Supporting Information

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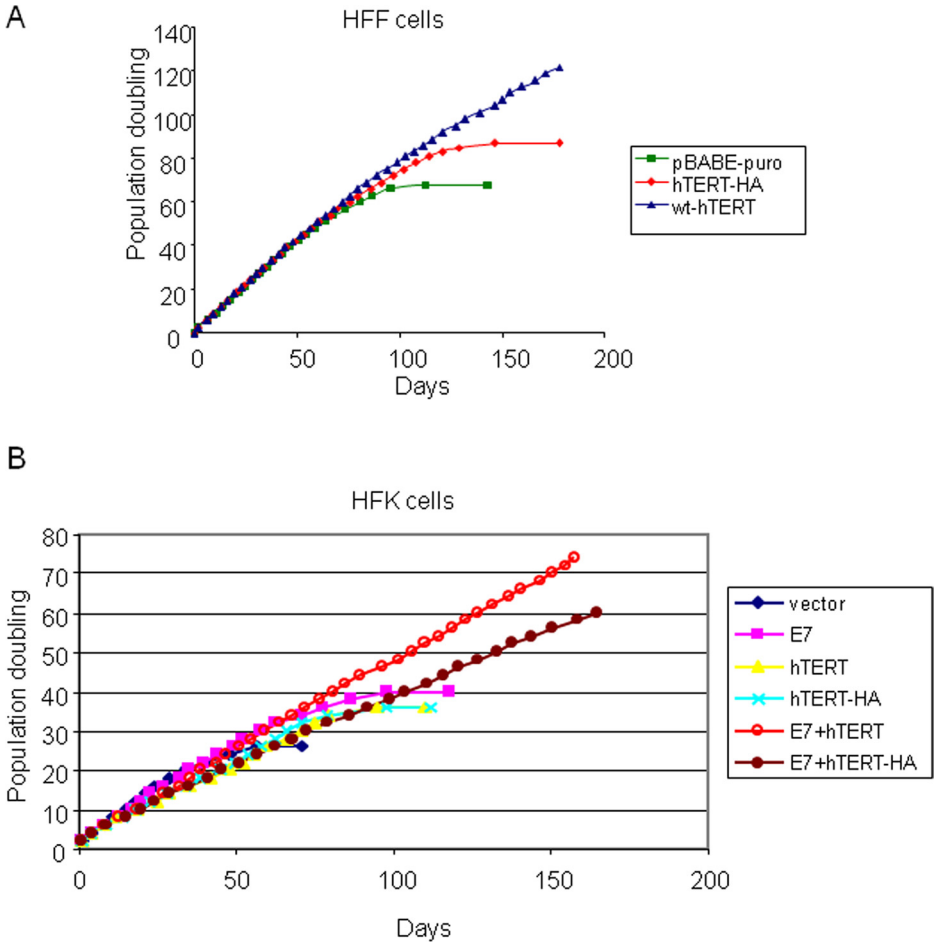

Fig. S1. hTERT-HA and HPV E7 oncoproteins are sufficient to immortalize primary HFK cells. Primary HFF (*A*) and HFK (*B*) were transduced with the indicated pLXSN-based retroviruses with vector, or E7, and pBABE retroviruses expressing vector, wt hTERT, or hTERT-HA. Retrovirus-infected cells were selected in puromycin (2 μg/mL) for HFF, in G418 (50 μg/mL)/puromycin (2 μg/mL) for HFK for 3–5 days. Resistant colonies were pooled and passaged every 3–4 days (1:4 ratio for HFK, 1:8 for HFF). The number of cell doublings calculated and plotted versus time in culture. Cultures that did not proliferate and expand in 20 days for HFK and 30 days for HFF were considered senescent and were terminated at the indicated times. Confirming the previous findings, hTERT-HA fails to immortalize HFF (*A*). However, this construct is sufficient to immortalize HFK together with HPV16 E7 (*B*). These experiments were repeated three times with similar results.

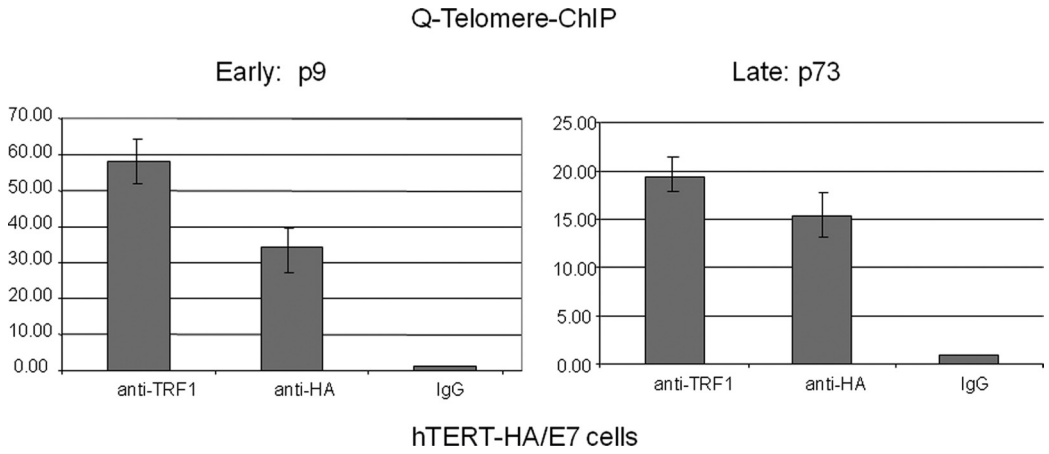

Fig. S2. hTERT-HA associates with telomere sequences in HFK cells immortalized with hTERT-HA and HPV E7. The early (p9, *Left*) and late (p73, *Right*) passages of HFK cells immortalized with hTERT-HA and HPV E7 were used for quantitative telomeric DNA ChIP assays as described in *Materials and Methods*. Antibody against TRF1 (telomere repeat binding factor 1) and normal IgG were used for positive and negative controls, respectively. The signal from IgG was set to 1. hTERT-HA was associated with telomere sequences in the both early and late passages of human HFK cells immortalized with hTERT-HA and E7.