

Supplemental Material:

MATERIALS AND METHODS

RNA interference

Mouse CHOP siRNA (ON-TARGETplus SMARTpool Cat# L-062068-00) and control siRNA (ON-TARGETplus Control) were purchased from Dharmacon. Transfection reagent designed for 3T3-L1 cells was also purchased from Dharmacon (Dharmafect 3). All transfection procedures were as per manufacturer's protocol. Cells were grown at a density of 2.5×10^5 cells/ml in 35 mm dishes and treated with 100 nM CHOP siRNA with DharmaFect transfection reagent in penicillin/streptomycin-free growth media 2 days prior to differentiation (day -2). Differentiation media was added on day 0 (48 hrs post-initial siRNA transfection). Every two days cells were re-transfected with CHOP siRNA. Cells were treated with 100 nM thapsigargin on day -2, day 0, and day 2 for 48 hrs to induce ER stress and CHOP expression. Total protein lysates were collected and separated on a 10% SDS polyacrylamide gel. On day 0 and 2, 10 mM 4-PBA was added to the media.

All other procedures were as described in the manuscript text.

FIGURE LEGENDS

Figure S1. Treatment of 3T3-L1 cells with tauro-ursodeoxycholic acid (TUDCA) blocks 3T3-L1 differentiation in a dose-dependent manner. **A)** Confluent day 0 3T3-L1 cells were cultured in differentiation media with increasing concentrations of TUDCA (0.1, 1, or 2 mg/ml). On day 4, cells were fixed and stained with Oil red O. Representative images of Oil red O stained cells are shown for each condition, with arrows indicating differentiated adipocytes. **B)** *Quantification of Oil red O indicated a significant decrease in lipid droplets with TUDCA treatment.* Following extraction and collection of Oil red O in isopropanol, absorbance was measured at 510 nm. Data are presented as the mean values +/- standard deviation (n=3). (**p<0.01*)

Figure S2. 4-PBA treatment inhibits 3T3-L1 differentiation through a mechanism independent of CHOP. **A)** *CHOP protein expression is repressed in CHOP siRNA transfected 3T3-L1 cells when treated with 100 nM thapsigargin or 10 mM 4-PBA.* Cells were treated with 100 nM CHOP siRNA in penicillin/streptomycin-free growth media (day -2). Differentiation media was added on day 0 (48 hrs post-initial siRNA transfection) and post-stimulation media was added on day 2. Cells were transfected with CHOP siRNA on day -2 and 0 and treated with 100 nM of the ER stress inducer thapsigargin (Tg) on day -2, day 0, and day 2 for 48 hrs to induce CHOP expression. On day 0 and 2, 10 mM 4-PBA was added to the media. Total protein lysates were collected at various time points during the experiment and run on a 10% SDS polyacrylamide gel. Western blotting was performed and the membrane was probed with an anti-CHOP antibody as well as anti- β -actin as a loading control. **A-** Day 0 Control, **B-** Day 0

after 48 hr 100 nM Tg, **C**- Negative Control siRNA + 100 nM Tg, **D**- Day 0 CHOP siRNA (48 hrs), **E**- Day 0 CHOP siRNA + 100 nM Tg (48 hrs), **F**- Day 2 CHOP siRNA (96 hrs), **G**- Day 2 CHOP siRNA (96 hrs) + 100 nM Tg (48 hrs), **H**- Day 2 48 hrs 4-PBA treated, **I**- Day 2 CHOP siRNA (96 hrs) + 48 hrs 4-PBA treated, **J**- Day 4 Control, **K**- Day 4 after 48 hrs 100 nM Tg, **L**- Day 4 CHOP siRNA treated day -2 to day 2 + 48 hrs 100 nM Tg, **M**- Day 4 96 hrs 4-PBA treated, **N**- Day 4 CHOP siRNA treated day -2 to day 2 + 96 hrs 4-PBA treated. **B**) *siRNA knock-down of CHOP did not rescue the cells from the anti-adipogenic effects of 4-PBA.* 3T3-L1 cells were seeded at 70% confluence and transfected with CHOP siRNA. Two days later on day 0, cells were transfected again and differentiation media was added in the presence or absence of 10 mM 4-PBA. On day 2, cells were washed and post-stimulation media with or without 10 mM 4-PBA was added. Cells were fixed and stained with Oil red O on day 4. Images were taken at 20X magnification. **C**) Oil red O quantification indicated a significant decrease in lipid content in 4-PBA and CHOP siRNA+4-PBA treated cells as compared to untreated control cells. However, there was no difference in adipogenic differentiation with 4-PBA when CHOP was knocked down versus non-transfected cells ($*p<0.001$).

