Biophysical Journal, Volume 97

Supporting Material

Combining laser microsurgery and finite element modeling to assess cell-level epithelial mechanics

M. Shane Hutson, Jim Veldhuis, Xiaoyan Ma, Holley E. Lynch, P. Graham Cranston, and G. Wayne Brodland

SUPPORTING MATERIAL

Note S1: Correlations with the local cell geometry. The FE model yields a wide range of recoil velocities – even when results are limited to a single angle with a specific set of boundary conditions. This wide range occurs despite a uniform viscosity and uniform cell-edge tensions. It is instead caused by local differences in cell geometry. To illustrate the geometric influences, we present a local approximation to the full finite element matrix equation. For a node at the end of a wounded edge (e.g. node A in Fig 1 of the main text), one can write an equation of dynamic equilibrium with respect to the edge direction (x'):

$$(\eta_{1xx} + \eta_{2xx} + \eta_{3xx})\mathbf{v}_{x0} \approx \gamma(\cos\alpha + \cos\varepsilon) + \langle \sigma_{in} \rangle \delta(d_{AC}\sin\alpha + d_{AE}\sin\varepsilon)/2 \qquad \text{Eqn S1}.$$

The right-hand side is the post-ablation force imbalance at node A due to tension along the two non-ablated edges and the internal stress in cell #3 (approximated by the average internal stress). The geometric parameters are defined in Fig 1 of the main text. The left-hand side is a viscous force where η_{mxx} is the effective drag coefficient for motion along *x'* due to cell *m*. For each cell, the drag coefficient is calculated on the basis of two orthogonal systems of dashpots (1). These dashpot systems define a 2×2 damping matrix in the cell's principle coordinate system

$$\boldsymbol{\eta} = \frac{4 \pi a \, \delta \mu}{n} \begin{bmatrix} \sqrt{I_{\min} / I_{\max}} & 0\\ 0 & \sqrt{I_{\max} / I_{\min}} \end{bmatrix}$$
Eqn S2

where a = 0.682 is an empirical constant, *n* is the number of nodes in the cell and I_{min} and I_{max} are the cell's principle moments of inertia. To find η_{mxx} , the damping matrix is rotated into the frame of the ablated edge ($R^T \eta R$). A primary impact of Eqn S2 is to make η_{nxx} smaller when a cell is extended along *x'*. For Eqn S1, we take the first order approximation in which only node A moves and only in the *x'*-direction.

Eqn S1 can be recast in dimensionless parameters as:

$$G_{xx}v_0 \approx C_{\gamma} + \langle \Sigma_{in} \rangle C_{\Sigma} \qquad \text{where} \quad G_{xx} = (\eta_{1xx} + \eta_{2xx} + \eta_{3xx})/(2\mu\delta) \qquad \text{Eqn S3}$$

$$C_{\gamma} = \cos\alpha + \cos\varepsilon$$

$$C_{\Sigma} = (\chi_{AC}\sin\alpha + \chi_{AE}\sin\varepsilon)/4$$

 G_{xx} , C_{γ} and C_{Σ} respectively capture the influence of cell geometry on viscous damping, the imbalance of cell-edge tensions and the imbalance of internal cell stress. v_0 always increases with C_{γ} and decreases with G_{xx} (Fig S1). Both dependencies reflect increases in v_0 as cells become more elongated in the direction parallel to the ablated edge – which decreases each η_{nxx} and makes the triple-junction angles more acute. The dependence of v_0 on C_{Σ} is much weaker, and is not readily apparent until one includes the viscous damping effects (Fig S1, v_0 versus C_{Σ} / G_{xx}). When all three effects are included, the local approximation accounts for 50-60% of the variance in v_0 .

The variation in G_{xx} has a critical influence on recoil velocity that is often overlooked. For example, Rauzi et al modeled the geometry dependence of v_0 based solely on the imbalance of cell edge tensions in an idealized geometry of stretched hexagonal cells (2). They assumed uniform viscous damping that canceled out of v_0 comparisons. Although they attributed

differences between model and experiments to cell-edge elasticity, we estimate that G_{xx} should actually decrease by a factor of $\sqrt{3}$ over the range of cell shapes considered – accounting for roughly half of their observed differences.

