

Table S1. Bacterial strains and plasmids used in this study

Strain or plasmid	Relevant characteristics	Source or reference
<i>S. gordonii</i> strains		
CH1	Parental strain Challis (sequenced genome)	(13)
DL1	Parental strain Challis	(8)
UB1968	DL1; 266 bp SGO_2006 replaced by 913 bp <i>aad9</i> with no terminator	This study
UB1890	DL1; 1960 bp SGO_2005 replaced by 1158 bp <i>aad9</i> with terminator	This study
UB1964	DL1; 1454 bp SGO_2004 replaced by 1158 bp <i>aad9</i> with terminator	This study
UB1545	<i>hsa::aphA3</i> (Kn ^r)	(7)
UB1360	$\Delta(sspA sspB)::aad9$ (Sp ^r)	(6)
UB1552	<i>hsa::aphA3</i> $\Delta(sspA sspB)::aad9$ (Kn ^r Sp ^r)	(7)
UB2031	DL1; <i>hsa::aphA3 padA::aad9</i> (Kn ^r Sp ^r)	This study
<i>E. coli</i> strains		
DH5 α	F ⁻ ϕ 80d <i>lacZ</i> Δ M15 $\Delta(lacZYA-argF)$ U169 <i>deoR recA1 endA1 hsdR17</i> (r _k -m _k ⁺) <i>supE44</i> λ ¹ <i>thi-1 gyrA96 relA1</i>	Invitrogen
JM109	F ⁺ <i>traD36 proA' B' $\Delta(lacZ)$M15/$\Delta(lac-proAB)$ glnV thi</i>	New England BioLabs
BL21	F ⁻ <i>ompT hsdS_B (r_B⁻ m_B⁻) gal dcm</i> (DE3) pLysS (Cm ^r)	Novagen
Plasmids		
pGEM-7:sp ^R	Spectinomycin-resistance (Sp ^r) <i>aad9</i> determinant with terminator in pGEM-7Zf, (Ap ^r Sp ^r)	(8)
pET46	PCR product cloning vector incorporating x6 His NH ₂ tag, (Ap ^r)	Novagen
pBluescript II SK(+)	Cloning vector, (Ap ^r)	Invitrogen
pBS:sp ^{Rt}	Sp ^r <i>aad9</i> determinant with no terminator (sp ^{Rt}) in pBluescriptII SK(+), (Ap ^r Sp ^r)	This study
pBS:sp ^{Rt} Δ 2006	Sp ^r determinant in pBS:sp ^{Rt} flanked by SGO_2006 and SGO_2005 regions, (Ap ^r Sp ^r)	This study
pGEM-7:sp ^R Δ 2005	Sp ^r determinant in pGEM-7:sp ^R flanked by SGO_2005 DNA amplicons, (Ap ^r Sp ^r)	This study
pGEM-7:sp ^R Δ 2004	sp ^r determinant in pGEM-7:sp ^R flanked by SGO_2004 DNA amplicons, (Ap ^r Sp ^r)	This study

Table S2. Primers used in this study

Primer	Primer location in genome	Sequence ^a	Annealing temperature ^b
<i>Sst</i> 2006usF	2069663-2069686	(TAGAGC) <u>TC</u> GATATTTGTACAGTGCTTGCTC	66
<i>Bam</i> 2006usR	2069366-2069387	(TAGGATCC <u>T</u>)AGCTAGCCCACTCTACTACCAG	66
<i>Eco</i> 2006dsF	2069078-2069098	(ATGAA) <u>TTC</u> CACGATGAAGCAGCGCTTG	62
<i>Xho</i> 2006dsR	2068668-2068690	(TACTCGA) <u>GTT</u> CTGGACTAACATATGTTGTC	62
2006tfJ	2069902-2069925	GTCTTCGACTGGCTTGAATATAGG	66
<i>Xho</i> 2005usF	2068881-2068903	(ATCTCG) <u>AG</u> TTGTTTCGAGCTGATGATCCTC	66
<i>Eco</i> 2005usR	2068511-2068535	(AT)GAATTCGCATCAGTATATACATTCG	66
<i>Hind</i> 2005dsF	2066528-2066550	(ATAAGCTT) <u>CT</u> GATCTTCAAGGCTAAGATTCG	64
<i>Sst</i> 2005dsR	2065799-2065821	(ATGAGCT) <u>CT</u> GATAACCGGTTTGTCTGTACC	64
2005tfJ	2069089-2069112	AGTAATGACAGACTTTTCACGATGA	64
<i>Xho</i> 2004usF	2057867-2057888	(TACTC) <u>GAG</u> CTGACACATGTCCCTACTG	62
<i>Mun</i> 2004usR	2057542-2057563	(ATCAAT) <u>TG</u> AGTCGTAATCTCTGGTTGAG	62
<i>Hind</i> 2004dsF	2056062-2056086	(ATAAGCT) <u>TCA</u> AGTAATTTACCATGGCACTATC	64
<i>Bam</i> 2004dsR	2055682-2055704	(ATGGATC) <u>CT</u> AGAGTATTGAGGTTGTTGTGG	64
2004tfJ	2058006-2058028	CTATGCTCTTCTGGTTACTATCG	64
HP1F	2068889-2068867	(GACGACGACAAGATGG)ATGATCCTCTGAATATTGAGACG	55
HP2R	2067913-2067938	(GAGGAGAAGCCCGGTTT)ATTGCTCATTAACATAGAAGGAAACAT	55
HP3R	2066920-2066943	(GAGGAGAAGCCCGGTTT)AGTCTTTTGTCTTGGCATGGGCATC	55
HP4R	2065006-2065029	(GAGGAGAAGCCCGGTTT)ATTCTTCCAAGGTATCAGAAAGTAC	55

