

## X-ray crystallographic analysis of the 6-aminohexanoate cyclic dimer hydrolase: catalytic mechanism and evolution of an enzyme responsible for nylon-6 byproduct degradation

### SUPPLEMENTARY DATA ON-LINE

#### Legend for supplementary figures

**Fig. S1. Multiple 3D-alignment of 6-aminohexanoate cyclic dimer hydrolase (NylA), glutamyl-tRNA<sup>Gln</sup> amidotransferase subunit A (GAT), malonamidase E2 (MAE2) and peptide amidase (PAM).**  $\alpha$ -Helices and  $\beta$ -strands of NylA shown in Fig. 2A are illustrated at the top with green cylinders ( $\alpha$ -helix) and orange arrows ( $\beta$ -strands). Multiple 3D alignments were carried out using Secondary Structure Matching (SSM) (45), and the secondary structures are shown as *green* (helix) or *orange* ( $\beta$ -strand) letters. The numbering of GAT, MAE2 and PAM is the same as the numbering registered in PDB. The Ser-*cis*-Ser-Lys catalytic triads conserved in AS family enzymes [Ser174-*cis*-Ser150-Lys72 (NylA), (Ser178-*cis*-Ser154-Lys79) (GAT), Ser155-*cis*-Ser131-Lys62 (MAE2), Ser226-*cis*-Ser202-Lys123 (PAM)] are shown as *red* letters marked with *yellow*. Acd-binding residues (Asn125 and Cys316) in NylA are marked with *light blue*.

**Fig. S2. Stereoviews of overall structures of 6-aminohexanoate cyclic dimer hydrolase (NylA).** Superimposition of NylA structure (*green*) with glutamyl-tRNA<sup>Gln</sup> amidotransferase subunit A (PDB ID: 2DF4; *blue*) (A), malonamidase E2 (PDB ID: 1ocm; *purple*) (B), and peptide amidase (PDB ID code: 1m21; *olive*) (C). Superimposition was carried out on the transformation matrix generated by Secondary Structure Matching (SSM) (45).

**Fig. S3. CD spectra at far UV for NylA and various mutants in 20 mM potassium phosphate buffer (pH 7.3) containing 10% glycerol.** CD measurements were carried out in a J-720WI spectropolarimeter (Jasco). Cuvette with a pathlength of 1 mm was used for far UV CD. The results are expressed as the mean residue molar ellipticity,  $[\theta]$ , defined as  $[\theta] = 100 (\theta_{\text{obs}} - \theta_{\text{back}}) / l c$ , where  $\theta_{\text{obs}}$  is the observed ellipticity in degrees,  $\theta_{\text{back}}$  is the observed ellipticity in degrees without enzyme as background,  $c$  is the molar concentration of the residue, and  $l$  is the length of the light path (in centimeters). The temperature was controlled at 25 °C with a Jasco PTC-348WI peltier system. The protein concentration used (in far UV CD measurements) was 0.11 mg ml<sup>-1</sup>. **A.** Wild-type NylA (*orange*), NylA-A<sup>150</sup> (*green*); **B.** Wild-type NylA (*orange*), NylA-A<sup>125</sup> (*blue*), NylA-S<sup>316</sup> (*yellow green*), NylA-D<sup>316</sup> (*pink*).

**Fig. S4. Temperature factors for NylA and NylA-A<sup>174</sup>/Acd complex.** **A.** Average values for  $C_{\alpha}$ ,  $C$ ,  $N$  in each amino acid residue are plotted as a function of residue number. **B.** Data for amino acid positions 300-450 are shown with the positions of  $\alpha$ -helices (H14-H18),  $\beta$ -strands ( $\beta$ 9- $\beta$ 11), and the loop region. Open circle, wild-type NylA; closed circle, NylA-A<sup>174</sup>/Acd complex.