Figure S3. Salubrinal, a selective inhibitor of eIF2 α dephosphorylation, blocks adipogenesis in a dose-dependend manner. **A**) 3T3-L1 cells at day 0 were stimulated to differentiate in the presence or absence of 10, 50 or 100 μ M salubrinal. On day 4, the cells were fixed and stained with Oil red O. Representative images are shown. **B**) *Salubrinal dose-dependently inhibited lipid accumulation.* Oil red O staining was quantified and a significant decrease in staining intensity was observed with increasing dose of salubrinal ($* p<0.05$,

**** $p < 0.01$**). **C)** Western blotting confirmed that 100 μ M salubrinal decreases splicing of XBP1 in differentiating day 4 adipocytes.

Figure S4. Rosiglitazone can partially rescue the differentiation of 3T3-L1 cells in the presence of 4-PBA by enhancing GRP78/GRP94 expression levels. **A)** Day 0 cells were treated with 4 μ M rosiglitazone in the absence or presence of 10 mM 4-PBA in differentiation media. Cells were re-supplemented with these drugs on day 2. On day 5, cells were fixed and stained with Oil red O. **B)** 4-PBA alone and the rosiglitazone/4-PBA co-treatment resulted in a significant decrease in Oil red O staining (*** $p < 0.001$**) though co-treatment allowed for some adipogenic conversion to occur. **C)** *Rosiglitazone and 4-PBA co-treatment allows for some adiponectin secretion by 3T3-L1 cells that was completely blocked by 4-PBA treatment alone.* Media was collected on day 5 of differentiation and an ELISA was performed to detect secreted adiponectin protein. Media from day 0 untreated and undifferentiated 3T3-L1 cells was used as a negative control. Data represent the mean values \pm standard deviation. (*** $p = 0.0003$** , **** $p \leq 0.0001$**) **D)** *Rosiglitazone exerts its pro-adipogenic effect in the presence of 4-PBA by enhancing GRP78 expression in differentiating adipocytes.* Western blotting using lysates collected on day 5 of differentiation with various treatments. Rosiglitazone treatment during 5 days of differentiation (day 0-5) increased GRP78 expression. 4-PBA blocked GRP78 expression, while rosiglitazone and 4-PBA co-treatment enhanced GRP78 expression.

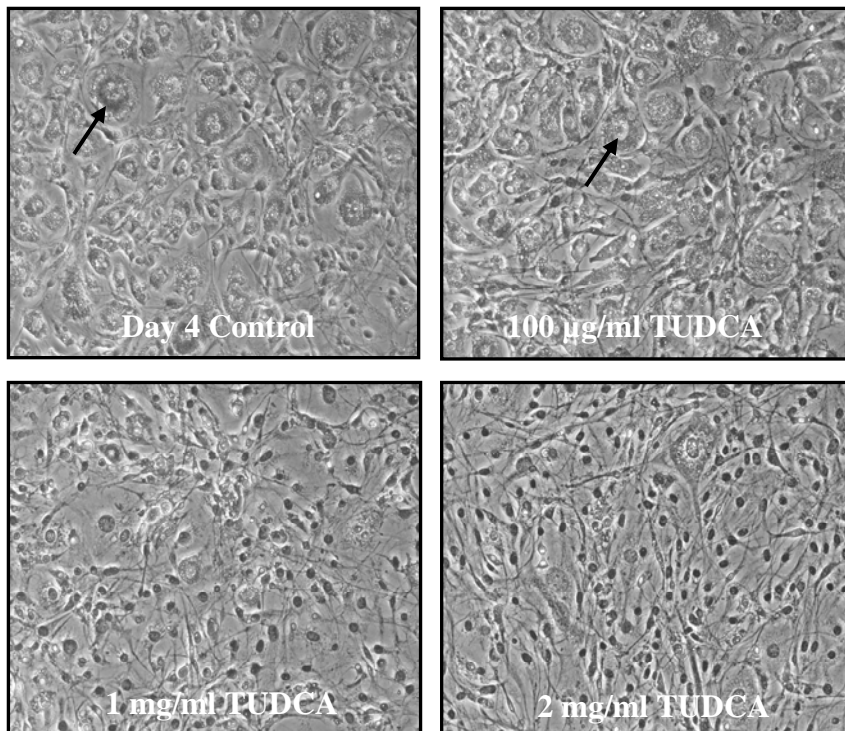
Figure S5. Markers of inflammation or oxidative stress were not altered with 4-PBA treatment. **A)** 3T3-L1 cells treated with varying doses of 4-PBA over 48 hours did not change the expression of NF κ B or peroxiredoxin-1 on Western blots. Expression of β -catenin, a known

