		
<i>cpxA</i> E2HS <i>Xba</i> I Fwd	ACGTGtctagaGATAGGCAGCTTAACCGCGCG	p6110, p61100, und pKNT25		
		pUT18 pUT18C and		
<i>cpxA</i> E2HS <i>Kpn</i> I Fwd	GCAGTggtaccCGACTCCtCTTATACAGCGGC	nKNT25		
Site-Directed Mutagenesis Primers (alterations shown in lower case)				
Primer Name Sequence (5',3') Cloping Vector				
mzrA A13-28 SDM	CGCATGTCGCTTCGCCAGCTAGCCTCCGCGGTTCG	nBAD24mzr4nho4		
Fwd	CCAGCAAGAGTC	nBAD24mzrA··lacZa		
mzrA A13-28 SDM	GACTCTTGCTGGCGAACCGCGGAGGCTAGCTGGCG	pBAD24m2rAnho4		
Rev	AAGCGACATGCG	nBAD24mzrA··lacZa		
Env7 R3971 Ewd	GCGGCGACAGeGCaCtCACCATTAGCGGCACGGGAT	nFTFnv7c		
$\Box \Pi V \Box \Pi \nabla J / \Box \Pi V U$	Jeggeonenovoeneren indeddenedddal			

	TAGGG	
EnvZ R397L Rev	CCCTAATCCCGTGCCGCTAATGGTGaGtGCgCTGTC	pETEnvZc
	GCCGC	