

Supporting Information
Journal of Organic Chemistry

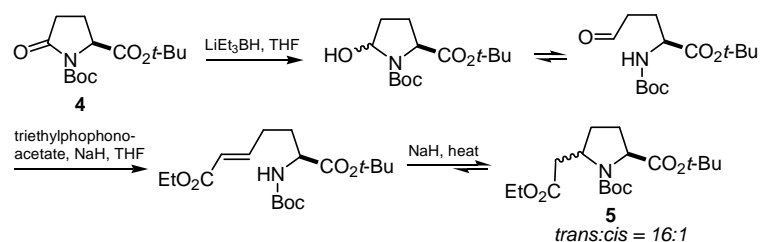
“Synthesis of Carbapoyochelins via Diastereoselective Azidation of 5-(Ethoxycarbonyl)methylproline Derivatives”

Wathsala Liyanage, Laksiri Weerasinghe, Roland K. Strong, and Juan R. Del Valle

CONTENTS

Experimental procedures	S2-S5
¹ H NMR Spectrum of 5	S6
¹³ C NMR Spectrum of 5	S7
¹ H NMR Spectrum of 10	S8
¹³ C NMR Spectrum of 10	S9
¹ H NMR Spectrum of 11	S10
¹³ C NMR Spectrum of 11	S11
¹ H NMR Spectrum of 12	S12
¹³ C NMR Spectrum of 12	S13
¹ H NMR Spectrum of 13	S14
¹³ C NMR Spectrum of 13	S15
¹ H NMR Spectrum of 14	S16
¹³ C NMR Spectrum of 14	S17
¹ H NMR Spectrum of 15	S18
¹³ C NMR Spectrum of 15	S19
¹ H NMR Spectrum of 2 · TFA	S20
¹³ C NMR Spectrum of 2 · TFA	S21
HPLC Trace for 2 · TFA	S22
ESI-MS (RT=10.38 min) 2 · TFA	S23
¹ H NMR Spectrum of 3 · TFA	S24
¹³ C NMR Spectrum of 3 · TFA	S25
HPLC Trace for 3 · TFA	S26
ESI-MS (RT=9.76 min) 3 · TFA	S27
¹ H NMR Spectrum of 16	S28
¹³ C NMR Spectrum of 16	S29
X-ray crystal structure of 16	S30-S31

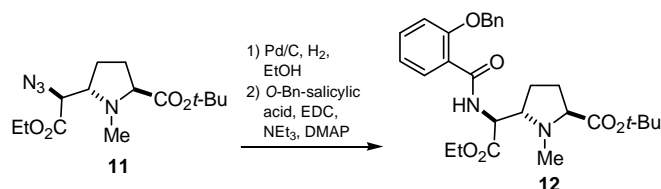
General. Unless stated otherwise, reactions were performed in flame-dried glassware under a positive pressure of argon using dry solvents. Commercial grade reagents and solvents were used without further purification except where noted. Diethyl ether, dichloromethane, and tetrahydrofuran were purified by solvent purification system. Other anhydrous solvents were purchased directly from chemical suppliers. Thin-layer chromatography (TLC) was performed using silica gel 60 F₂₅₄ pre-coated plates (0.25 mm). Flash chromatography was performed using silica gel (40 μm particle size). The purity of all compounds was judged by TLC analysis (single spot/ two solvent systems) using a UV lamp or CAM or ninhydrin or basic KMnO₄ for detection purposes. NMR spectra were recorded on 300 MHz and 400 MHz spectrometers. ¹H and ¹³C NMR chemical shifts are reported as δ using residual solvent as an internal standard. Analytical high performance liquid chromatography (HPLC) was performed C₁₈ reverse phase analytical column. HPLC elution was carried out with a 20 min linear gradient of 0% - 90% MeCN in water (containing 0.1% formic acid buffer).



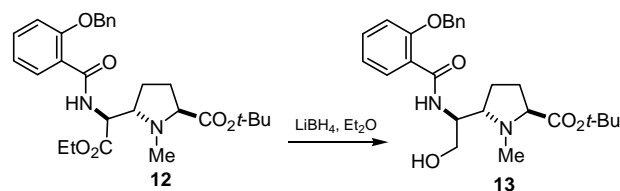
(2S,5S)-5-Ethoxycarbonylmethylpyrrolidine-1,2-dicarboxylic acid di-tert-butyl ester (5). A solution of **4** (3.00 g, 10.51 mmol) in THF (30 mL) was treated dropwise at -78 °C with LiEt₃BH (1M solution in THF, 12.62 mL, 12.65 mmol), stirred at -78 °C for 30 min, and carefully quenched with sat. aq. NaHCO₃ (10 mL). The mixture was warmed to -15 °C and hydrogen peroxide (30% in water, 5.0 mL) was added dropwise. After stirring at 0 °C for 30 min, the solution was decanted from a white solid residue and concentrated. The crude material was taken up in Et₂O, washed with sat. aq. NaHCO₃, dried over MgSO₄, filtered, and evaporated under reduced pressure. The crude *N*-acyloxyaminal was used in the next step without further purification.

A suspension of sodium hydride (60% dispersion in mineral oil, 0.59 g, 14.72 mmol) in THF (30 mL) was treated dropwise with triethylphosphonoacetate (3.16 mL, 15.77 mmol) and stirred for 2 h at RT. The reaction was cooled to -15 °C and a solution of the above *N*-acyloxyaminal in THF (20 mL) was added dropwise. After the addition, the reaction was allowed to warm to RT and stirred 18 h, then treated with an additional 0.21 g of 60% NaH (5.26 mmol) and heated to 50 °C for 4 h, then cooled to RT and poured carefully into sat. aq. NaHCO₃ (failure to heat the reaction with additional NaH resulted in isolation of the ring-opened enoate as the major product). The mixture was extracted with EtOAc, dried over MgSO₄, filtered, and evaporated under reduced pressure to furnish the crude diester. Purification by flash chromatography over silica gel (20% EtOAc/hexanes as eluant) gave **5** as white solid (3.16 g, 84% from **4**). ¹H NMR of crude **10** (*vide infra*) later revealed compound **5** to be a 16:1 trans:cis mixture of diastereomers. ¹H NMR (300MHz, CDCl₃) for the major trans isomer δ 4.41-4.25 (m, 1H), 4.20-4.08 (m, 3H), 2.97 and 2.87 (2dd, J = 15.2, 3.4 Hz, 1H, rotamers), 2.32-2.05 (m, 3H), 1.91 (q, J = 5.9 Hz, 1H), 1.74 (m, 1H), 1.47 and 1.43 (2s, 9H, rotamers), 1.44 and 1.42 (2s, 9H, rotamers), 1.25 (q, J = 7.0 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 171.9, 171.4, 154.0, 153.6,

90.0, 80.1, 79.9, 60.4, 60.3, 54.7, 39.4, 38.4, 28.4, 28.3, 28.0, 27.9, 14.2; ESI-MS (m/z) [MH]⁺ calcd for C₁₈H₃₁NO₆ 358.22, found 358.35.