### Supplementary Table

Table S1. Primer DNA used for site-directed mutagenesis of *nylA* gene

Primer	Mutation	Sequence
<b>A</b>		
FE1K72A	K72A	5'-TGCCCTATCTTCTG <u>GCG</u> GACCTCACC-3'
FE1S150A	S150A	5'-TCGTTGGCGGAG <u>GCG</u> AGCGGCGGCTCA-3'
FE1S174A	S174A	5'-GACGCGGCAGGT <u>GCT</u> GTGCGCATACT-3'
RE1C316E	C316E	5'-GATCGCGACGTC <u>CCT</u> CAATCGTCGAGTAG-3'
RE1C316X	C316D, C316G C316S,	5'-GATCGCGACGTC <u>NNN</u> AATCGTCGAGTAG-3'
<b>B</b>		
FE1N125A RE1N125A	N125A	5'-GAGATGGGCG <u>GCT</u> CAGGTAACGACGGAGCCC-3' 5'-CGTTACCTG <u>AGC</u> GCCCATCTCCGGTGTATT-3'
FE1C316-3 RE1C316-3	C316A	5'-ACGATT <u>NCC</u> GACGTCGCGATCGCGCGA-3' 5'-GACGTC <u>GGN</u> AATCGTCGAGTAGTCCTT-3'
FE1C316-2 RE1C316-2	C316K	5'-ACGATT <u>NAG</u> GACGTCGCGATCGCGCGA-3' 5'-GACGTC <u>CTN</u> AATCGTCGAGTAGTCCTT-3'
FE1C316-1 RE1C316-1	C316N	5'-ACGATT <u>NAC</u> GACGTCGCGATCGCGCGA-3' 5'-GACGTC <u>GTN</u> AATCGTCGAGTAGTCCTT-3')

- A. Site-directed mutagenesis was carried by "modification of restriction-site (MR)" method (23).  
 B. Site-directed mutagenesis was performed using the PrimeSTAR mutagenesis kit (Takara Bio Inc., Otsu, Japan). Mutated sites in the primer sequences are underlined.