inhibitor of adipogenesis was also not affected by 4-PBA. **B)** Western blot analysis of the WAT and liver lysates from the 4-PBA supplemented mice showed no change in NF κ B or peroxiredoxin1 expression as compared to the non-supplemented group. β -actin was used as a loading control.

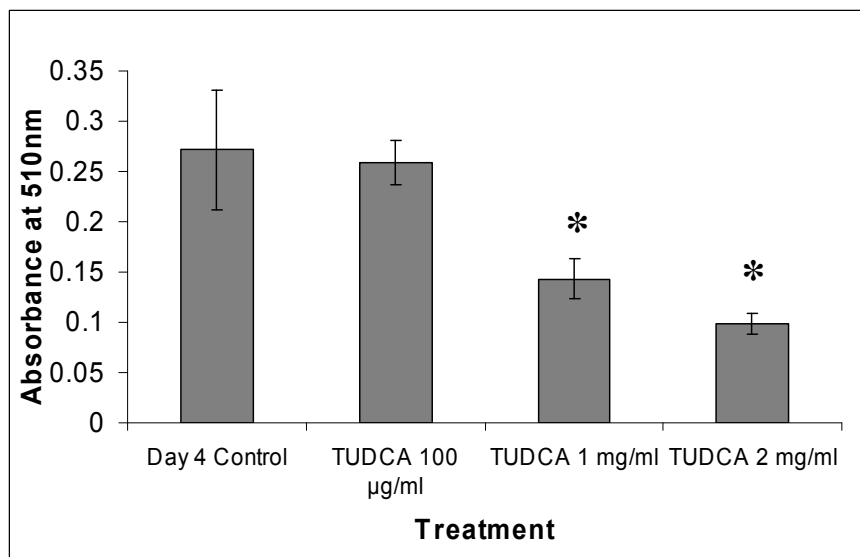
Figure S6. XBP-1 deficient MEFs show reduced expression of adipogenic markers. Protein was collected on days 0, 2, or 4 of differentiation following stimulation of XBP-1 deficient or wildtype MEFs with adipogenic media. Protein from 3T3-L1 cells was collected as an additional control. Western blotting was performed to detect PPAR γ , C/EBP α isoforms, as well as β -actin as a loading control. MEFs deficient in XBP-1 express reduced adipogenic markers as compared to wildtype control MEFs.

Supplementary Figure S1.