^a Primers were designed based on the annotated genome sequence of *S. gordonii* CH1 (GenBank accession number CP000725). Some primers were engineered with additional sequence (in parentheses) for cloning e.g. to generate restriction sites (underlined), or to encode an in-frame translational stop codon (italicized).

^b *S. gordonii* CH1 chromosomal DNA template was amplified by PCR with an initial denaturation of 95 °C, 30 cycles of denaturation at 95 °C, annealing at the noted temperatures and extension at 72 °C, followed by a final extension at 72 °C for 10 minutes.

Table S3. Characteristics of ORFs interrupted in *S. gordonii* DL1 by allelic replacement

<i>S. gordonii</i> CH1 SGO number ^a	ORF size ^b		Leader peptide (aa) ^{b,c}	Putative functional sub-domain ^d		Region of ORF replaced by <i>aad9</i> ^{b,e,f}		Predicted MW of encoded protein ^g	
	bp	aa		Type	Location in ORF (aa) ^b	bp	aa	Parent	Mutant
SGO_2006	564	187	27-28	thioredoxin	56-172	253-519	85-173	21924	9531
SGO_2005	10941	3646	33-34	vWF type A	72-229	479-2438	161-813	397305	9618
SGO_2004	2424	807	21-22	G5	140-698	431-1885	145-630	86908	18391

^a ORFs were located in the annotated genome sequence of *S. gordonii* CH1 (GenBank accession number CP000725).

^b bp and aa indicate base pairs and amino acid residues, respectively.

^c Predicted signal peptide cleavage site for Gram-positive bacteria, determined using Signal P (3)

^d The presence of protein families of thioredoxin (PF00085) and vWF (von Willebrand Factor) type A domain (VWA; PF00092) were identified using BlastP (1). The presence of a thioredoxin domain in SGO_2006 and vWA in SGO_2005 suggests that these ORFs may be involved in cytochrome maturation (10) and platelet-binding to subendothelial connective tissue (reviewed in ref. 12). The protein family of G5 (PF07501) was determined using Pfam (5). Seven 78-amino acid G5 direct repeats were found in SGO_2004; the presence of such repeats is implicated in binding to *N*-acetylglucosamine (2).

^e The *aad9* gene has its own putative promoter region. In allelic replacement of SGO_2005 and SGO_2004, the *aad9* gene had a terminator region; in SGO_2006, the *aad9* gene did not have this terminator.

^f The reverse primer used to generate the upstream *S. gordonii* DL1 amplicon in SGO_2006 was engineered with a translational stop codon (Table S2), thereby truncating the encoded protein upstream of the *aad9* gene.

^g Predicted molecular mass (Daltons) of the encoded protein (including the predicted signal peptide) in the *S. gordonii* CH1 parent and respective mutant, in which part of the putative functional sub-domain was replaced by the *aad9* gene

Table S4. Quantitative RT-PCR expression of SGO_2005 and SGO_2004

$\Delta\Delta C_t$ Analysis of SGO_2005						
Experiment	Strain	SGO_2005 C_t	<i>gyrA</i> C_t	ΔC_t	$\Delta\Delta C_t$	Fold Change ($2^{-\Delta\Delta C_t}$) ^a
1	DL1	25.06	24.43	0.64	0	1
	UB1890	26.74	25.59	1.15	1.52	0.70
	UB1964	25.83	25.27	0.55	-0.08	1.06
2	DL1	25.48	25.72	-0.24	0	1
	UB1968	23.14	23.32	-0.18	0.06	0.96

$\Delta\Delta C_t$ Analysis of SGO_2004						
Experiment	Strain	SGO_2004 C_t	<i>gyrA</i> C_t	ΔC_t	$\Delta\Delta C_t$	Fold Change ($2^{-\Delta\Delta C_t}$) ^a
1	DL1	24.51	24.43	0.08	0	1
	UB1890	26.55	25.59	0.96	0.88	0.54
	UB1964	26.03	25.27	0.75	0.67	0.63
2	DL1	24.93	25.72	-0.79	0	1
	UB1968	22.62	23.32	-0.70	0.09	0.94

^a Relative expression normalized to *gyrA* of the mutant strains with disrupted SGO_2004 (strain UB1964), SGO_2005 (strain UB1890) or SGO_2006 (strain UB1968) as compared to strain DL1 in same experiment.

