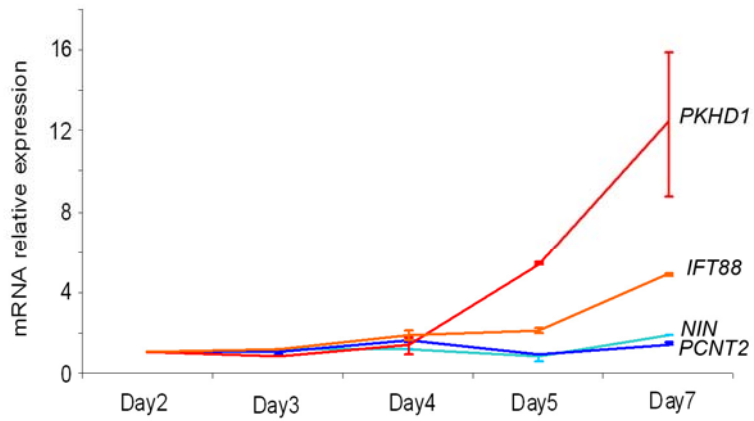


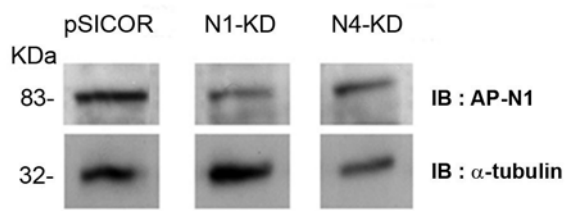
**Supplementary Table: Oligonucleotide primers for real-time RT- PCR.**

<b>Gene</b>	<b>Specie</b>	<b>Forward</b>	<b>Reverse</b>
<i>NPHP1</i>	canine	AGCAGGAGGGGAAGAAGT	TGGAAGAGTGTGGAAGGC
<i>INVS/NPHP2</i>	canine	CATGAAGTGATCCAGTTCATGTTG	GGCGGCGATGTCCTGTAT
<i>NPHP3</i>	canine	TCTCTGTGGAGTAACGCTGTCA	ATCCCATTGTCTCACACATTC
<i>NPHP4</i>	canine	ATAGTTCGCTGGGCTGTT	CTGGCAGGGGTACTTTA
<i>NPHP4</i>	human	CCGTGTACCTTGGAAGT	CCACAGTCAGGCAGAGCA
<i>CEP290/NPHP6</i>	canine	ATGGAGCGACAGCTTAGGAA	TAAATCGTTGCAAACGTCCA
<i>GLIS2/NPHP7</i>	canine	GCCAAGTGTAACCAGCTCTTTGA	CCCTCCCAGTGACAGCAGTAC
<i>RPGRIP1L/NPHP8</i>	canine	TTGAAGCAGTGACCCAGAAGA	TGGTGCCATAGGCAATATCCT
<i>NEK8/NPHP9</i>	canine	GGTGCCTTCGGGATTGTG	CCACTGGGATCTGCTTGATGA
<i>PCNT2</i>	canine	AGGCTTCAGGAAAGCTCTGATTTATC	GTGGTCGTCTTCTTATCAGCTTTG
<i>NIN</i>	canine	CCATCTGCATCTACGCCATACA	CGAAAGCCAATGGTACTTGTC
<i>PKHD1</i>	canine	TGAAATACACTCAGGTGTCTCT	CAGGCATCTCATGAATAAACC
<i>IFT88</i>	canine	GATGGGAAGACCAATGACAGGG	TCAAATGCAGAGCCTCTCAA
<i>GAPDH</i>	canine	AAGGTAGTGAAGCAGGCA	GGTGGAAGAGTGGGTGTC

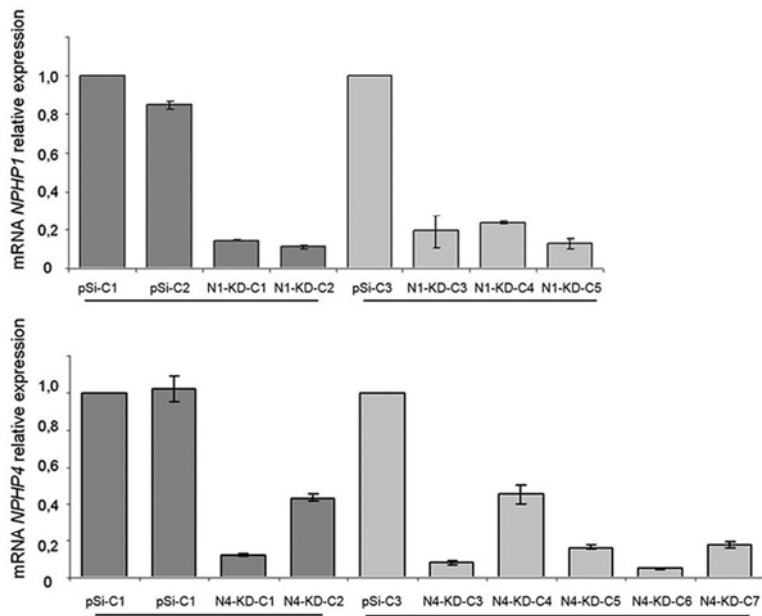


**Suppl. Figure 1: Expression analysis of centrosomal and ciliary genes during cell polarization.** Quantitative RT-PCR analysis of centrosomal genes (*NIN*, encoding ninein and *PCNT2*, pericentrin) and ciliary genes (*PKHD1* and *IFT88*) during time-course experiments performed on MDCK cells.