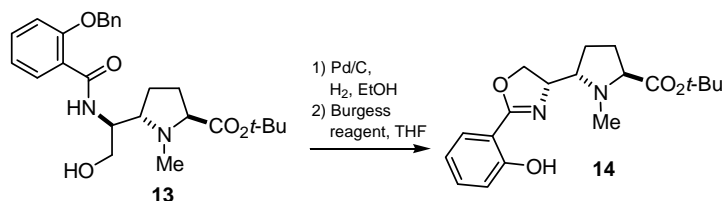


(2S,5S,1'S)-5-[(2-Benzyloxybenzoylamino)-ethoxycarbonylmethyl]-1-methylpyrrolidine-2-carboxylic acid *tert*-butyl ester (12): A solution of azide **11** (250 mg, 0.80 mmol) in EtOH (5 mL) was treated with a catalytic amount of 10% Pd/C and placed under H₂ atmosphere (balloon). After stirring at RT for 18 h, the mixture was purged with a stream of Ar and filtered through celite with EtOAc rinsing. Evaporation of the filtrate gave the intermediate diamine as a white solid (220 mg, 96% yield) which was taken up directly in CH₂Cl₂ (2mL) and treated with 2-benzyloxy-benzoic acid ethyl ester (200 mg, 0.90 mmol), EDC·HCl (180 mg, 0.96 mmol), NEt₃ (130 μL 0.96 mmol), and a catalytic amount of DMAP. The reaction was stirred 12 h at RT, concentrated, and the crude residue was purified by chromatography over silica gel (5-30% EtOAc/hexanes gradient as eluent) to give amide **12** as a sticky white foam (370 mg, 93% from **12**). ¹H NMR (300 MHz, CDCl₃) δ 8.38 (d, J = 6.8 Hz, 1H), 8.23 (dd, J = 7.9, 2.1 Hz, 1H), 7.50-7.35 (m, 6H), 7.05 (m, 2H), 5.21 (m, 2H), 4.86 (dd, J = 6.7, 3.8 Hz, 1H), 4.19 (q, J = 7.0 Hz, 2H), 3.50 (ddd, J = 8.8, 5.0, 3.8 Hz, 1H), 3.37 (dd, J = 7.6, 1.5 Hz, 1H), 2.40 (s, 3H), 1.84-1.35 (m, 4H), 1.43 (s, 9H), 1.26 (t, J = 7.0 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 173.4, 171.2, 165.4, 157.0, 135.6, 132.8, 132.6, 128.9, 128.7, 128.2, 121.6, 121.4, 112.7, 80.8, 71.2, 67.2, 63.0, 61.0, 53.6, 34.8, 28.2, 27.5, 25.0, 14.2; HRMS (ESI-TOF) (m/z) [MH]⁺ calcd for C₂₈H₃₆N₂O₆ 497.2646, found 497.2647.



(2S,5S,1'S)-5-[1-(2-Benzyloxybenzoylamino)-2-hydroxyethyl]-1-methylpyrrolidine-2-carboxylic acid *tert*-butyl ester (13). A solution of **12** (390 mg, 0.78 mmol) in Et₂O (4 mL) was treated with LiBH₄ (2M solution in THF, 580 μL, 1.16 mmol) and stirred for 12 h at RT. The reaction was quenched by addition of 1M NaOH (6 mL), stirred for an additional 10 min, then diluted with water and extracted with EtOAc. The combined organic layers were dried over Na₂SO₄, filtered, and evaporated and the crude residue purified by chromatography over silica gel (60% EtOAc/hexanes eluant) to give alcohol **13** as a sticky white foam (249 mg, 70%). ¹H NMR (300 MHz, CDCl₃) δ 8.58 (d, J = 4.0 Hz, 1H), 8.19 (d, J = 8.0, 2.0 Hz, 1H), 7.41-7.34 (m, 6H), 7.06 (m, 1H), 6.98 (d, J = 8.4 Hz, 1H), 5.4 (bs, 1H), 5.23 (s, 2H), 4.04 (m, 1H), 3.78 (dd, J = 11.6, 5.6 Hz, 1H), 3.63 (dd, 11.6, 2.4 Hz, 1H), 3.42 (dd, J = 7.6, 1.6 Hz, 1H), 3.30 (ddd, J = 8.8, 5.6, 3.2 Hz, 1H), 2.31 (s, 3H), 2.00 (m, 1H), 1.84-1.62 (m, 2H), 1.49 (m, 1H), 1.43 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ 172.9, 167.0, 156.6, 135.7, 132.8, 132.3, 128.9, 128.6, 127.4, 122.0,

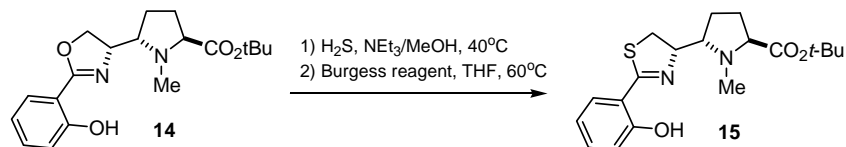
121.8, 121.5, 112.9, 81.1, 71.0, 67.3, 65.3, 63.6, 54.6, 35.4, 28.2, 27.6, 25.3; HRMS (ESI-TOF) (m/z) [MH]⁺ calcd for C₂₆H₃₄N₂O₅ 455.2546, found 455.2550.



(2*S*,5*S*,4'*S*)-5-[2-(2-Hydroxyphenyl)-4,5-dihydrooxazol-4-yl]-1-methylpyrrolidine-2-carboxylic acid *tert*-butyl ester (14). A solution of **13** (190 mg, 0.42 mmol) in EtOH (2 mL) was treated with a catalytic amount of 10% Pd/C and placed under H₂ atmosphere (balloon). After stirring 12 h at RT, the mixture was purged with a stream of Ar, filtered over celite with EtOAc rinsing and evaporated to give the crude phenol intermediate. Purification by chromatography over silica gel (60% EtOAc/hexanes eluant) gave the pure phenol (120 mg, 78%).

Data for (2*S*,5*S*,1'*S*)-5-[1-(2-Benzyloxybenzoylamino)-2-hydroxyethyl]-1-methylpyrrolidine-2-carboxylic acid *tert*-butyl ester: ¹H NMR (300 MHz, CDCl₃) δ 12.17 (s, 1H), 7.44-7.33 (m, 3H), 6.99 (dd, J = 8.0, 1.1 Hz, 1H), 6.86 (ddd, J = 8.0, 1.1 Hz, 1H), 3.97-3.92 (m, 2H), 3.75-3.68 (m, 2H), 3.52-3.47 (m, 1H), 2.28-2.01 (m, 2H), 1.94-1.87 (m, 1H), 1.72-1.62 (m, 1H), 1.47 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ 172.5, 170.9, 161.4, 134.3, 125.6, 118.7, 114.24, 81.4, 67.5, 64.0, 63.9, 54.0, 36.1, 28.1, 28.0, 25.7; ESI-MS (m/z) [MH]⁺ calcd for C₁₉H₂₈N₂O₅ 365.20, found 365.34.

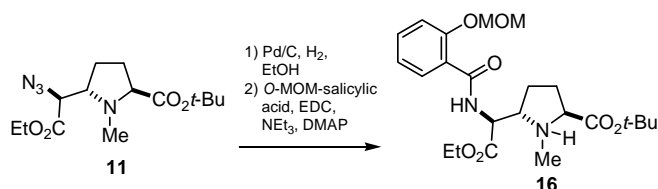
A solution of the above phenol (130 mg, 0.36 mmol) in THF (2mL) was treated with 1.5 equiv. of Burgess reagent, stirred for 30min at RT, then at 60 °C for an additional 30min. The mixture was concentrated and the crude residue purified by chromatography over silica gel (60% EtOAc/hexanes eluant) to give **14** as a white solid (110 mg, 89%). ¹H NMR (300 MHz, CDCl₃) δ 12.24 (s, 1H), 7.64 (dd, J = 7.9, 1.5 Hz, 1H), 7.37 (ddd, J = 8.5, 7.3, 1.8 Hz, 1H), 7.00 (m, 1H), 6.86 (ddd, J = 7.9, 7.3, 1.2 Hz, 1H), 4.49 (m, 2H), 4.22 (m, 1H), 3.65 (dd, J = 7.9, 1.8 Hz, 1H), 3.34 (m, 1H), 2.48 (s, 3H), 2.22 (m, 1H), 2.02 (m, 1H), 1.78 (ddt, 12.9, 8.8, 2.1 Hz, 1H), 1.58 (m, 1H), 1.47 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ 173.6, 165.7, 159.9, 133.3, 128.0, 118.6, 116.7, 110.5, 80.8, 69.0, 68.7, 68.5, 65.4, 36.7, 28.2, 28.1, 26.7; mp = 75-78 °C; HRMS (ESI-TOF) (m/z) [MH]⁺ calcd for C₁₉H₂₆N₂O₄ 347.1965, found 347.1974.



(2*S*,5*S*,4'*S*)-5-[2-(2-Hydroxyphenyl)-4,5-dihydrothiazol-4-yl]-1-methylpyrrolidine-2-carboxylic acid *tert*-butyl ester (15). A solution of **14** (120 mg, 0.35 mmol) was dissolved in 10 mL of 1:1 NEt₃:MeOH at RT in a pressure flask. Hydrogen sulfide gas was then bubbled through the stirring solution for 20 min, after which the pressure flask was closed and warmed to 40 °C. The reaction was monitored periodically by LCMS and stirred at 40 °C for 5d (> 90% consumption of starting material). The reaction was then purged of H₂S by bubbling a stream of

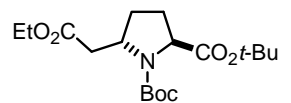
Ar through the solution for 15 min. Concentration and drying under vacuum gave a yellow solid which was used directly in the subsequent cyclodehydration.