Primer	Sequence 5'-3'	T_m (°C)
2005 Upstream	AATCTTCGCAATGGCTTCTG	60.36
2005 Downstream	GGCAACCCTATCAAAGATGC	59.54
2004 Upstream	AAGATCAAGCGAAACCAGGA	59.81
2004 Downstream	CAAAGCCATTTGCATGTTCA	60.64
<i>gyrA</i> Upstream	GAACGTGCAACGGAGCTTAT	57.86
<i>gyrA</i> Downstream	ACCTCCACGTTTTTGAGCTG	58.77

SUPPLEMENTAL MATERIALS AND METHODS

RNA was prepared from mid-log *S. gordonii* cells grown in Todd Hewitt broth, as previously described (13). Briefly, cultures were extracted with hot acid-phenol and precipitated RNA was suspended in DEPC water and purified via an RNeasy Midi spin column (QIAGEN). After DNase treatment to remove any remaining DNA, the RNA was purified with a second RNeasy midi spin column, eluted in RNase-free H₂O with 0.4 Units RNaseOUT™ (Invitrogen) /μl to prevent degradation, and stored in portions at -80°C.

Primer3 software (<http://frodo.wi.mit.edu/primer3/input.htm>) (11) was used to design primers for real-time PCR. Input parameters for primer pairs were a 150-bp target amplicon size, 20-bp primer sequence length, a G-C content of 40-60% for the primer sequences, and an annealing temperature range of 59 - 61°C. All primers were checked against the *S. gordonii* genome sequence to confirm specificity.

RNA from strains DL1, UB1890, UB1964 and UB1968 were templates in real-time PCR to determine the expression of SGO_2005, SGO_2004 and *gyrA*. The iScript One-Step RT-PCR Kit with SYBR Green (BioRad) was used according to the manufacturer's directions for reverse transcription and labeling of cDNA. Real-time data were collected using the MyiQ™ Single-Color Real-Time PCR Detection System (Bio-Rad). Data were analyzed using the Livak ($2^{-\Delta\Delta C_t}$) Method (9) and expression of SGO_2005 or SGO_2004 were reported as a fold-changes normalized to *gyrA* expression in each strain.

Legends to Supplemental Figures

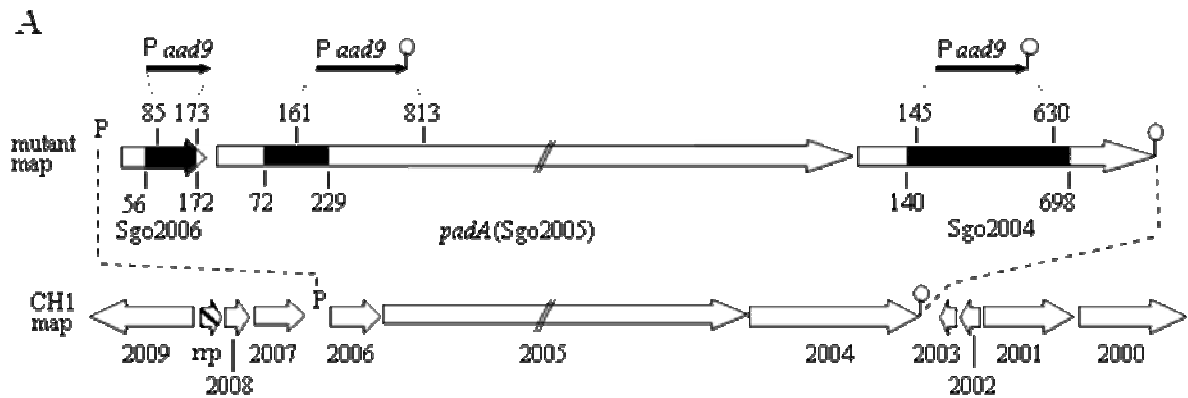
Fig. S1. Panel A: diagrammatic representation of a 19,060-bp *S. gordonii* Challis CH1 (Genbank CP000725) chromosomal region. The arrows in the CH1 map indicate the direction of transcription and relative size and position of each ORF. A 4,500-bp region within SGO_2005 (at 1000-2500 amino acids) is indicated by hatch marks. ORFs designated in the CH1 annotated sequence are labeled with SGO numbers. A predicted promoter (P) sequence was found upstream of SGO_2006 using BPROM software available from Softberry (Mount Kisco, NY; www.softberry.com) and a putative terminator (stemloop) sequence was found downstream of SGO_2004 using mfold (14). The hatched arrow labeled “rrp” is not designated in the current version of the *S. gordonii* CH1 annotated genome. However, this ORF is present and a BlastP search (1), determined that it encodes a 50S ribosomal protein L33 with 84 % amino acid identity to spr1822 (NP_359414) in *S. pneumoniae* R6 (Genbank AE007317). The expanded mutant map above shows details of the location of the putative functional subdomains (in black) of SGO_2006 (thioredoxin domain; PF00085), SGO_2005 (von Willebrand Factor type A domain; PF00092) and SGO_2004 (seven G5 domain direct repeats of 78 amino acids each; PF07501). The amino acid position of these domains is indicated by the numbers below each arrow. The amino acid numbers above each arrow designate the position of the cloned fragment containing the *aad9* gene (encoding spectinomycin resistance, Sp^R), amplified from pGEM7:*aad9* with its native promoter and with (SGO_2005 and SGO_2004), or without (SGO_2006), a transcriptional terminator. Linear fragments from plasmids containing the appropriate homologous streptococcal DNA flanking the directionally-cloned *aad9*-containing amplicon were used to construct three individual gene mutants by allelic replacements in SGO_2006, SGO_2005 and SGO2004. Details of construction of the mutant strains are provided in the **Materials and Methods** section. Panel B: predicted product of each ORF shown in the *S. gordonii* CH1 chromosomal map in panel A.

