SUPPLEMENTARY INFORMATION

Mathematical derivation Kinetochore microtubule (kinetochore-microtubule) dynamics.

Calculating the detachment rates and the mean lifetime of kinetochore-microtubules from measured kinetochore-microtubule half-lives

After photoactivating a $\sim 2\mu m^2$ area on the spindle the fluorescence decays at a double exponential decay rate (Supp. Figure 1A, B) that we can fit to the following equation:

$$F(t) = A_1 e^{-k_1 t} + A_2 e^{-k_2 t}$$
 (Eq. 1),

where F(t) is the observed fluorescence at time t, A_1 and A_2 represent less (nonkinetochore-microtubule) and more (kinetochore-microtubule) stable microtubule populations with decay constants of k_1 and k_2 , respectively [1-3].

Therefore, we can calculate the probability that a single microtubule will detach from a kinetochore within time (T) to be

$$\int_0^T k_2 e^{-k_2} \partial t \quad \text{(Eq. 2),}$$

from the definition of the half-life $t^{1/2}$ for an exponential decay process therefore,

$$\frac{1}{2} = \int_{0}^{1/2} k_2 e^{-k_2} \partial t \quad \text{(Eq. 3),}$$

where $t^{1/2}$ is experimentally estimated as the time taken for half of the kinetochoremicrotubules with a photoactivated fluorescent label to detach from the kinetochore

From equation 3,

$$k_2 = \frac{\ln(2)}{t^{1/2}}$$
 (Eq. 4).

We can then calculate the detachment rate (r_d) of microtubules from kinetochores, which is directly proportional to the kinetochore-microtubule occupancy (n). Thus,

$$r_d = n \frac{\ln(2)}{t^{1/2}}$$
 (Eq. 5)

The mean-lifetime (τ) of an individual microtubule attached to kinetochore is

$$\tau = \frac{n}{r_d} = \frac{t^{1/2}}{\ln(2)}$$
 (Eq. 6)

Calculating kinetochore-microtubule occupancy from kinetochore-microtubule half-lives

In order to calculate kinetochore-microtubule occupancy for a given experimentally measured half-life of a kinetochore-microtubule, high resolution information detailing kinetochore-microtubule occupancy is required. The most wellstudied kinetochore microtubule interface is in PtK₁ cells where serial section electron microscopy was used to measure the kinetochore-microtubule occupancy during different phases of mitosis (at metaphase, n = 24.3, ref. 4). Moreover, in these cells, kinetochoremicrotubule half-lives have also been measured at the different stages of mitosis (at metaphase, $t^{1/2} = 7.4$ min., ref. 2). It was also shown that kinetochores are saturated at a *saturation occupancy* (*S*) of 35 microtubules [4].

We assume that, at metaphase, kinetochore-microtubule occupancy is at a steadystate so that the attachment rate (r_a) and detachment rate (r_d) of microtubules from kinetochores are equal:

$$r_a = r_d$$
 (Eq. 7),

We also assume that kinetochore microtubule attachment r_a is directly proportional to the number of unoccupied sites at the kinetochore. Therefore,

$$r_{a} = K_{a}(S-n)$$
 (Eq. 8),

where K_a is the rate of kinetochore-microtubule attachment if there was only one single unoccupied site at the kinetochore and we assume that it remains constant throughout mitosis.

From equations 7 and 8,

$$K_a(S-n) = r_d = n \frac{\ln(2)}{t^{1/2}}$$
 (Eq. 9)

Therefore, from equations 5 and 9, the relationship between *n* and $t^{1/2}$ is governed by the following equation:

$$n = \frac{K_a t^{1/2} S}{K_a t^{1/2} + \ln(2)} = S\left(1 - \frac{\ln(2)}{K_a t^{1/2} + \ln(2)}\right)$$
(Eq. 10)

From previous data, if S = 35 (ref. 4), and during metaphase $t^{1/2} = 444$ s. (ref. 2), and n = 24.3 (ref. 4), therefore

 $K_a \approx 3.53 \text{ x } 10^{-3} \text{ s}^{-1}$

SUPPLEMENTARY REFERENCES

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SUPPLEMENTARY FIGURE LEGENDS

Supplementary Figure 1. Cancer cell lines have a functional SAC. Mitotic indices in cell lines that were untreated (blue bars) or treated for 16 hours in the presence of nocodazole. Bars represent mean \pm s.e.m., n > 500 cells, 5 experiments.

Supplementary Figure 2. Kinetochore-microtubule dynamics. (A) Microtubulerelease rates, r_d (s⁻¹), from kinetochores as a function of kinetochore-microtubule half-life (min.). r_d -values from several cancer cell lines at metaphase are denoted as empty circles. Prometaphase values have been omitted for clarity. (B) The mean lifetime, τ (min.), of an individual microtubule attachment at the kinetochore as a function of kinetochoremicrotubule half-life (min.). τ -values from several cancer cell lines at metaphase are denoted as empty circles.

Supplementary Figure 3. RNAi depletion. (A) Total cell lysates from untreated and MCAK-depleted cells were immunoblotted with antibodies specific for MCAK, tubulin and actin as indicated. (B) Total cell lysates from untreated or APC-depleted RPE-1 cells were immunoblotted with antibodies specific for APC and actin as indicated.