Fig. S1

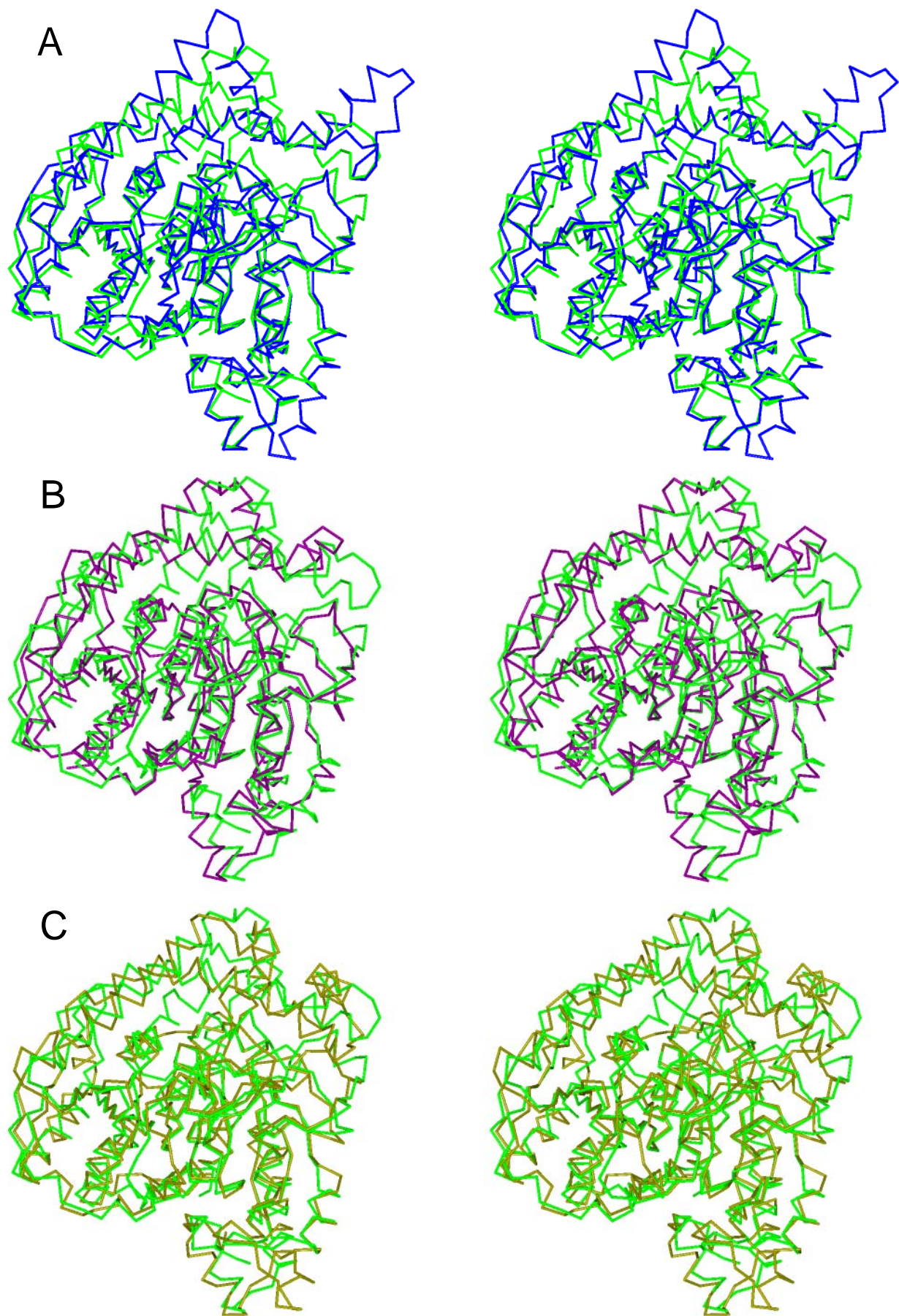


Fig. S2

