## Structural Perturbations Induced by the α-Anomer of the Aflatoxin B<sub>1</sub> Formamidopyrimidine Adduct in Duplex and Single-Strand DNA

## **Supporting Information**

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Nucleotide	Atom	Chemical Shift (ppm)	Nucleotide	Atom	Chemical Shift (ppm)
$C^1$	H1'	5.81	$A^4$	H1'	6.27
$C^1$	H2'	2.02	$A^4$	H2'	2.69
$C^1$	H2"	2.40	$A^4$	H2"	2.51
$C^1$	H3'	4.53	$A^4$	H3'	4.62
$C^1$	H4'	3.92	$A^4$	H4'	4.17
$\mathbf{C}^1$	Н5	5.54	$A^4$	H5'	4.00
$C^1$	H5'	3.58	$A^4$	H5"	3.96
$C^1$	H6	7.48	$A^4$	H8	8.29
$T^2$	H1'	6.19	AFB <sup>5</sup>	H5	5.72
$T^2$	H2'	2.21	AFB <sup>5</sup>	H5B	5.71
$T^2$	H2"	2.37	AFB <sup>5</sup>	H6a	6.13
$T^2$	H3'	4.71	AFB <sup>5</sup>	H8	5.66
$T^2$	H4'	4.28	AFB <sup>5</sup>	H9	6.16
$T^2$	H5'	3.92	AFB <sup>5</sup>	H9a	3.66
$T^2$	H5"	3.89	AFB <sup>5</sup>	OCH <sup>3</sup>	3.76
$T^2$	H6	7.54			
$T^2$	M7	1.70			
X <sup>3</sup>	H1'	5.62			
X <sup>3</sup>	H2'	2.30			

**Table S1.** Resonance Assignments for AFB- $\alpha$ -FAPY Modified 5'-d(C<sup>1</sup>T<sup>2</sup>X<sup>3</sup>A<sup>4</sup>)-3'.

$X^3$	H2"	1.88
X <sup>3</sup>	H3'	4.54
X <sup>3</sup>	H4'	4.11
X <sup>3</sup>	H5'	3.82
X <sup>3</sup>	H5"	3.74
$X^3$	H8	8.27

**Figure S1**. Electronic circular dichroism of the AFB- $\alpha$ -FAPY and AFB- $\beta$ -FAPY anomers in oligodeoxynucleotides. The duplex 5'-d(CTATXATTCA)-3'•5'-d(TGAATCATAG)-3' (panel A) was compared to 5'-d(CTATXATTCA)-3' (panel B). Unmodified oligodeoxynucleotides (solid black line) were compared to oligodeoxynucleotides containing the AFB- $\beta$ -FAPY (broken black line) and AFB- $\alpha$ -FAPY (solid red line) adducts.



**Figure S2.** UV spectra of AFB-FAPY modified 5'-d(CTATXATTCA)-3'. The spectra of the  $\alpha$  anomer (—) is compared to that of the  $\beta$  anomer (—) in **panel A**. The absorbance units on the Y-axis are normalized to the maximum absorbance of 1.0. The spectrum of the  $\alpha$  anomer (—) is compared to that of the chromatographically inseparable shoulder preceding the main  $\alpha$  anomer peak (—) in **panel B**.



Figure S3. NOESY assignments of purine H8 and pyrimidine H6 protons to

deoxyribose H1' protons for the AFB- $\alpha$ -FAPY modified duplex 5'-d(CTATXATTCA)-3'•5'd(TGAATCATAG)-3'. **Panel A:** The connectivity for the modified strand was broken between T<sup>4</sup> H1' and A<sup>6</sup> H8. **Panel B:** The connectivity for the complementary strand was broken between C<sup>16</sup> H6 and A<sup>17</sup> H1'. The intra-nucleotide purine H8 and pyrimidine H6 to deoxyribose H1' cross peaks are labeled.