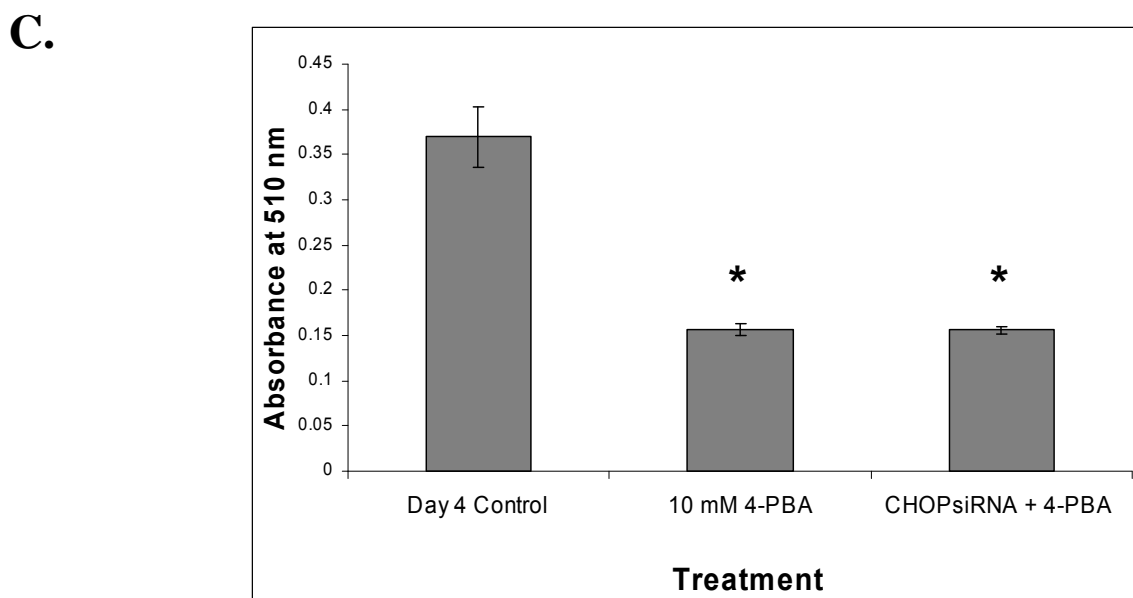
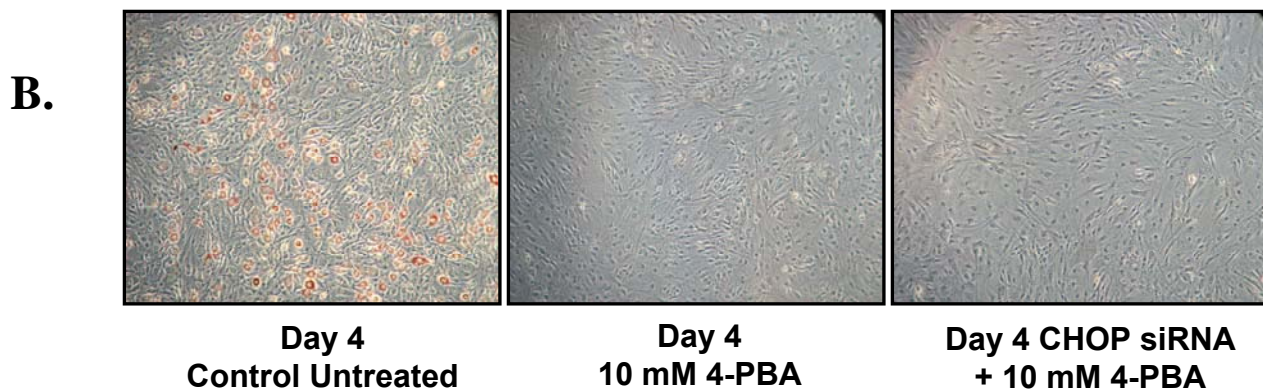
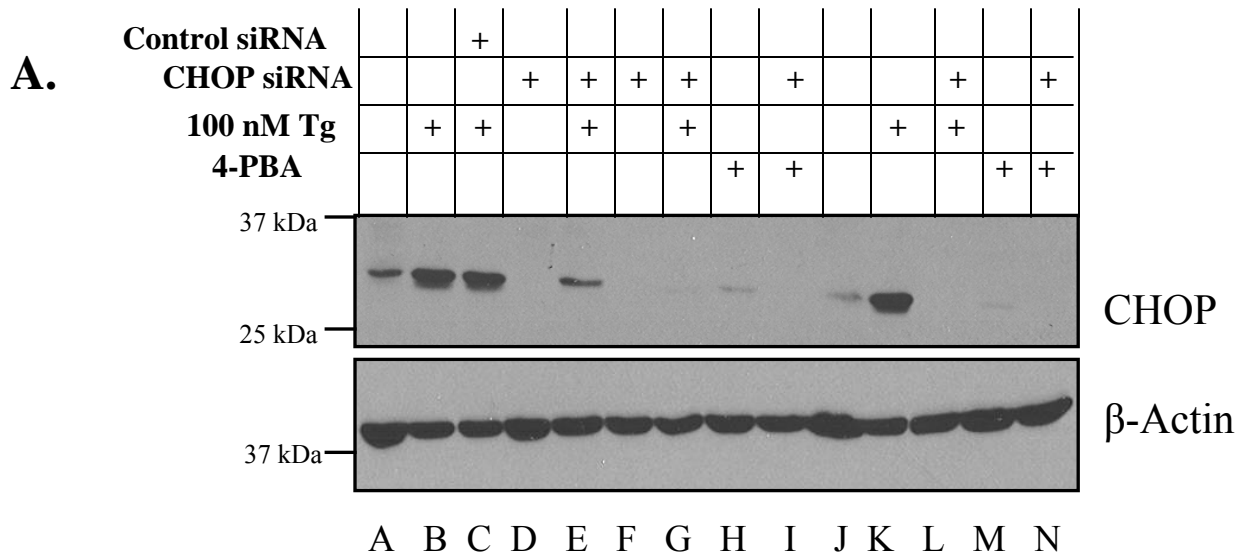
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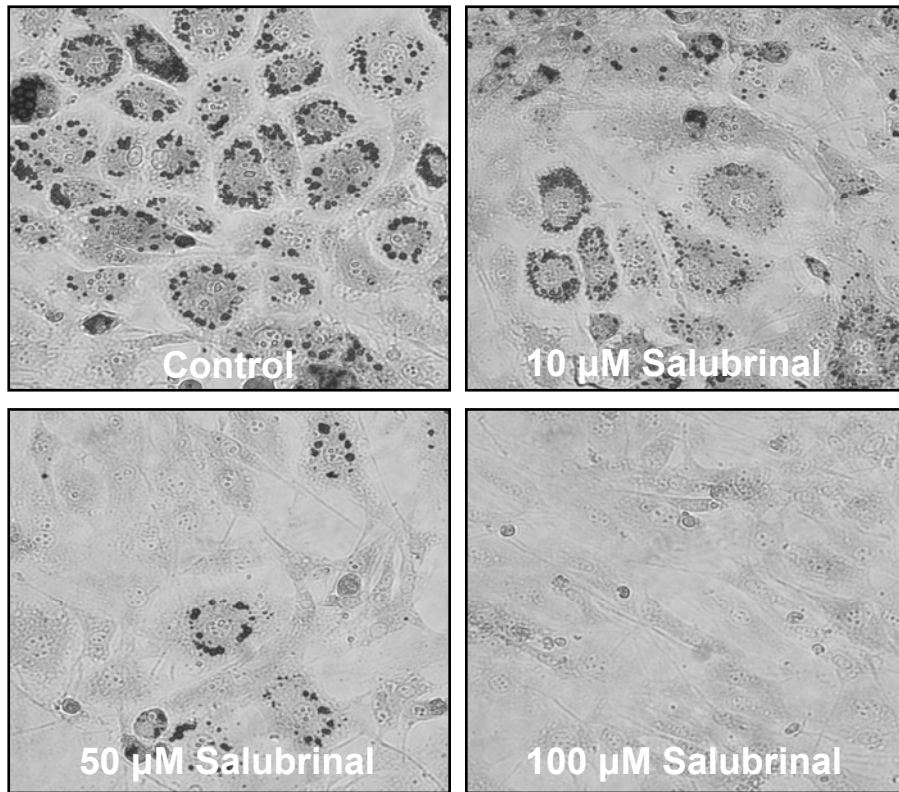