Fig. S2. Features of PadA (3646 aa residues) primary sequence include: leader peptide (1-33 aa); N-terminal region (blue print, equivalent to Fragment II in Fig. 1) comprising vWF-like domain (72-229 aa, underlined) and two cysteine residues (C55 and C508, purple shaded); fourteen aa residue repeat blocks (1329-3485 aa, green, yellow and blue shading); cell wall anchorage region (3541-3618 aa); sortase motif LPKGTG (3614-3618 aa, purple shaded).. For repeat blocks numbers 1 (green shaded), 13 (green shaded) and 14 (shaded light blue) the sequences tend to diverge. Repeat blocks 1-12 each contain 148-152 aa residues, while repeat blocks 13 and 14 contain 176 aa residues and 179 aa residues respectively. More specifically there is duplication of two super repeat blocks each of 755 aa residues consisting of 5 aa residue repeat blocks: repeat blocks 2-6, 1479-2230 aa residues (designated RP1, 755 aa) (red typeface underlined in red) and repeat blocks 7-11, 2231-2982 aa residues (designated RP2, 755 aa) (black typeface underlined in black). The von Willebrand factor type A (vWA) domain is the prototype for a domain of widespread occurrence involved in a variety of cellular functions (4). It possesses no significant sequence similarity to any known protein structure. Secondary structure predictions suggest a largely alternating pattern of six alpha-helices and six beta-strands. A protein fold for this domain is proposed to correspond to a doubly-wound open twisted beta-sheet structure flanked by alpha-helices (α/β para-rossman type of fold). In vWf the domain forms multimers.

Fig. S3. Analysis of recombinant PadA polypeptides by SDS-PAGE stained with Coomassie Blue. Fragment I (346 aa residues) with predicted M_r 36.2 kDa showed retarded migration in SDS-PAGE with apparent M_r approximately 53 kDa. Fragment II (657 aa residues) with predicted M_r 73 kDa migrated at apparent M_r 80 kDa. Fragment III (1295 aa residues) showed predicted and apparent M_r 142 kDa. M_r , molecular mass markers in kilodaltons.

Fig. S4 Glycoprotein (GP) IIb/IIIa transfected CHO cells or mock infected cells (vector only), were incubated with 10 µg/ml phycoerythrin (PE) labelled primary antibody (mouse anti-CD41, clone SZ22) or isotype control antibody for 20 min at room temperature. The samples were then diluted in 1 ml PBS and analyzed on a FACS-CALibur flow cytometer (San Jose, CA, USA) using CellQuest Pro software. Panel A, separation of mock or GPIIb/IIIa transfected cells; panel B, CD41 gated binding levels for mock or GPIIb/IIIa transfected CHO cells; panel C, Western immunoblot of lysates from mock or transfected cells reacted with anti-CD41 antibody and peroxidase-conjugated anti-mouse secondary antibody, showing GPIIb/IIIa expression.

Fig. S5. Alignment of amino acid sequences of vWf (GenBank accession number P04275) and PadA. There is no significant amino acid sequence identity. Shaded yellow is the A1 domain of vWf showing only weak homology to PadA. The motif in vWf interacting with platelet GPIIb is highlighted in green. Shaded red is the C1 domain of vWf again showing only weak homology with PadA. The vWf motif interacting with platelet GPIIb/IIIa is highlighted in green.