The crude thioamide was taken up in 4 mL of THF, treated with Burgess reagent (124 mg, 0.52 mmol) and heated to 60 °C. After 1 hr, LCMS indicated complete consumption of the thioamide intermediate. The reaction was cooled to room temperature and filtered through a short silica pad with EtOAc rinsing. The filtrate was concentrated in vacuo and the crude residue purified by chromatography over silica gel (15% EtOAc/hexanes eluant) to give **15** as an off-white solid (48 mg, 38% from **14**). ¹H NMR (400 MHz, CDCl₃) δ 12.80 (bs, 1H), 7.40-7.30 (m, 2H), 6.98 (dd, J = 8.2, 0.8 Hz, 1H), 6.86 (m, 1H), 4.81 (ddd, J = 10.2, 9.0, 3.1 Hz, 1H), 3.67 (dd, J = 7.8, 1.6 Hz, 1H), 3.52 (dt, J = 9.4, 3.9 Hz), 3.34 (dd, J = 10.5, 8.6 Hz, 1H), 3.21 (t, J = 10.5, 1H), 2.51 (s, 3H), 2.25 (dq, J = 12.9, 9.4 Hz, 1H), 2.03 (m, 1H), 1.79 (ddt, J = 12.9, 9.0, 2.0 Hz, 1H), 1.67 (m, 1H), 1.47 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 173.9, 171.5, 159.4, 133.0, 130.6, 118.9, 117.2, 116.6, 81.0, 79.6, 68.6, 66.0, 36.7, 33.0, 28.5, 28.4, 27.0; mp = 133 °C; HRMS (ESI-TOF) (m/z) [MH]⁺ calcd for C₁₉H₂₆N₂O₃S 363.1737, found 363.1753.

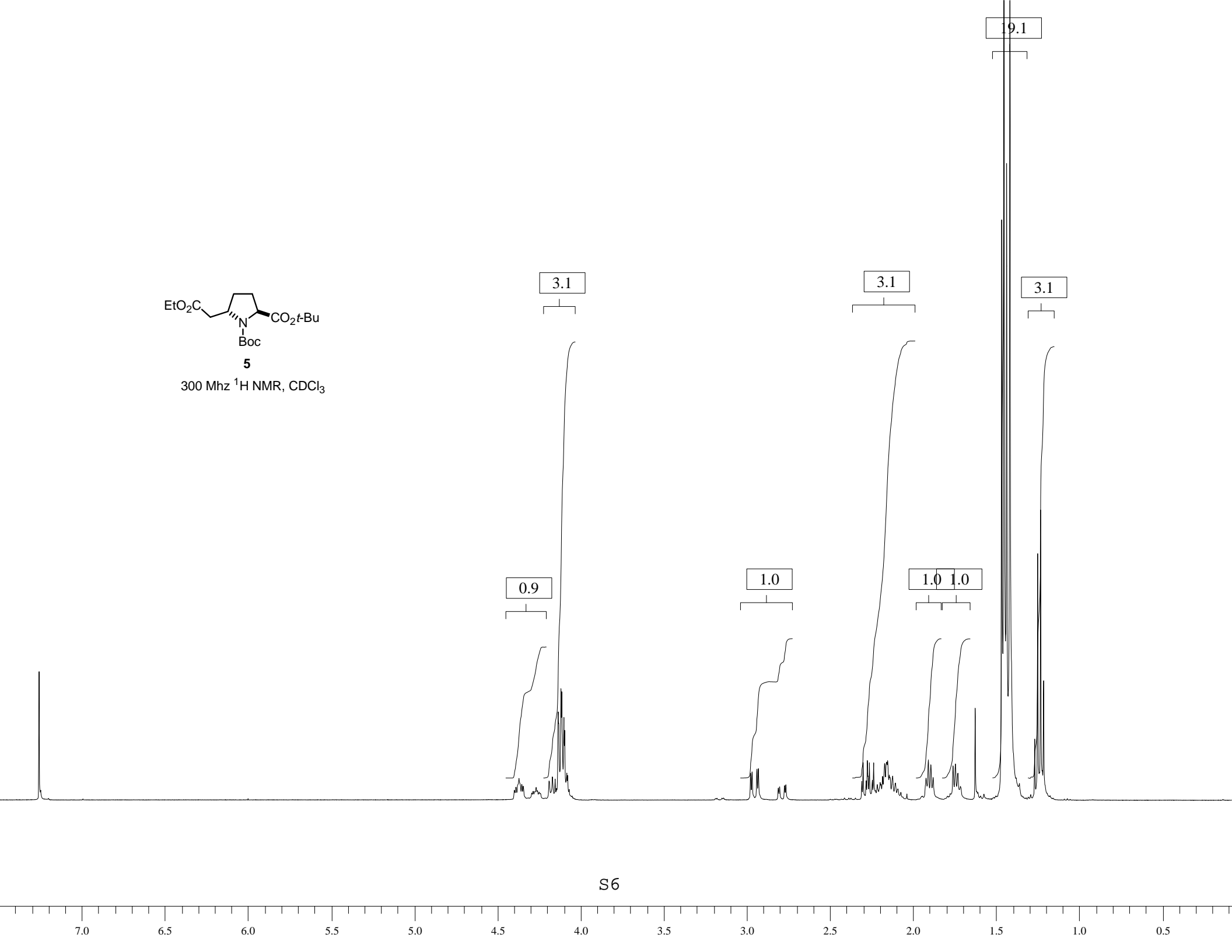


(2S,5S,1'S)-5-[(2-Methoxymethoxybenzoylamino)-ethoxycarbonylmethyl]-1-

methylpyrrolidine-2-carboxylic acid *tert*-butyl ester (16): Amide **16** was prepared from **11** (700 mg, 2.23 mmol) following the procedure described for the synthesis of **12**, and using 2-methoxymethoxybenzoic acid in place of 2-benzyloxybenzoic acid. The crude amide was purified by chromatography over silica gel (35% EtOAc/hexane eluant) to give **16** as a white solid (850 mg, 84% from **11**). ¹H NMR (300 MHz, CDCl₃) δ 8.47 (d, J = 5.3 Hz, 1H), 8.19 (dd, J = 7.9, 1.8 Hz, 1H), 7.43 (ddd, J = 8.5, 7.3, 1.8 Hz, 1H), 7.13 (m, 2H), 5.31 (m, 2H), 4.79 (dq, J = 7.0, 1.5 Hz, 2H), 3.65 (d, J = 6.7 Hz, 1H), 3.58 (m, 1H), 3.54 (s, 3H), 2.50 (s, 3H), 2.19-1.80 (m, 4H), 1.48 (s, 9H), 1.29 (t, J = 7.0 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 173.2, 171.2, 165.3, 155.4, 132.8, 132.3, 122.3, 122.0, 114.6, 95.0, 81.1, 67.6, 62.9, 61.1, 56.6, 54.0, 35.0, 28.2, 27.6, 25.4, 14.2; mp = 79-81 °C; HRMS (ESI-TOF) (m/z) [MH]⁺ calcd for C₂₃H₃₄N₂O₇ 451.2444, found 451.2431.