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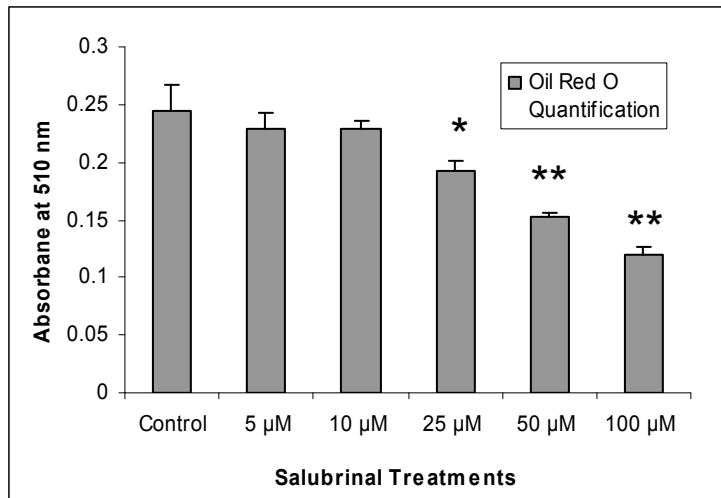
Basseri S et al. The chemical chaperone 4-phenylbutyrate inhibits adipogenesis by modulating the unfolded protein response.