B

- SGO_2009: zinc proteinase
- rrp: 50S ribosomal protein L33 homolog
- SGO_2008: preprotein translocase SecE component-related protein
- SGO_2007: transcription termination/antitermination factor NusG
- SGO_2006: thioredoxin signature protein
- SGO_2005: LPXTG cell wall surface protein (PadA)
- SGO_2004: LPXTG cell wall surface protein
- SGO_2003: structural tRNA (Cys tRNA)
- SGO_2002: hypothetical protein
- SGO_2001: *rpsB* ribosomal protein S2
- SGO_2000: translation elongation factor Ts

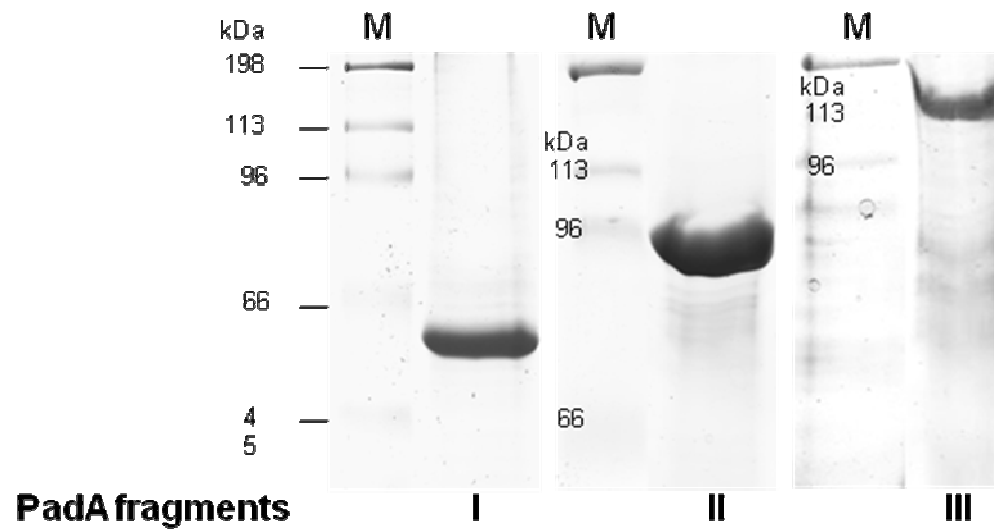
Supplemental material

Figure S1. Structure of the *padA* locus and sites of insertion of *aad9* to generate mutants.

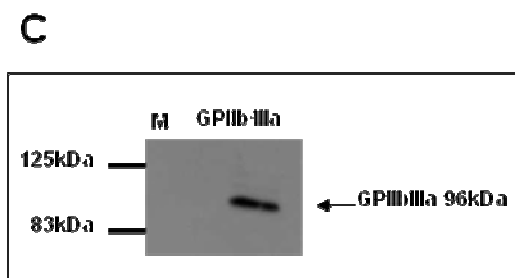
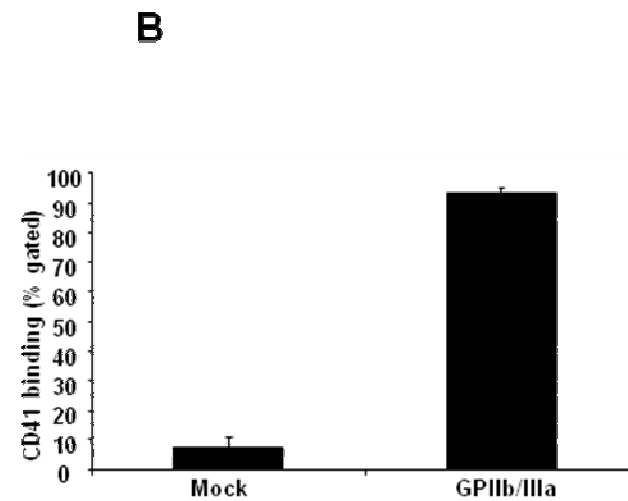
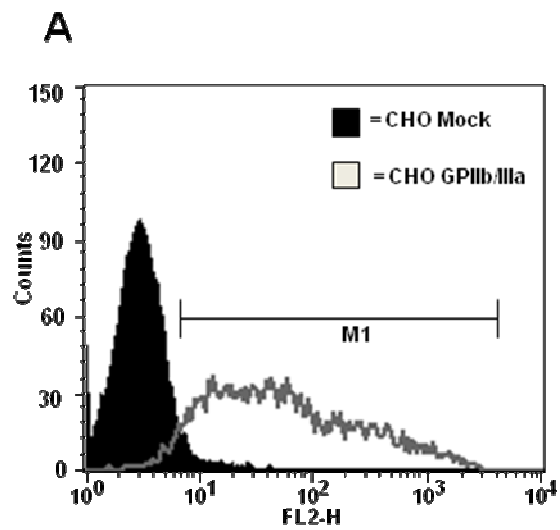
Leader peptide

