# Supporting Information

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## SI Materials and Methods

**Cell Culture, Treatments, and Transfection.** Human pancreatic cancer BxPC-3 cells were transfected with the human 1.35-kbp sst2 cDNA or mock vector. Stable BxPC-3 transfectants were cultured in DMEM (LONZA). Human dermal microvascular endothelial HMEC cells were grown in MCBD131 (GIBCO), and human pancreatic neuroendocrine tumor BON cells in DMEM/HAM F12 (50%–50%) (LONZA). All media were supplemented with 10% FCS (LONZA), 2.5 $\mu$g/mL fungizone, 2 mM L-glutamine, and 5 U/mL streptomycin/penicillin (Invitrogen). When indicated, cells were treated with recombinant VEGF (BD Biosciences), or with the somatostatin analog RC-160 (provided by one of us, A.V. Schally), or with the PI3K inhibitor LY294002 (Calbiochem). For characterization of the TSP-1 IRES, BxPC-3 cells were transfected with 2 $\mu$g of the pSL2-TSP-1S, pSL2-TSP-1AS, or pGL3-TSP-1 vector. Cells were harvested in Passive Lysis Buffer (Promega), and extracts were assayed for firefly and *Renilla* luciferase activities using the Dual Luciferase Reporter Assay System (Promega), detected with a Luminoscan Ascent Microplate Luminometer (Thermo Labsystems).

**Real-Time Quantitative RT-PCR.** Total RNA (RNeasy Kit Qiagen) was reverse-transcribed (random hexamers) and resulting cDNAs were used in qRT-PCR using a GeneAmp 7000 Sequence Detection System and SYBR green (Applied Biosystems). Primer sequences (Invitrogen) were as follows: TSP-1-forward, 5′-GTAACCCCTGCACGGATGG-3′; reverse, 5′-TGCACTTGGCGTTCTTGTTG-3′; VEGF-forward, 5′-CCAATCGAGACCCTGGTG-3′; reverse, 5′-CACACAGGATGGCTTGAAGA-3′; and 18S-forward, 5′TGCATGGCCGTTCTTAGTTG-3′; reverse, 5′-TGGCTGAACGCCACTTGTC-3′. PCR efficiencies were >90%. Target gene expression was normalized by using 18S expression as an internal control.

**RNA Interference.** One hundred nanomolar of TSP-1, SRIF, and CTR siRNAs were designed (SMARTpools, Dharmacon) and transfected using DharmaFECT1 reagent (Dharmacon).

**Immunoprecipitation and Immunoblotting.** CM was centrifuged to remove cell debris. Clarified supernatant was used for treatment of HUVEC cells, or concentrated by ultrafiltration (Amicon, Millipore) for immunoprecipitation and/or direct immunoblot analysis. HUVEC cells were treated with 30 ng/mL recombinant VEGF, or with CM for 5 min, and lysed in [50 mM Tris, pH 7.5, 150 mM NaCl, 1 mM EDTA, 1% Triton X-100, 2.5 mM sodium pyrophosphate, 1 mM $\beta$-glycerophophosphate, 1 mM sodium orthovanadate, and protease inhibitor mixture (Roche Diagnostics)]. Protein immunoprecipitation from cell lysates or CM was performed using the appropriate antibody precoupled to protein A/G Sepharose beads. Immunocomplexes or CM were resolved on SDS/PAGE, transferred onto nitrocellulose membranes (Pall Life Sciences). Following probing with a specific primary antibody and a horseradish peroxidase-conjugated secondary antibody, the protein bands were detected by enhanced chemiluminescence (Pierce). Primary antibodies used were as follows: TSP-1, phosphotyrosine clone 4G10 and MMP-2 (Calbiochem), VEGFR2/Flk-1, and VEGF (Santa Cruz Biotechnologies).