Supplementary Figure S3.

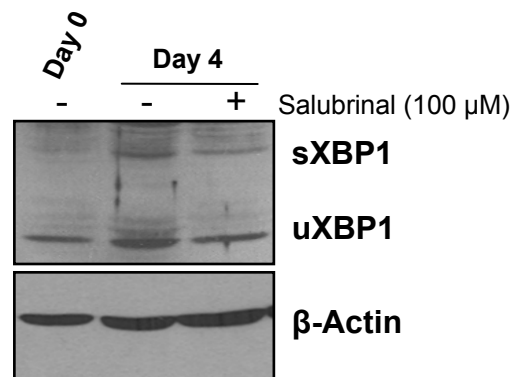
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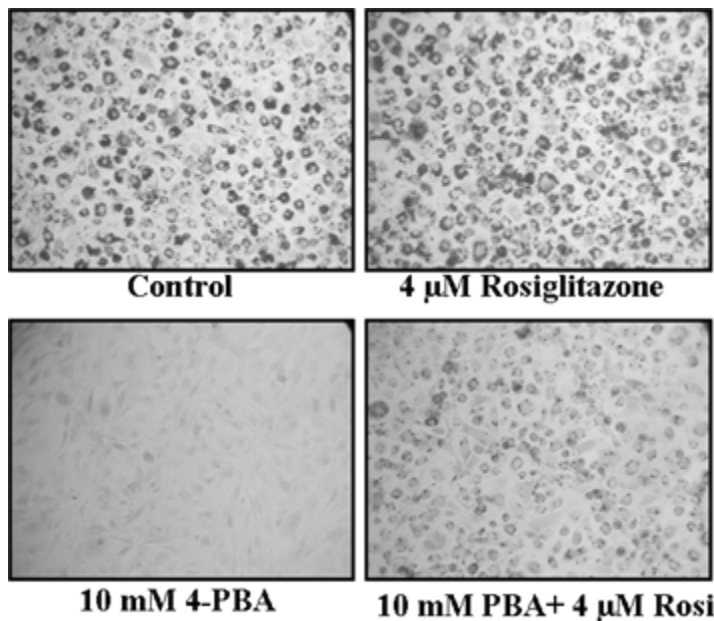
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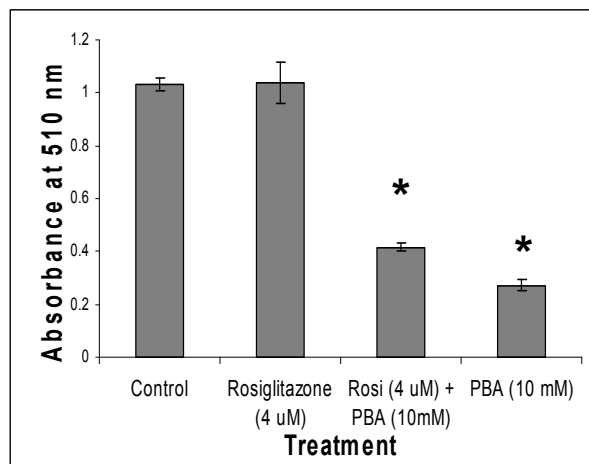
Basseri S et al. The chemical chaperone 4-phenylbutyrate inhibits adipogenesis by modulating the unfolded protein response.

Supplementary Figure S4.