300 Mhz ^1H NMR, CDCl_3



171.916
171.396

153.955
153.643

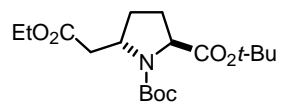
80.981
80.061
79.911

60.443
60.297
54.715
54.686

39.428
38.447

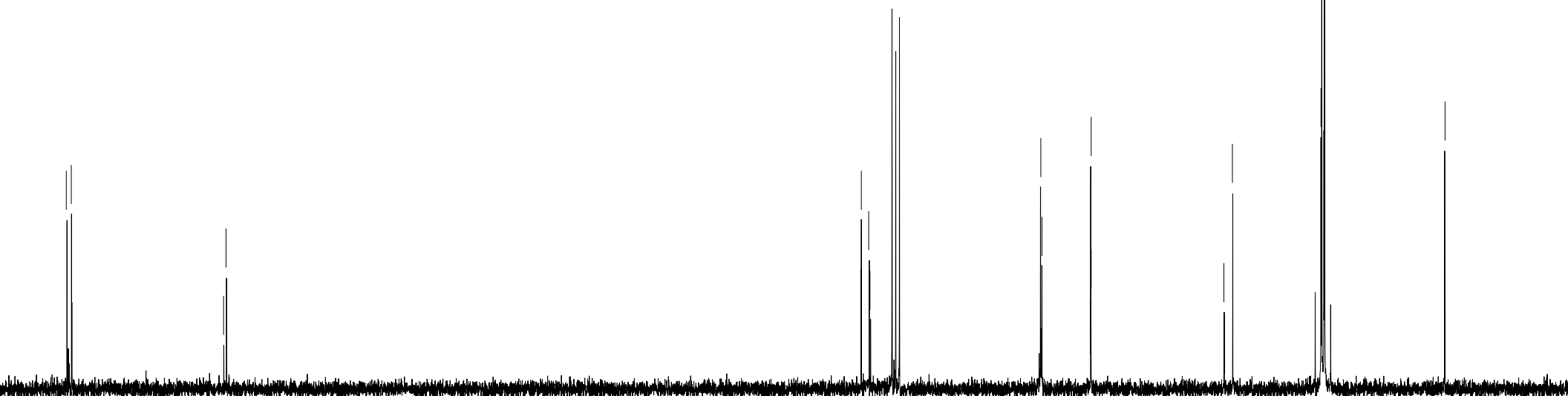
28.360
28.270
27.975
27.915

14.186

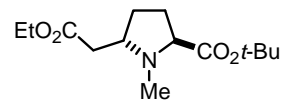


5

75 Mhz ¹³C NMR, CDCl₃

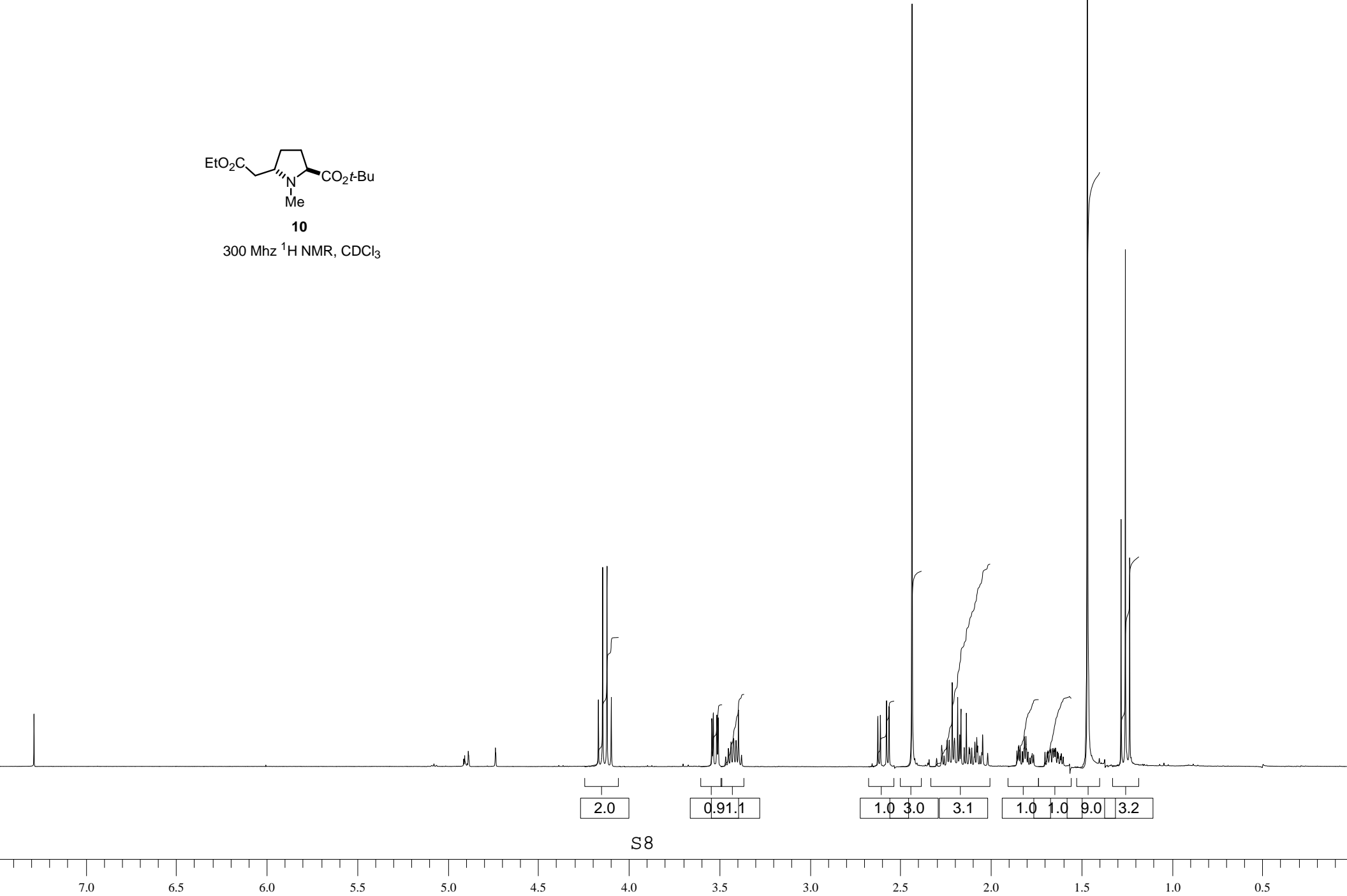


170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10