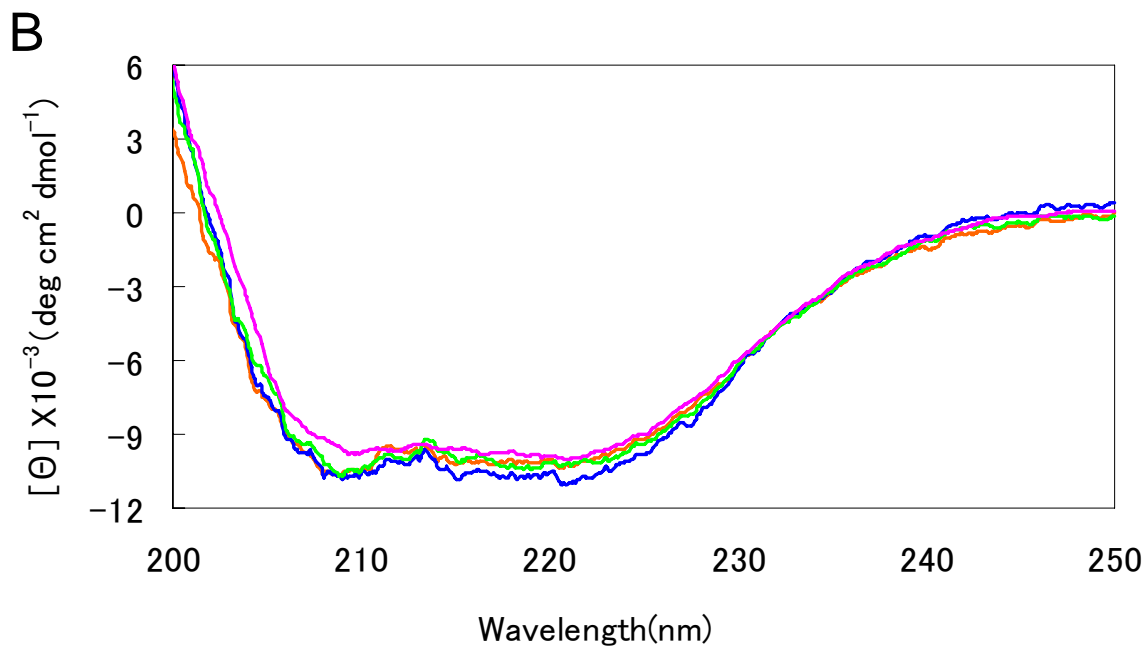
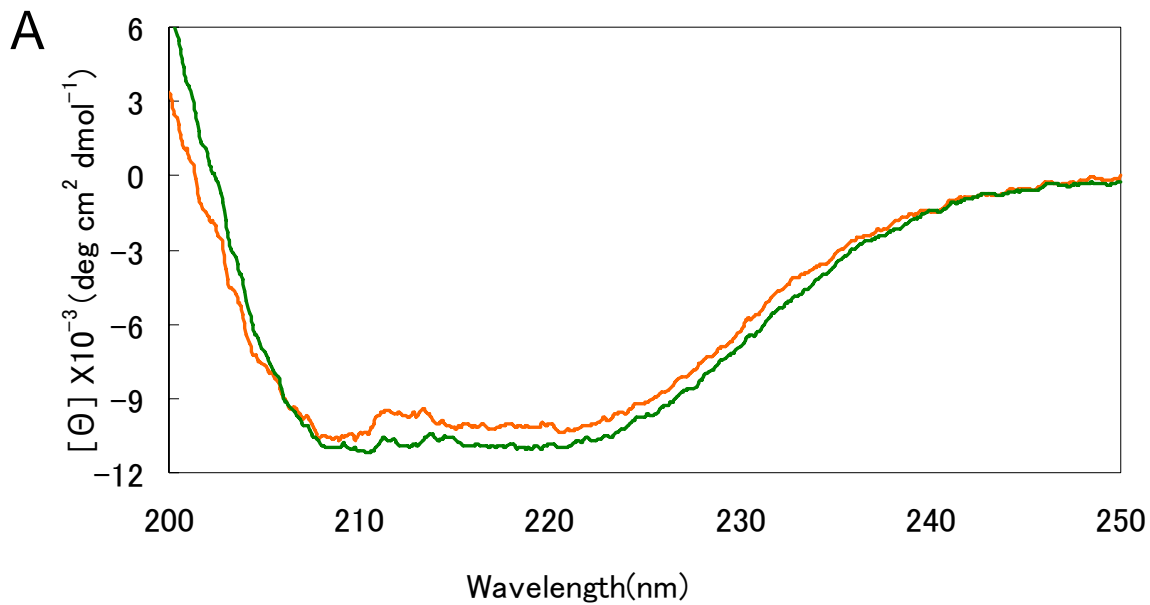