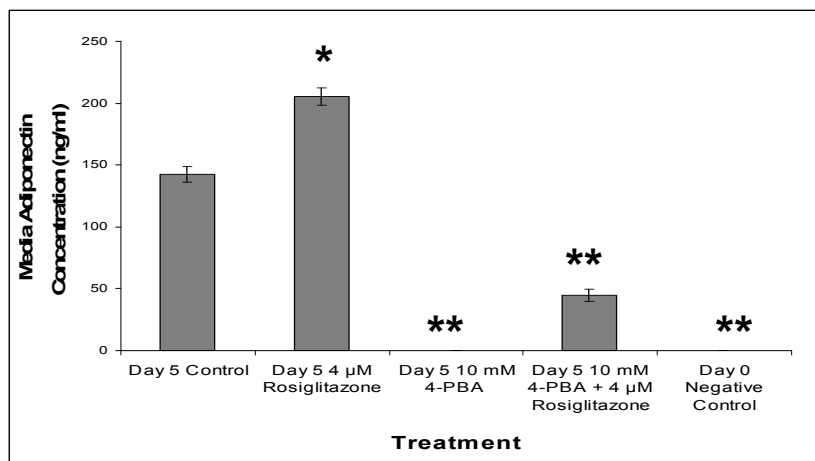
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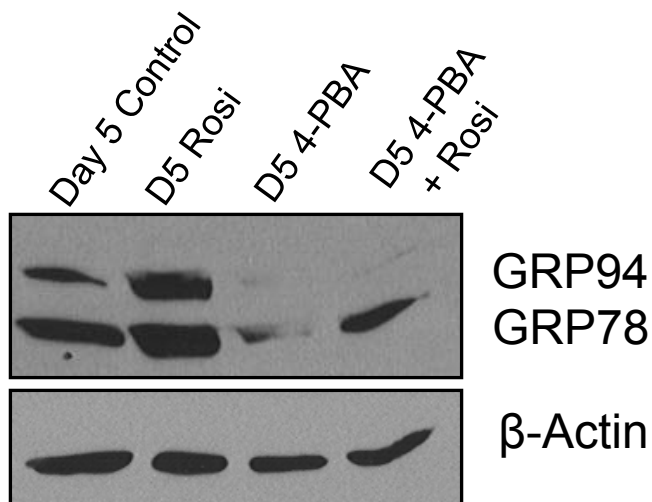
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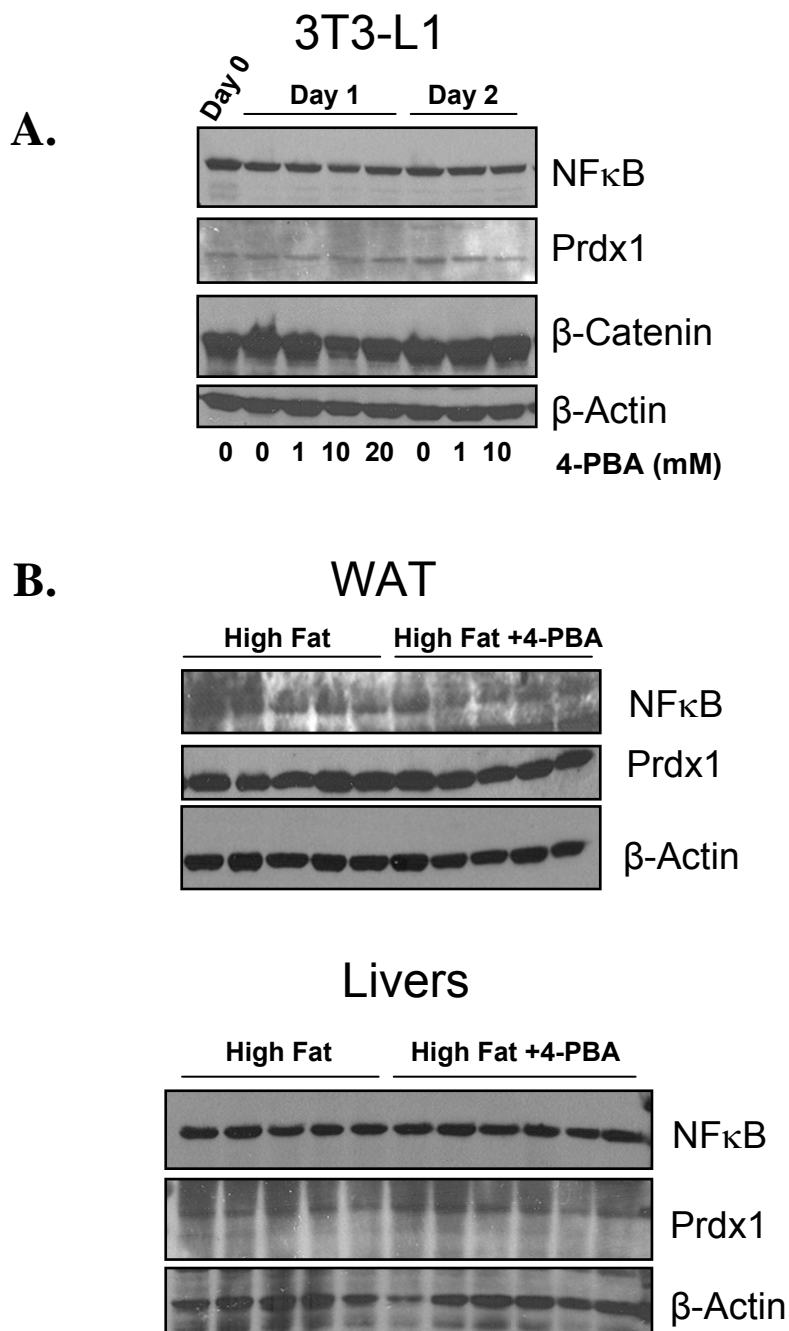