- 1. Brodland, G. W., D. Viens, and J. H. Veldhuis. 2007. A new cell-based FE model for the mechanics of embryonic epithelia. Comp. Meth. Biomech. Biomed. Engr. 10:121-128.
- 2. Rauzi, M., P. Verant, T. Lecuit, and P. F. Lenne. 2008. Nature and anisotropy of cortical forces orienting Drosophila tissue morphogenesis. Nature Cell Biology 10:1401-1410.



FIGURE S1. Correlations between the recoil velocity and local cell geometry. Each row of graphs corresponds to a different far-field stress (as noted). The geometric factors G_{xx} , C_{γ} and C_{Σ} are defined in Eqn S3. The last plot on each row compares the simulated v₀ to that predicted based on Eqn S3 and the local cell geometry (solid line is y = x).



FIGURE S2. Dependence of initial recoil velocity on direction under anisotropic external stress. (A-C) Histograms of cell-edge orientations. The external stresses are as shown for the sample cell patches. Note that $\Sigma < 0$ implies cell edges that are under compression. For (B,C), the cell patch was previously stretched in the vertical direction. (**D-F**) Initial recoil velocities versus direction for cell-edge (\Box) and cell-center (×) wounds. For celledge wounds, the tracked direction was always parallel to the ablated edge.

FIGURE S3. Comparison of the v_0 -distributions for experiments (A) and simulations (B-F) for late dorsal closure when $\Sigma < 0$, i.e. cell edges are under compression: (B) best-matching uniform simulations; (C) simulations with inter-embryo lognormal

1.5 А 1.0 0.5 0.0 В $\Sigma_x = -5.38 - \Delta \Sigma$ 2.0 $= -5.38 + \Delta \Sigma$ 1.5 1.0 0.5 0.0 С 1.0 Probability density 0.5 0.0 D 1.0 0.5 0.0 Е 1.0 0.5 0.0 F 1.5 1.0 0.5 0.0 2 5 3 4 $\nu_{0}/\langle\nu_{0,C}\rangle$

variations in $|\Sigma|$; (**D**) simulations with inter-embryo lognormal variations in all |force/viscosity| ratios; (**E**) simulations with intra-embryo lognormal variations in viscosity; (**F**) non-equilibrium simulations with intra-embryo lognormal variations in the interfacial tension magnitude. Cell-center wounds are in red, cell-edge wounds in grey. $\langle v_{0,C} \rangle$ and $\langle v_{0,E} \rangle$, are marked by the red *C* and grey *E* respectively. The sample cell patch in (B) shows the cell geometry after equilibration at the noted stress $\Sigma_{x,y}$.

Viscoelastic rods along cell edges only									
Fit to:	R^2	$\Phi \Psi_{M}$	$\Phi\xi_{ m M}$	$\Phi \Psi_{K}$	$\Phi \xi_{ m K}$				
mean – st. dev.	.999242	1750	767	27.5	0				
mean	.999653	750	349	10.0	0				
mean + st. dev.	.999447	500	174	6.25	0				

Table S1. Viscoelastic parameters used in the simulations presented in Figure 7.

Viscoelastic rods as a pre-stressed intracellular mesh									
Fit to:	R^2	$\Sigma_{ m mesh}$	$\Phi \Psi_{M}$	$\Phi \xi_{ m M}$	$\Phi \Psi_{K}$	$\Phi \xi_{\mathrm{K}}$			
mean – st. dev.	.999256	3.27	1.00	0.506	2.50×10^{-3}	1.74×10^{-2}			
mean	.999651	3.16	0.625	0.251	1.25×10^{-3}	1.74×10^{-2}			
mean + st. dev.	.999474	2.90	0.375	0.293	1.25×10^{-3}	1.74×10^{-2}			
Double-wound simulation		3.75	0	0	0.497	1.34×10^{-2}			