1 MKDFLKKVLILFTVLLMSMPSSVNLNLTGSVVRADDDPLNIETRRIDEHTTITQNGCYRKIE
61 KTDATDWTVPRKPIDLVLIQDASGSFRITIPSVKNALKRLTTYVSPEQYDENDPHLVKTD vWF-like domain
121 DPTTDRVFVASYQGLDQVRYFENNDFSGNPANVYTDANSTGKNYTYGNSGLTSDQNKVH
181 NFIDNIAVDGGTPTVPAIDDTIAQYNRVKGNMENGKRTVFLVTDGVDVANGYRLPGTNTVV
241 MDKSWTRTDAIQAWRVDSYPEAAQDIIGRANELKAAGNQLKAAVGVSEGSVVVGFWERVD
301 NFKTEKYYQYGPAYLNGFGNTINIGDNRVQAI FHDALQSMASPDKVVNGKNVSYFYVNEQN
361 NIDVFSQKILESVAALVKDDITGEFDITEGYKVD AIRINGKKIVPKVTDPSKEIRGTIT
421 QTGNKVKISVPDSVFNPGKNSFDYDLSKEARAPETDEDSEVDPPENYVPEKEEITVPELT
481 GKFKAGDFETRQIGGRNQTVVEVQKLEYCYPSATKTVKDADASNDIGVIPDPLELTKKPSY
541 SAQLSKKDEEFTYTVDYNFNVPYEFKENVMLTDPIDYRLEVVSASHAQGPDGQSWPTRV
601 TQQDAGGNSQSVVADVPPQKDYNYLIMKKAKLKMTVRLKEEYRKNQASKAYLAILQNN
661 NGYGLVNGQINIMWNGEDDSPNQDAHAKTKDKASTIRRSNPIYVKPPLDTEVDKKNVNEKEH
721 EGLQADGEEFEYKVTAPWPGIADKFTLTDTVVDELEIVPNSAKVTIVAGKSYNALTKAISI
781 NGQTIDITLTKAQLTSLNRLISRRGGSEVQEI EELIFKAKIRPGADLSKYKKNGAVNIPNT
841 ADVILNDKKKTSKEVTVTPPKPKPTVSKKINNTLDSLVTFDGQPYTYNITTAVPSDVAG
901 YKFFVISDKLDLADDFDQASISGLPADVFEIQTNQGTVTATVKEGKFKELAKYSFVELT
961 IPAKVKAGVTGKTIENKAKISFTNENNAKEVESNPVTVTPPPVTKKINENLDHLDIATG
1021 QPYKYNVKTTLPSDITSYKEFVIDTLEDELSVINEGTDKPVISGPAEFFDVTVSGQKV
1081 TATMKNFAGASALAGQEI EELVIPA KINDGVTRSNIPNKATFSFKDKNDHKGEKETIPVTV
1141 TPTTEPNVSKKINGDQDNATIAAETDFTYNIKTTLPNIDITYKSAITDTLDENLGVVNP
1201 EPSISEEAKKFFDITVSGNTVATMKDFAKASALANKEI EELVIHAKVKKESVLP EIPNTA
1261 KITTYTNKNNESKEKETEPVKVTPPPITKKVNGKDQEDLASLTSFKYTVDSKVP I VADKF
1321 VLSDTLEKVLTFDGDATVTDIGQTVTDVTVAKKDQKLTVTDFDKDQVKKYAGKAVQVAFDA
1381 KIKSGYTVDQVLVAKYPNGDKAAIPNKASFVNDNPETEKF SNPVTVTPPPPNTPEIEKKV
1441 NGADSYNLQTRLEEFYSLNTAMPTNATEFTVDELKSVLEFAGKKGDVQVKIDGKAAND RP1 755 aa
1501 QATISTDKNLTVAFAEKAVKANAGKSI EIVTFKAKIREGANLLDYLVPGGQIRIPNKASY
1561 DIDHNPKFHKDSNEVPVTPPSPEQPPIEKDVNDKAEATLEARDEEFTYHVTKIPYEATA
1621 FNITDTLKEVLEFSGEKGQATVVDGKLLSDPHIAINGQITVTLNQEELKANADKEIKL
1681 TPKAKIRPNANLAAYVVGDKVIVNNQASYNVLDLFDNPGVHKDSNIVPVTPPSPEKPEIEY
1741 TVNDAKEATLANRDEIFTYKVKTRVFFDATAFSDDTIKVLEFADAGSATLNGEAL EAD
1801 RISIADQKITLTLTEDQVKNNGGKEVVLTFKAKIROGANLSGYIEKGKTVINNQASYNAA
1861 FPNDFPNFHKDSNIVPVTPPNPNPPIEKKVNEAESANL GARDEEFTYITDITVPLDVTGF
1921 AVYDTIEKVLRESGENGQASAVDQQLDASHITIKGQKITVKNTEDEAKALGKAVHVS
1981 FRAKIRAGANLSDYIERDGTIRIYNTAKYFNNDPGTEQSSKPVVPIPTTPEPELKKEV
2041 NGKEABTDANRDEEFTYITVKTVPVQDATAFSDGSLVLEFAGEDAEASLTLNGEKLDA
2101 KQIKLKDQTI SAELTEAQVKANGGKEVVLNFKAKIREGANLADYIEADGVTRVPNKASYV
2161 ANFPHRPKVEKDSNIVPVTPPSPENPPVEKKVNNKPSATLDSRDEEFTYITDKVPVDAT
2221 GFKITDELKDLEFSGKAGQAVTVVDGADVLEFSQITVQAVTVVTLTADQVKNYGNKA
2281 VNVSEKAIKKNVSLAGYIANGVTRIPNIAKYLINDDFIKKSTEEVYVPPSPEEGL
2341 KPEVNGQPTALPEYERFTYKVTTSVVDATAFVSDDTAVVLEFSGEKGQATATLDGQ
2401 EIDANRINVAQDITSMALTEDEVKANGGKEVTLTFKAKIREGANLSAYIEKGKTSIPNTA
2461 SYTAGFPNRPDIHKDSNRVPTPPTPEEPEIKKDVNGKEETLANRND EFTYHINTKVPF
2521 DATAFSINDELKDLEFADGTGRATASLNGALDADRISINGQITVNLTEDEVKNNGC
2581 DQVTFANFQVNSCHINQKTSINQASVVDTPNNDQVKNIEVFTPTPEE
2641 FIEKKVNEAESANL GARDEEFTYITDITVPLDVTGFAVYDTIEKVLRESGENGQASATVD
2701 GQPLDASHITIKGQKITVKLTEDEAKALGGKAVHVSFKAKIKAGANLSDYIEKDGTTRIY
2761 NTAKYFNNDPGTEQSSKPVVPIPTTPEPELKKEVNGKEAETLANRDDVFTYTVKTTVP
2821 QDATAFSISDKLEDLEFAGKAGATLAGEDKADQITTDGQIKLTLTEDQVANGGKEV
2881 VLNFKAKIREGANLSAYIEKADKAEPNTASYVNGFNKPAYKDSNEVPVTPPSPEEGL
2941 ERDYNKPSSETIADNTEFTYNIHTTNPQDAIGFTVYDELKIVLEFAGDVQVTLGGKKAD
3001 AAVAKNGQTL E VTFPEETVKANGGKQVTFKAKIKADADLTPYETANSYSVPNTASYLI
3061 NNNPTSKKETKPVTVVEVPKQPGPEVTKKINRTLDHLDVDRDVPYMYNVNTQIPKDIRLYK
3121 EFTVTDITLEFVLEITGTPVAVVDGYATDAVETKVEGNTVTVTKDFARISGYKEIQLYIF
3181 AKLKADSDLSAYENQTVPNKATI AFKDSNGKNGTKESNPVTVRPRDPEKPEEPKPNPAK
3241 TVGPDGNSNPSTAYRLKELKEGFRFDVTAKVPTDPVDESGNPIKDAQGRDVKTELNSFTV
3301 TDELEKVLKVDRAVAVKVEENKVAEAI AKITAKIEKAESDLKELEGKETNGTFAKKLAEAE
3361 KKVEELTAQLAAAKEKAAAAPATPAPASDS DAGNATATPAPADNNAEVAALEESLAAAQA
3421 ELEQLKADGAKAGNLATPEEQKVEQDKLNKNLEQLKESKEKLEKALEAFTTVNDKGEITD
3481 EALAKIAKVTVEGQKVTVEVTDKAVLEALKGSTFRVIIYSSIKDGADLSSYLNKENNETK
3541 IPNKATVTFNDKPKVTNTVNVYPPPEPTTPPQTPPHPTPTTPGTPPPTTPDTPPAPKGDLP
3601 PAPTPEPEKPKNI LPKTGTSATMVNEVIIGMILVLMGLLLRRRPPKH CWA