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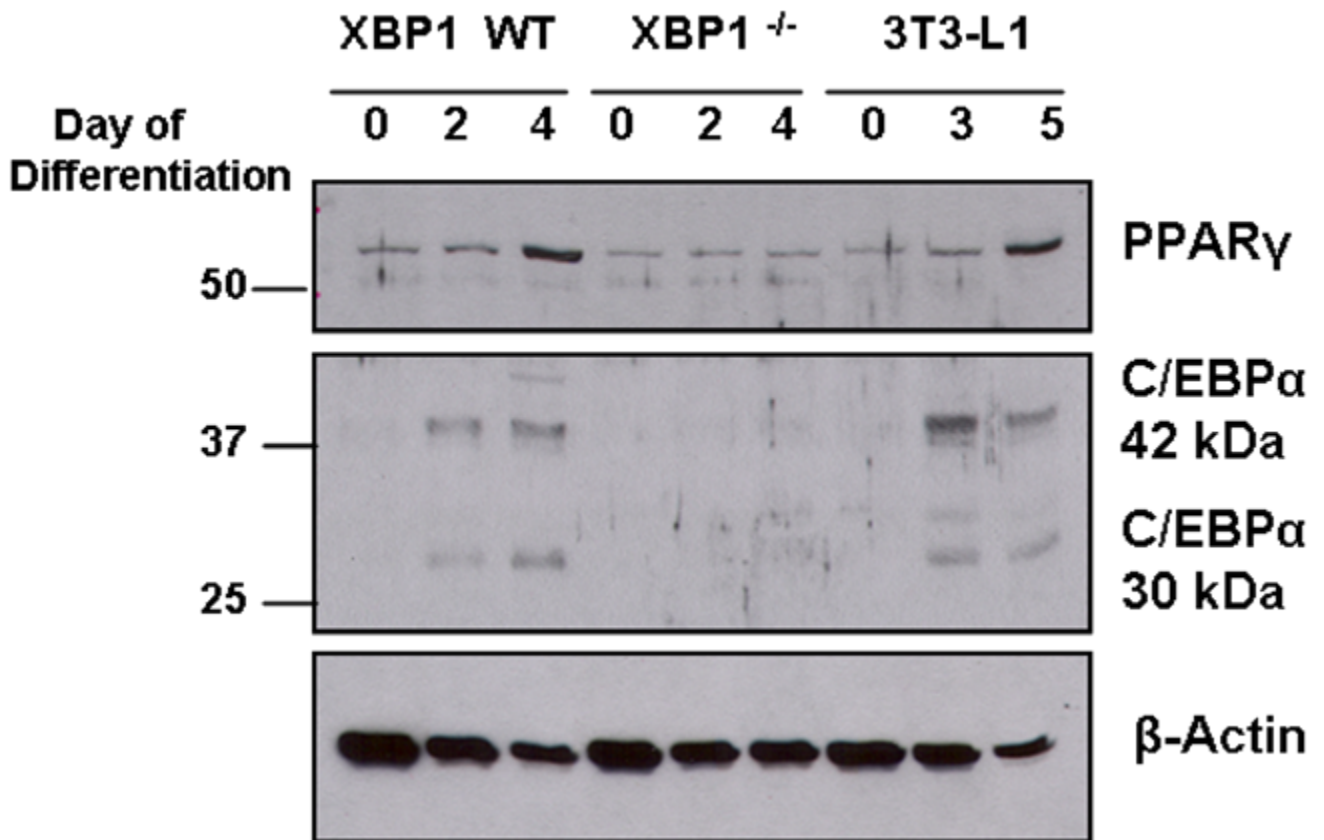


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Supplementary Figure S5.



Supplementary Figure S6.



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