10

300 Mhz ^1H NMR, CDCl_3



173.342
172.228

80.729

66.987

60.228

59.693

38.805

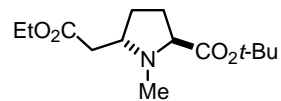
35.269

29.921

28.183

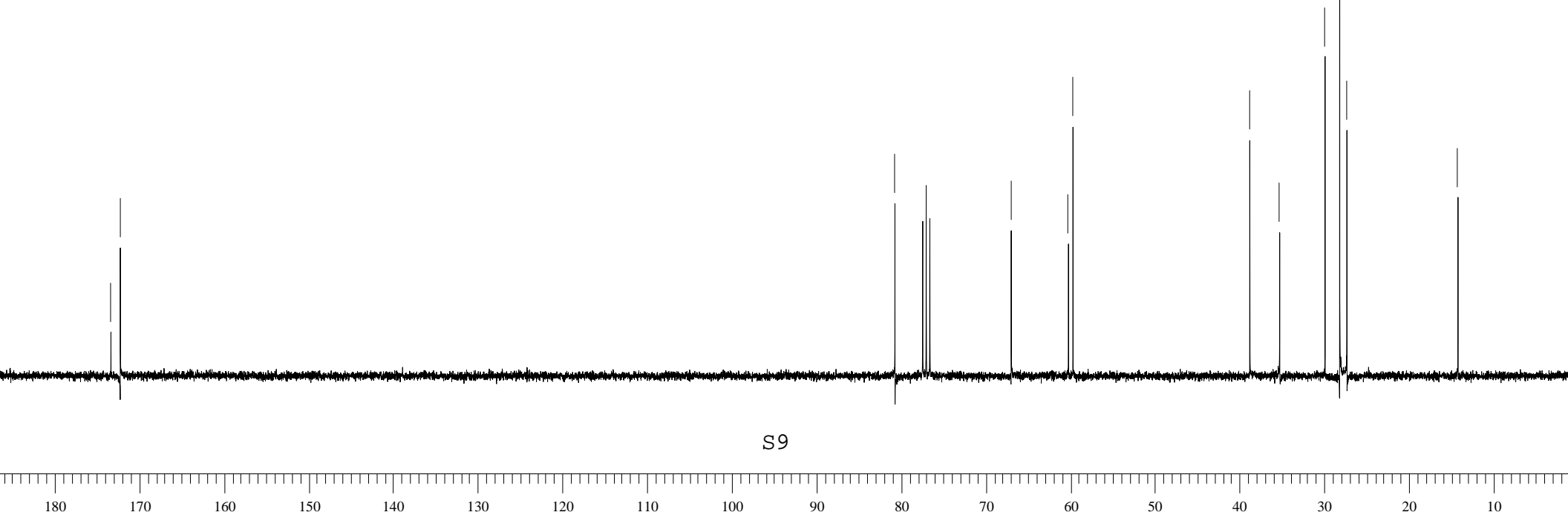
27.336

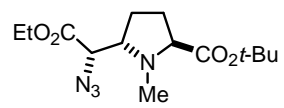
14.218



10

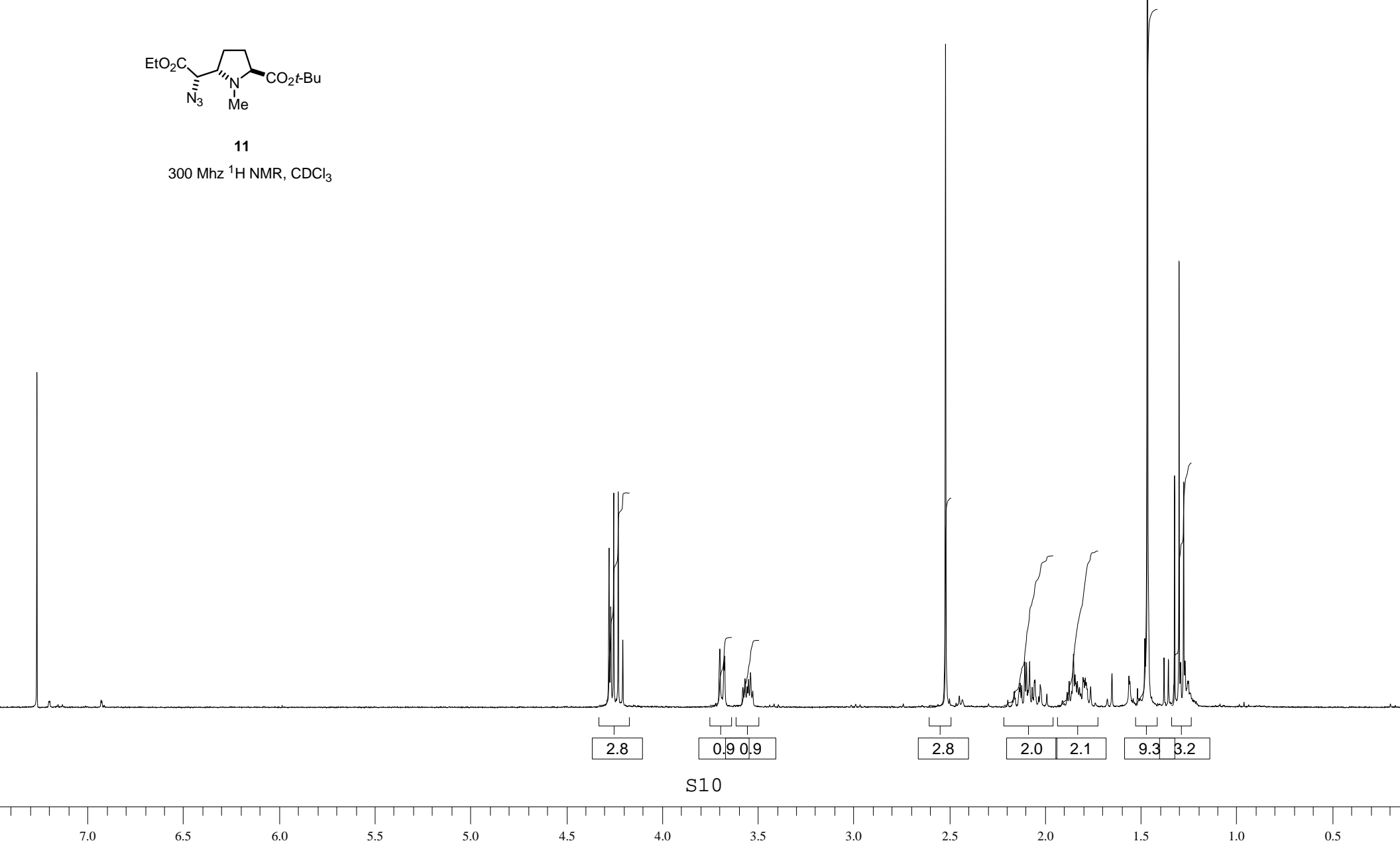
75 Mhz ¹³C NMR, CDCl₃

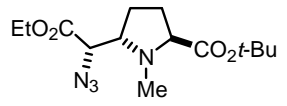




11