Supplemental material
Figure S2. Primary sequence of PadA highlighting regions of interest and potential functional significance



Supplemental material
Figure S3.
SDS-PAGE gels of purified PadA protein fragments I, II and III.



Supplemental material
Figure S4.
CHO cells expressing GPIIb/IIIa

Petersen et al., 2009. Platelet Receptor GPIIb/IIIa Interaction with PadA

vWf YVLVQDYCGS-----NPGTFRILVGNKGCSPVSKCKKR-----V 925
 PadA TATMKNFAGASALAGQEIELVIPAKINDGVTRSNIPNKATFSFKDKNDHKGEKETIPVTV 1140
 . : : : : * : * * . * : * * . * : : * *

vWf TILVEGGEIELFDG-----EVNVKRPMDETHFEVVESGRYIILLGKALSVVWDR 976
 PadA TPTTEPNVSKKINGDQDNATIAAETDFTYNIKTTLPNIDITYKSFATDTLDENLGVVNP 1200
 * . * . : : * * . * . : : * : : * . * :

vWf HLSISVVLKQTYQEKVCG-----LCGNFDGIQNNDLTSSNLQVEEDPVDFGNSW 1025
 PadA EPSISSEAKFFDITVSGNVTATMKDFAKASALANKEIELVIHAKVKESVLPPEIPNTA 1260
 . * * * * : : : * * * . . : : : : : : * . : : * :

vWf KVSSQCADTRKVPDLSSPATCHNNIMKQTMVDSSCR-----ILTSDFVQDCNKL 1074
 PadA KITYTNNKNESKEKETEPVKVTPPPITKKVNGKDQEDLASLTSTFKYTVDSKVPVADKF 1320
 * : : . . . : : * . . : : : : . . . : * * * : : :

vWf VDPEPYLDVCIYDTCSCESIG----- 1095
 PadA VLSDTLEEVLTDFGDATVTIDGQVTDVTVAKKDQKLTVTFDKDQVKKYAGKAVQVAFDA 1380
 * . . . : * : * : : * .