Fig. S3

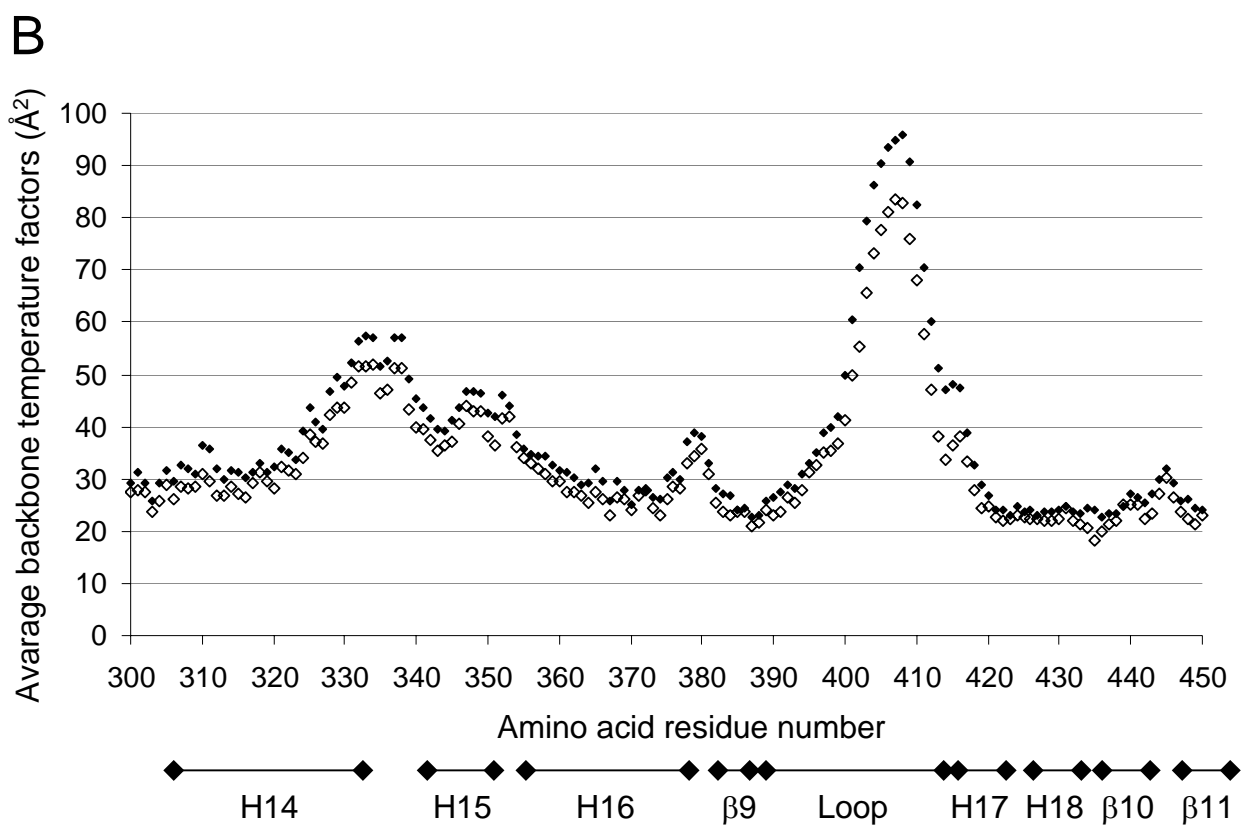
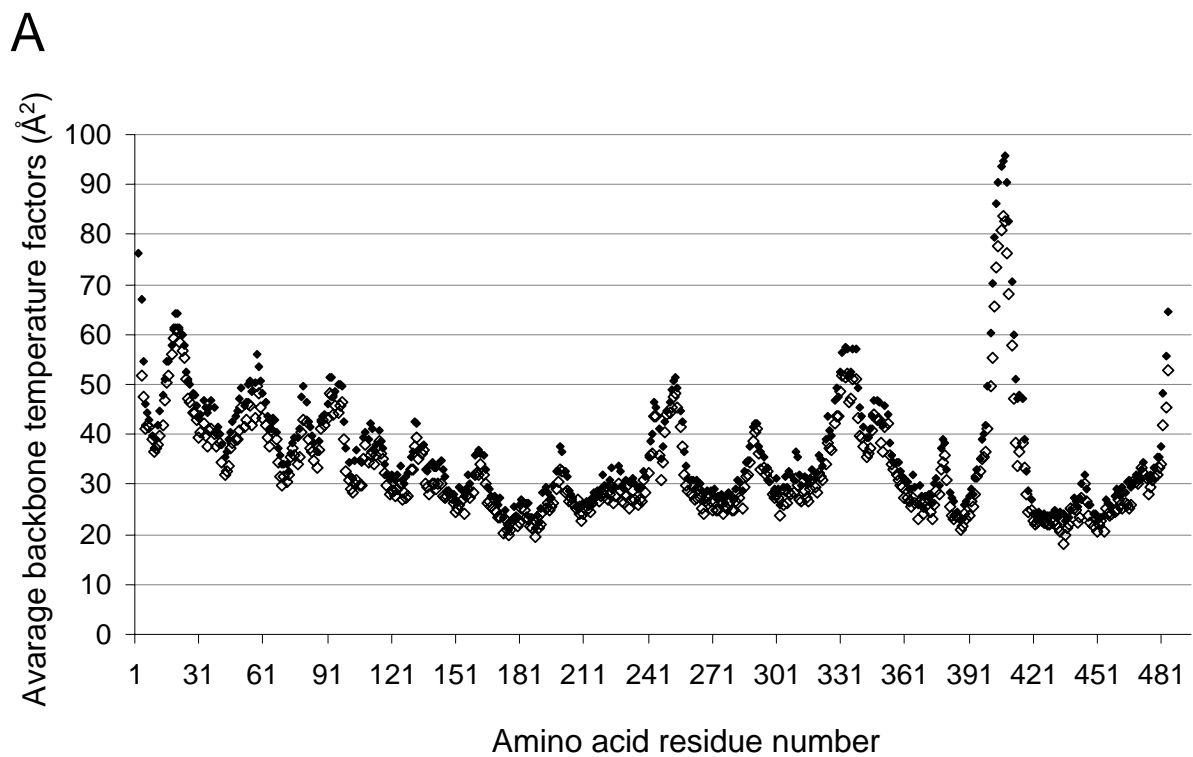


Fig. S4