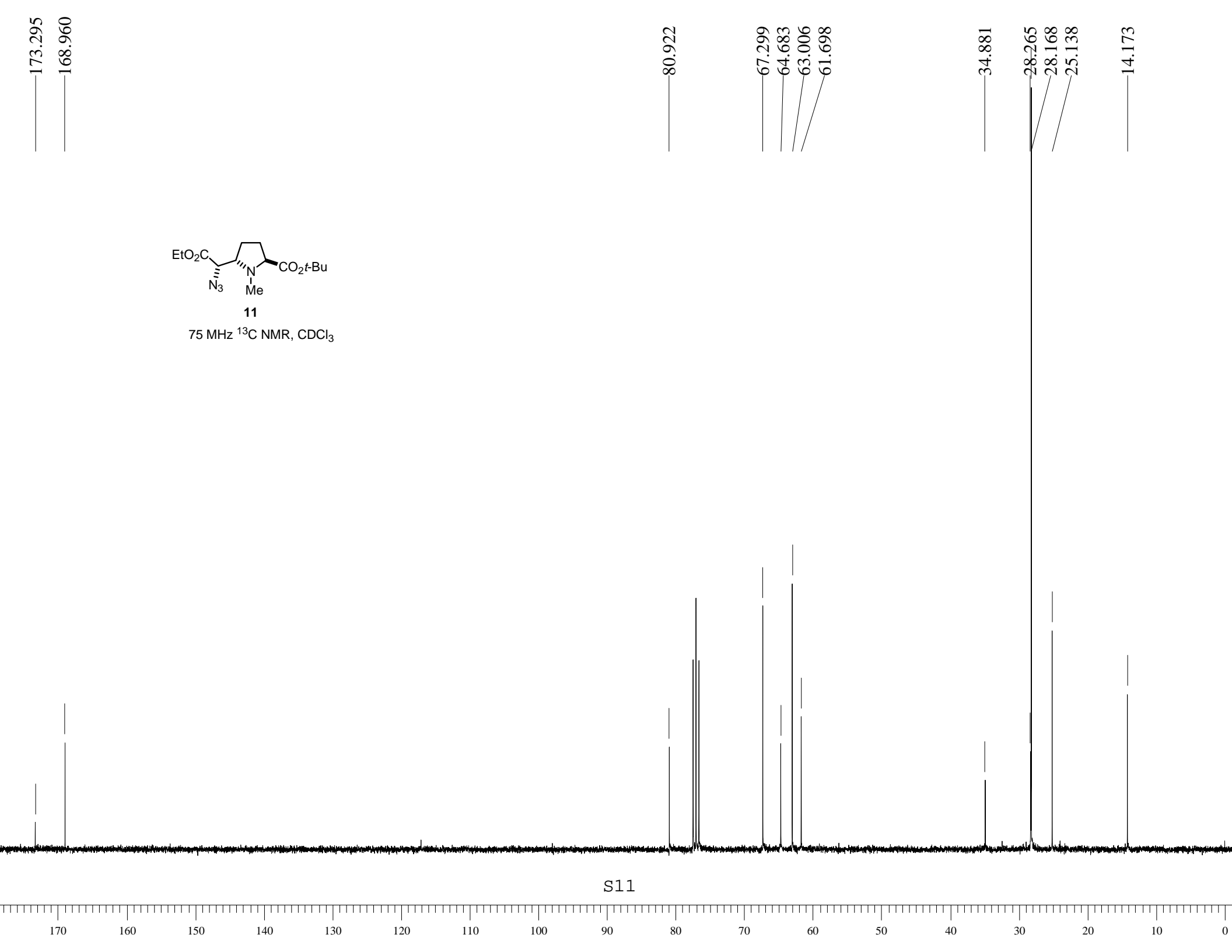
300 Mhz ¹H NMR, CDCl₃





11

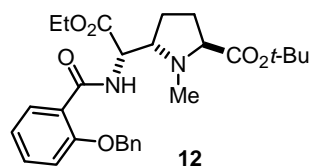
75 MHz ¹³C NMR, CDCl₃



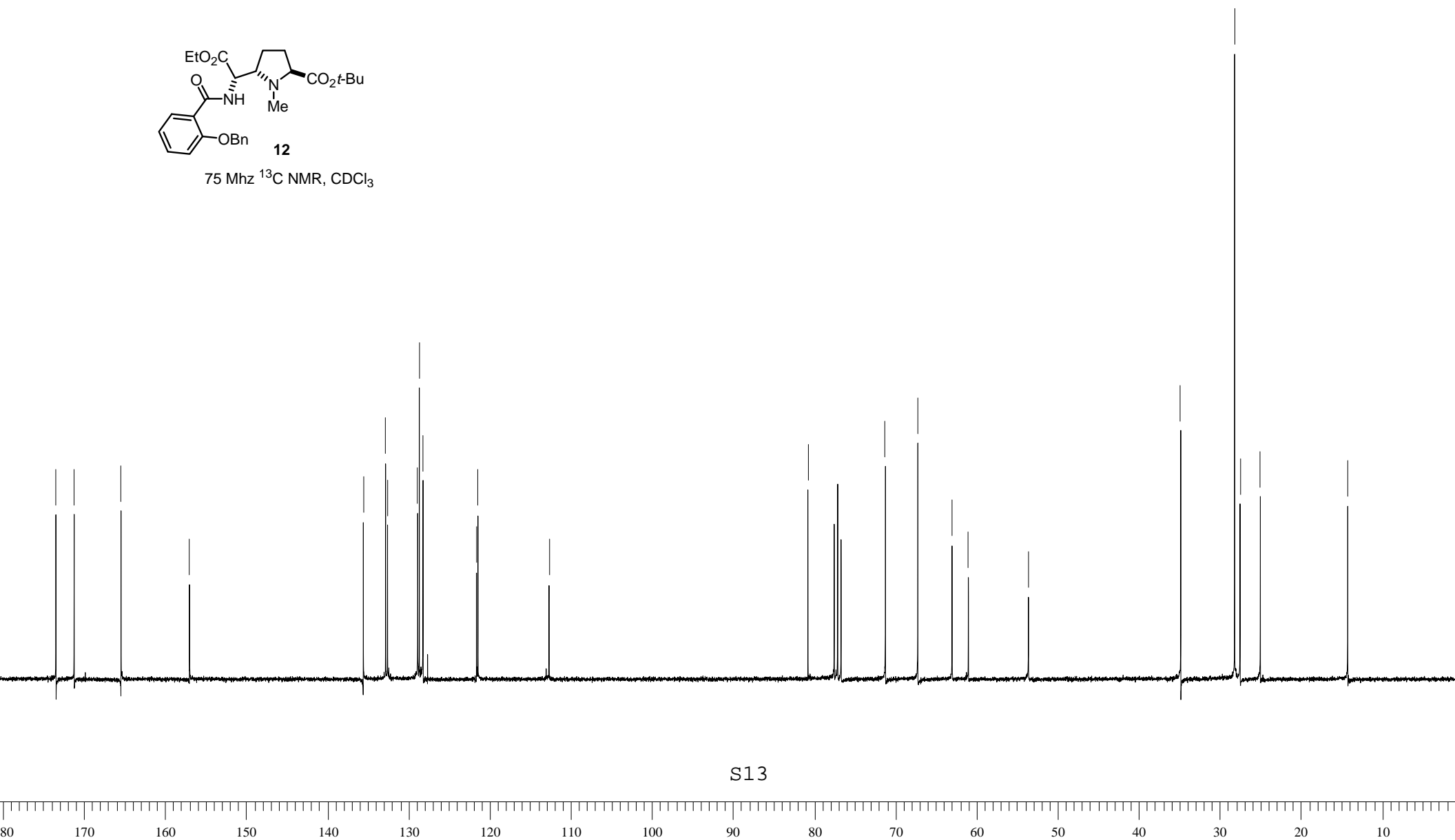
173.446
171.203
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132.815
132.576
128.860
128.670
128.209
121.613
121.434
112.665

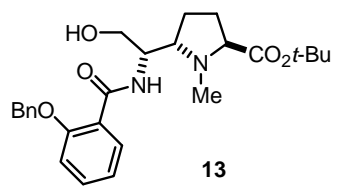
80.774
71.221
67.224
63.004
60.996
53.595