vWf -----DCACFCDTIAAYAHVCAQHGVVTVRWTATLCP----- 1127
 PadA KIKSGYTVDLVAKYPNGDKAAIPNKASFVVDNDNPETEKFVSNPVTVPVPPNTPEIEKKV 1440
 * * : : : : : : * . . * * *

vWf QSCEERNLRENGYECWEVRYNSCAPACQVTCQHPEPLACPVCVEG-----CHAHCP 1178
 PadA NGADSYNLQTRLEEFYSLNTAMPNATEFTVTDLKSVELEFAGKKGDVQVKIDGKAAND 1500
 : . . . * * : . * : * : * : . : * . : : . : *

vWf PGKILDELLQTCVDPEDCPVCEVAGRRFASGKKVTLNPSDPEHCQICHCDVVNLTCEACQ 1238
 PadA QATISTDKNTLTVAFAEKAVKANAGKSIEVTFKAKIREGANLLDYLVPGGQIRIPNKASY 1560
 . . * : * : * * * : * . . . : : : : : * .

vWf EPGG-----LVVPPTDAPVSPTTLYVEDISEPPLHD----- 1269
 PadA DI **DHNP**KFHKDSNEVPVTPPSPEQPPIEKDVNDKAEATLEARDEEFYHVTKIPYEATA 1620
 : . : * * . . . * * * * * * .

vWf **FYCSRLDLVFLLDGSSRLSEAEFEVLKAFVVDMMERLRISQKWRVAVVEYHGDGHAYI** 1329
 PadA **FNITDITLKEVLDVDFSGEKGQAEATVDGKKLSDDHIAINGQTIITVTLNQEELKANADKEIKL** 1680
 * : * . * : : * . . : * * : : * . : : : : : : : : : : : : : : : : : :

vWf **GLKDR**KRPSE--LRRIASQVKYAGSQVASTSEVLKYLTFQIFSKID----RPEASRIAL 1382
 PadA **TFKAKIRPNANLAAYVVGDKVINNOQASYNVDLPDNPVGHKDSNIVPVTPPSPEKPEIEK** 1740
 : * : * * . : : : . . * : : . : : . : : * : * * * * . . *

vWf **LLMASQEPQMRSRNFVRYVQGLKKKKVIVIPVGIGPHANLKQIRLIEKQAPENKAFVLS** 1442
 PadA **TVNDAKEATLANRDEIFTYKVKTKVPFDATAFSIDDTIKDVFLEFADAGSATLNGEALEAD** 1800
 : : : * . . * : : : : * * . : : * . * : : :

vWf **VDELEQQORDEIVSYLCDLAPAEPPPTLPPHMAQVTVGP**GLLVSTLG-----PKRNSM 1495
 PadA **RISIAQDKITLTLTEDQVKNNGGKEVVLTFKAKIRQGAN**LSGYIEKGTVINNOQASYNAA 1860
 . : : * : : . : : * : : * . * * * . . * :

vWf VLDVAFVLEGSDKIGEADFNRSKEFMEEVIQRMDVG-----QDSIHVTVLQY 1542
 PadA FPNDPNFHKDSNIVPVTPPNPENPPIEKKVNEAESANL GARDEEFTYITDITVPLDVTGF 1920
 . : . . : * : : : * : : * : : : . : : : : * :

vWf SYMVTVEYPFSEAQSKGDILQRVREIRYQGGNRTNTGLALRYLSDHSFLVSQGDREQAPN 1602
 PadA AVYDTIEKVLFEFSGENQASATVDGQPLDASHITIKGQKITVKLTEDEAKALGGKAVHVS 1980
 : * : * : . : : * : * : : : * . * : . . : * : : .

vWf LVYMTGNPASDEIKRLPGDIQVVPVIGVGNANVQELERIG-----WPNAPIILQDF 1654
 PadA FKAKIKAGANLSDYIEKDGTTRIYNTAKYFNNDPGTEQSSKFPVVPPTPEPELKKEV 2040
 : : : . . * : : . * * : . * . * * * : : .

vWf ETLPREAPDLVLQRCCSGEGLQIPTLSPAPDCSQPLDVILLDGSFFPASVYFDEMKSFA 1714
 PadA NGKEAETLANRDDVFTYTVKTTVPQDATAFSISDSLVPVLEFAGEDAEASLTLNGEKLDA 2100
 : * : : : * . . * . * * * : * : * . . : : : * *

vWf KAFISKAN-IGPRLTQVSVLQYGSITTDVPWNVVPEKAHLLSLVDVMQREGGSPQIGDA 1773
 PadA KQIKLKDQTI SAELTEAQVKANGGKEVVLNFKAKIREGANLADYIEADGVTRVPNKASYV 2160
 * : * : * . . * * . . * . . : : * * * * . : : . * . . .

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