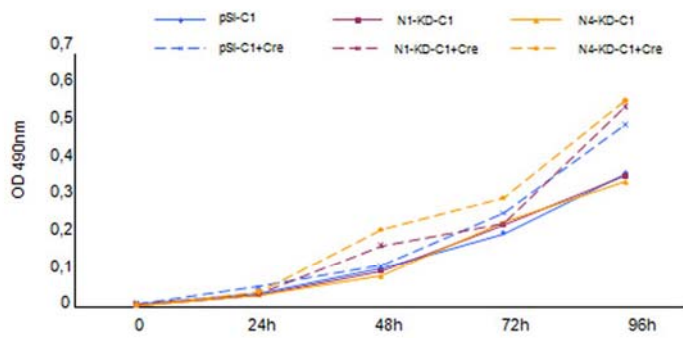
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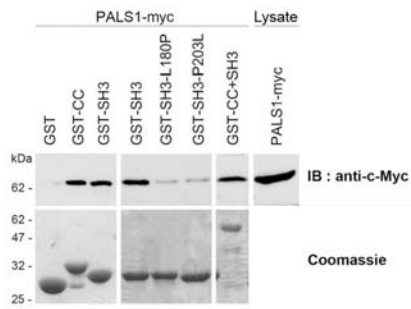
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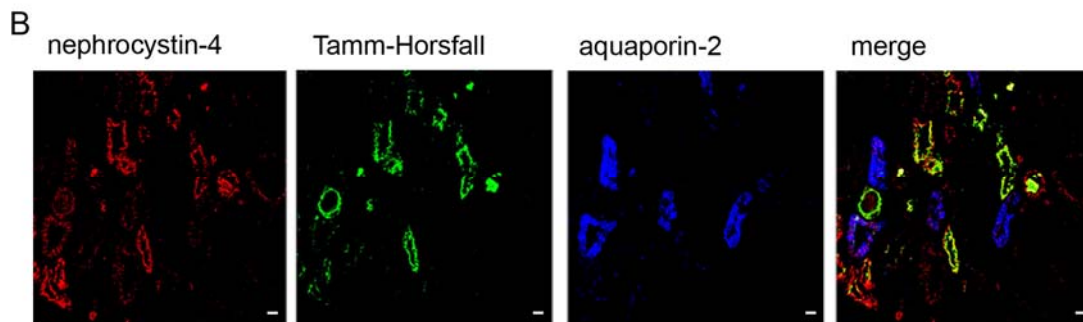
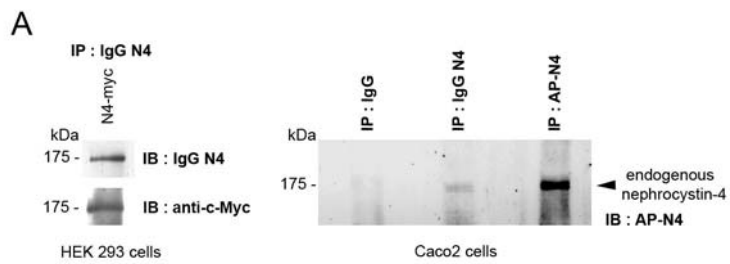
**Suppl. Figure 2: Analysis of *NPHP1* and *NPHP4* expression in polyclonal and clonal knockdown cell lines. (A) Western blot analysis of nephrocystin-1 expression level in polyclonal control pSICOR, N1-KD and N4-KD cell lines. (B) Quantitative RT-PCR analysis of *NPHP1* and *NPHP4* gene expression in control (pSi-C1 to 3), *NPHP1* knockdown (N1-KD-C1 to 5) and *NPHP4* knockdown (N4-KD-C1 to 7) clones, respectively. As clones were selected in two independent experiments, the clones which are compared are underlined.**



**Suppl. Figure 3: NPHP knockdown cells have no defect in cell proliferation.** Control and NPHP-depleted cells ( $5 \cdot 10^3$  cells), before and after Cre infection, were seeded in triplicate in 96-well plates. At each time point, number of cells was quantified using CellTiter 96® Aqueous Cell Proliferation Reagent (Promega) following the manufacturer's instructions and OD490nm was measured using a spectrophotometer microplate reader (Thermolab systems).



**Suppl. Figure 4: Nephrocystin-1 interacts with PALS1 via its coiled-coil and SH3 domains.** *In vitro* pull down assay of PALS1-myc expressed transiently in HEK293T using GST fusion protein of the CC (GST-CC) and SH3 domains (GST-SH3) of nephrocystin-1 and GST alone. Bound protein was revealed with anti-c-Myc antibody and coomassie staining revealed the amount of GST fusion proteins used.



**Suppl. Figure 5: Characterization of the mouse monoclonal anti-nephrocystin-4 antibody.** (A) Immunoprecipitation of overexpressed (N4-myc, left panel) or endogenous nephrocystin-4 (right panel) protein with mouse monoclonal (IgG N4) or rabbit polyclonal (AP-N4) anti-nephrocystin-4 antibody in HEK293 cells and caco2 cells, respectively. (B) Co-labelling of human kidney section with IgG N4, anti-Tamm-Horsfall (Argen) (thick ascending loop of Henle and distal tubules) and anti-aquaporin2 (Santa Cruz) (collecting ducts) antibodies. Scale bars are 20 $\mu$ m.