**Figure S4. Upper Panel.** NOE connectivity for the Watson-Crick base paired amino protons of the AFB-α-FAPY modified duplex. The cross peaks are: a, AFB OCH<sub>3</sub>→X<sup>5</sup> H3; b, C<sup>16</sup> H41→X<sup>5</sup> H3; c, X<sup>5</sup> H21→X<sup>5</sup> H3; d, X<sup>5</sup> H22→X<sup>5</sup> H3; e, A<sup>6</sup> H61→X<sup>5</sup> H3; f, C<sup>16</sup> H42→X<sup>5</sup> H3; g, C<sup>9</sup> H42→G<sup>12</sup> H1; h, C<sup>1</sup> H42→G<sup>20</sup> H1; i, A<sup>13</sup> H61→G<sup>12</sup> H1; j, C<sup>9</sup> H41→G<sup>12</sup> H1; k, C<sup>9</sup> H5→G<sup>12</sup> H1; l, AFB OCH<sub>3</sub>→T<sup>4</sup> H3; m, A<sup>3</sup> H61→T<sup>18</sup> H3; n, A<sup>17</sup> H62→T<sup>4</sup> H3; o, A<sup>17</sup> H61→T<sup>4</sup> H3; p, A<sup>3</sup> H2→T<sup>18</sup> H3; q, A<sup>17</sup> H2→T<sup>4</sup> H3; r, A<sup>6</sup> H2→T<sup>15</sup> H3; s, A<sup>14</sup> H61→T<sup>15</sup> H3; t, A<sup>6</sup> H62→T<sup>15</sup> H3; u, A<sup>14</sup> H62→T<sup>7</sup> H3; v, A<sup>14</sup> H61→T<sup>7</sup> H3; w. A<sup>14</sup> H2→T<sup>7</sup> H3; x; A<sup>13</sup> H2→T<sup>8</sup> H3; y, A<sup>13</sup> H61→T<sup>8</sup> H3; z, A<sup>13</sup> H62→T<sup>8</sup> H3. **Lower Panel.** NOE connectivity for the Watson-Crick hydrogen bonded imino protons of the AFB-α-FAPY modified duplex. The data was collected at 500 MHz. using a mixing time of 250 ms. The temperature was 5 °C.



**Figure S5.** <sup>31</sup>P NMR of AFB- $\alpha$ -FAPY in 5'-d(C<sup>1</sup>T<sup>2</sup>A<sup>3</sup>T<sup>4</sup>X<sup>5</sup>A<sup>6</sup>T<sup>7</sup>T<sup>8</sup>C<sup>9</sup>A<sup>10</sup>)-3'•5'd(T<sup>11</sup>G<sup>12</sup>A<sup>13</sup>A<sup>14</sup>T<sup>15</sup>C<sup>16</sup>A<sup>17</sup>T<sup>18</sup>A<sup>19</sup>G<sup>20</sup>)-3' (upper panel) and in 5'-d(C<sup>1</sup>T<sup>2</sup>X<sup>3</sup>A<sup>4</sup>)-3' (lower panel). The phosphate resonance of C<sup>16</sup> was observed at 0.9 ppm (upper panel).



## Figure S6. <sup>1</sup>H NMR of AFB- $\alpha$ -FAPY modified tetramer aromatic

protons. A single formyl proton resonance assigned as the *E* isomer was observed at 5 °C (lower panel). A formyl resonance assigned as the *Z* isomer was apparent at 30 °C (upper panel).



Figure S7. NOE assignments of AFB inter-nucleotide and deoxyribose intra-nucleotide crosspeaks for the AFB- $\alpha$ -FAPY modified 5'-d(C<sup>1</sup>T<sup>2</sup>X<sup>3</sup>A<sup>4</sup>)-3'. The cross peaks are assigned as (a) A<sup>4</sup> H4' $\rightarrow$ A<sup>4</sup> H1' (b) AFB OCH<sub>3</sub> $\rightarrow$ T<sup>2</sup> H1' (c) AFB H9a $\rightarrow$  AFB H9 (d) AFB H9a $\rightarrow$  AFB H6a (e) A<sup>4</sup> H2' $\rightarrow$  A<sup>4</sup> H1' (f) A<sup>4</sup> H2" $\rightarrow$  A<sup>4</sup> H1' (g) T<sup>2</sup> H2" $\rightarrow$ T<sup>2</sup> H1' (h) T<sup>2</sup> H2' $\rightarrow$ T<sup>2</sup> H1' (i) C<sup>1</sup> H2' $\rightarrow$ C<sup>1</sup> H1' (j) C<sup>1</sup> H2" $\rightarrow$ C<sup>1</sup> H1' (k) AFB OCH<sub>3</sub> $\rightarrow$  AFB H5 (l) X<sup>3</sup> H5" $\rightarrow$  X<sup>3</sup> H1' (m) X<sup>3</sup> H2' $\rightarrow$  AFB H5 (n) X<sup>3</sup> H2" $\rightarrow$  AFB H5 (o) X<sup>3</sup> H2" $\rightarrow$  X<sup>3</sup> H1' (p) X<sup>3</sup> H2' $\rightarrow$  X<sup>3</sup> H1'.