34.824
28.183
27.514
25.018
14.248

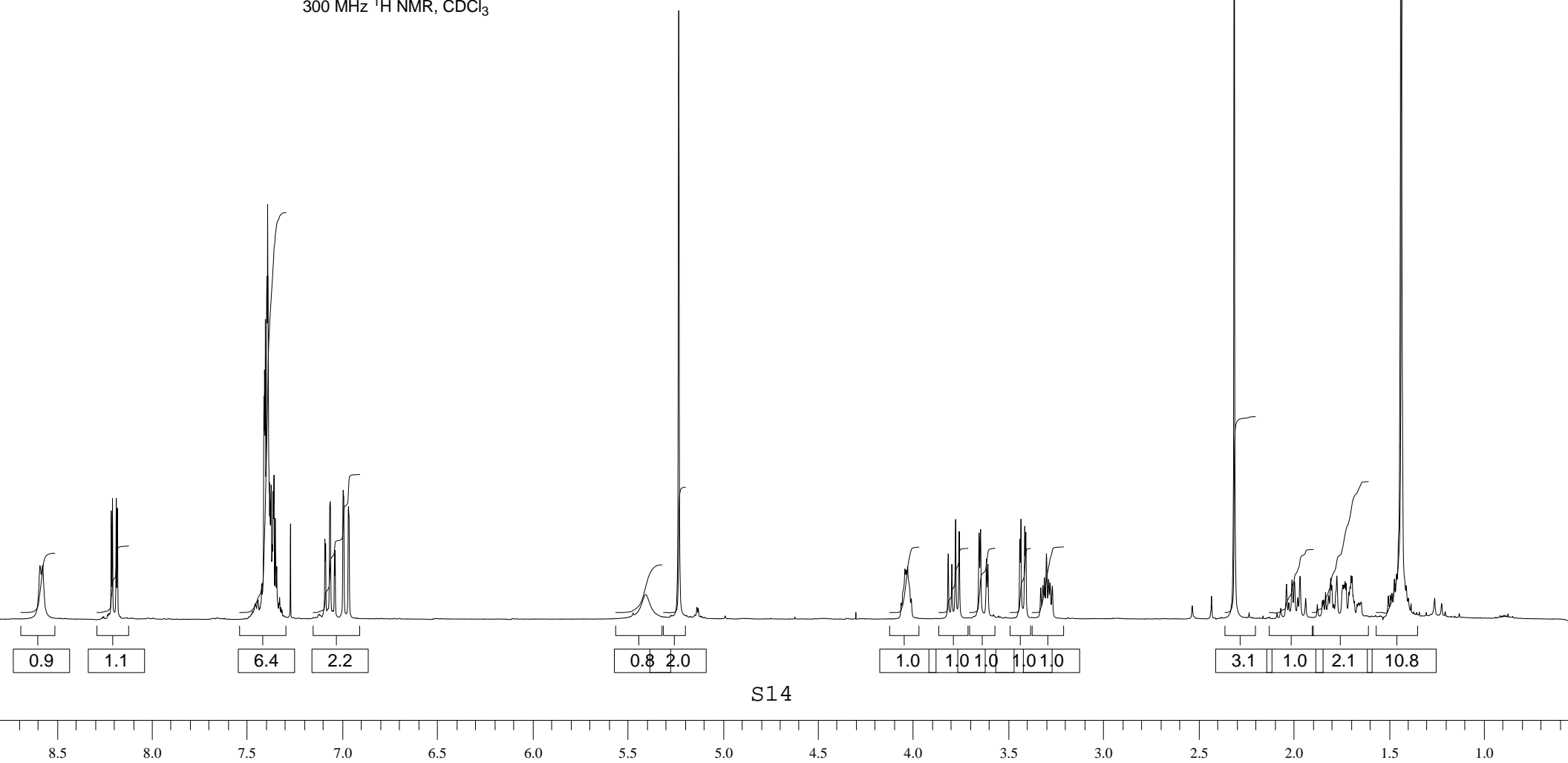


75 Mhz ^{13}C NMR, CDCl_3





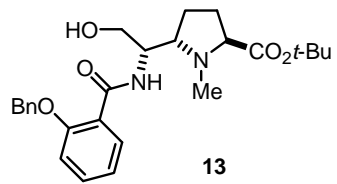
300 MHz ¹H NMR, CDCl₃



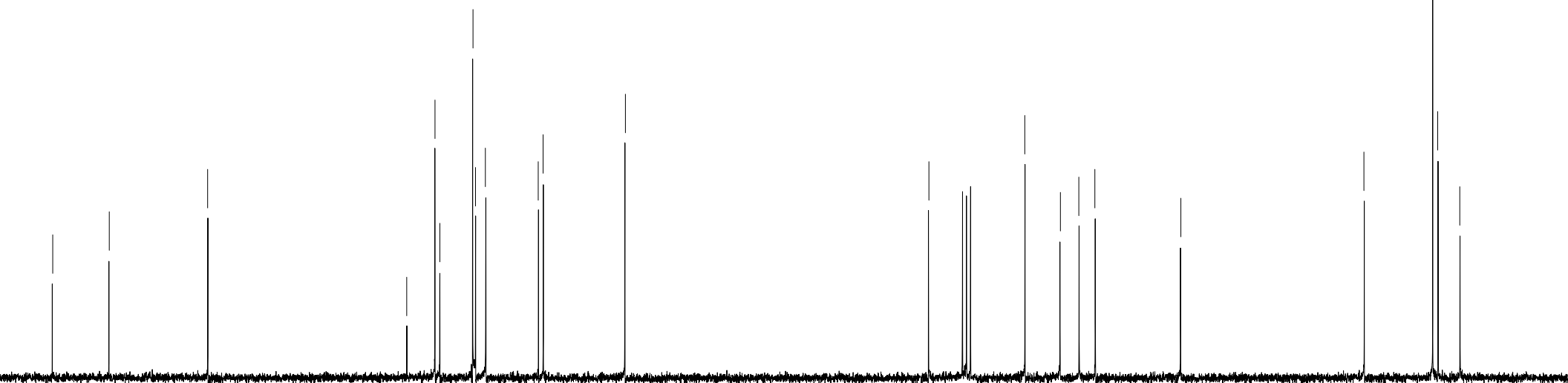
172.956
167.013
156.644
135.770
132.829
132.304
128.877
128.565
127.494
121.970
121.464
112.907

81.070
70.954
67.284
65.279
63.585
54.641

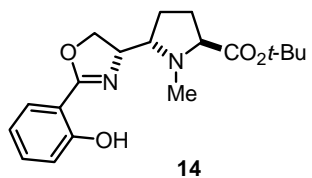
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28.198
27.634
25.329



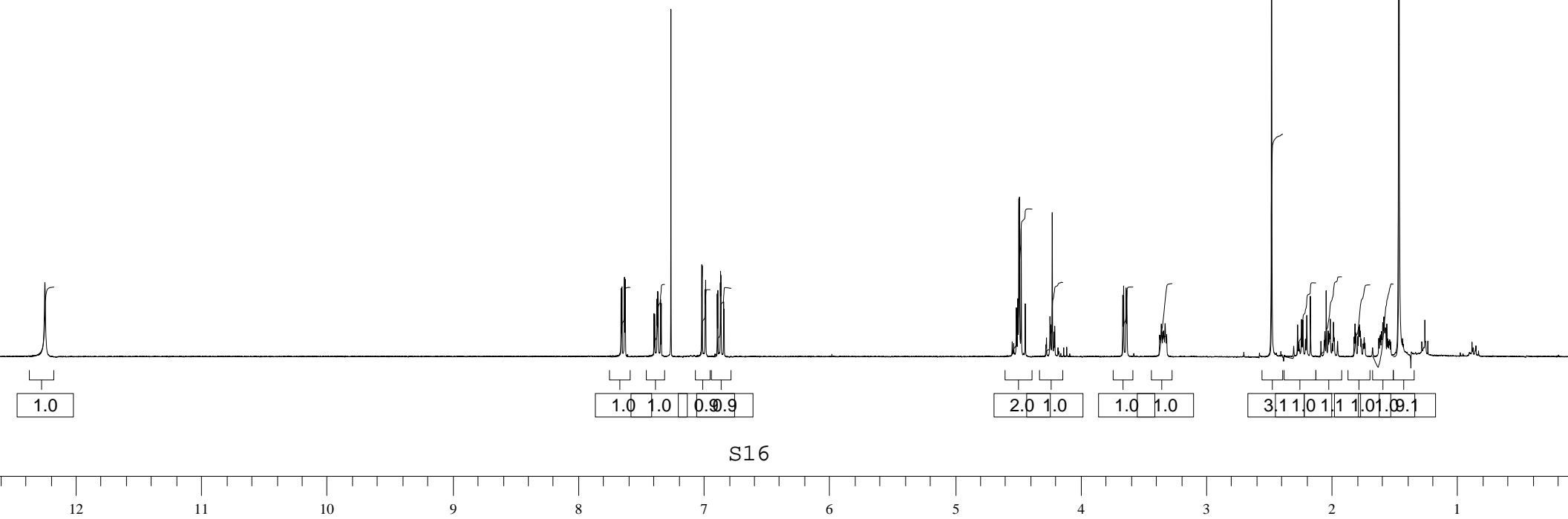
75 MHz ^{13}C NMR, CDCl_3



170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20



300 MHz ^1H NMR, CDCl_3



173.580
165.735
159.957

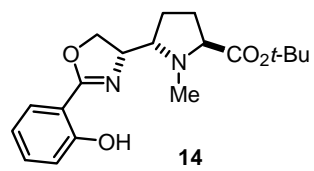
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80.818

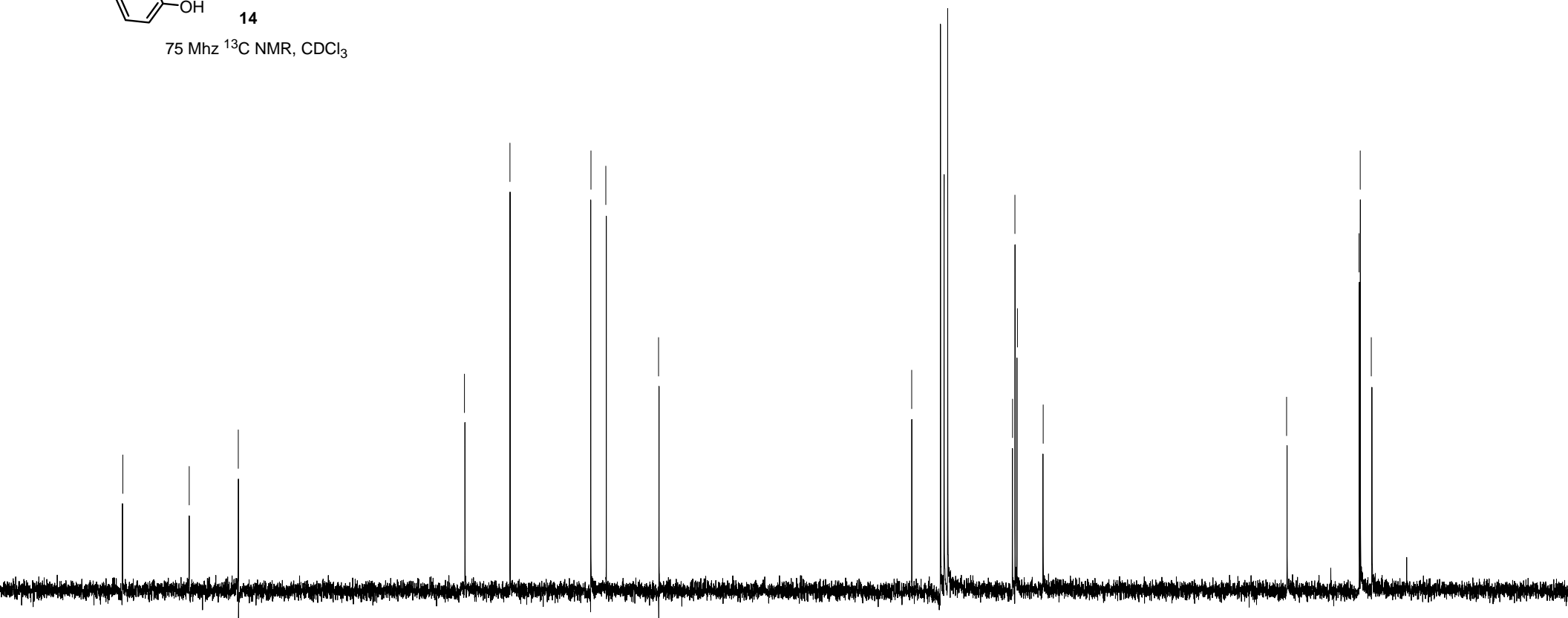
68.991
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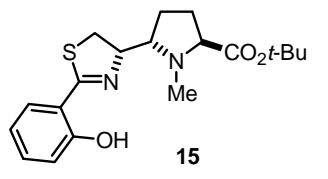
36.710

28.207
28.108
26.742

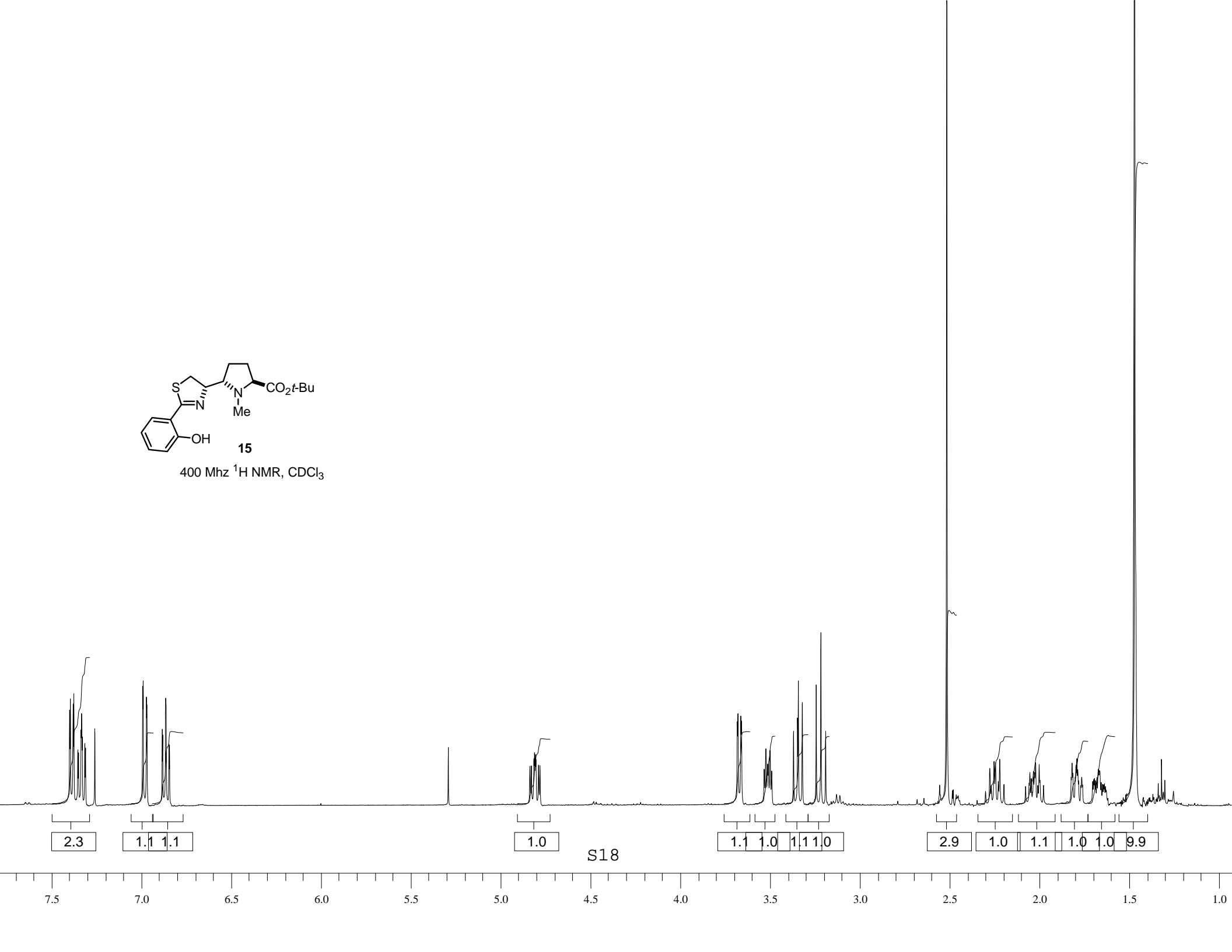


75 Mhz ^{13}C NMR, CDCl_3





400 Mhz ¹H NMR, CDCl₃



173.861
171.537

159.392

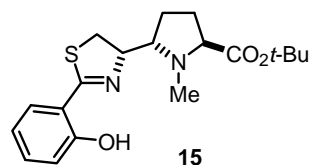
133.042
130.608

118.929
117.226
116.579

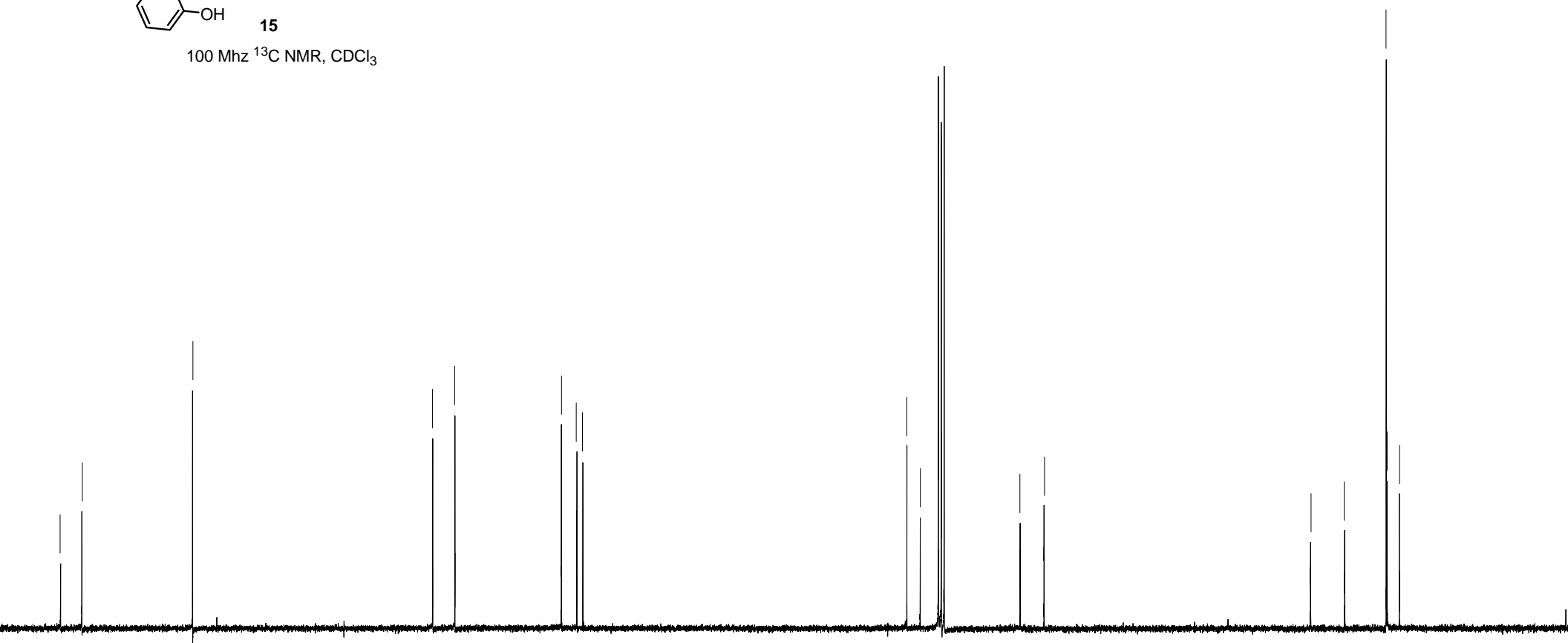
81.029
79.569

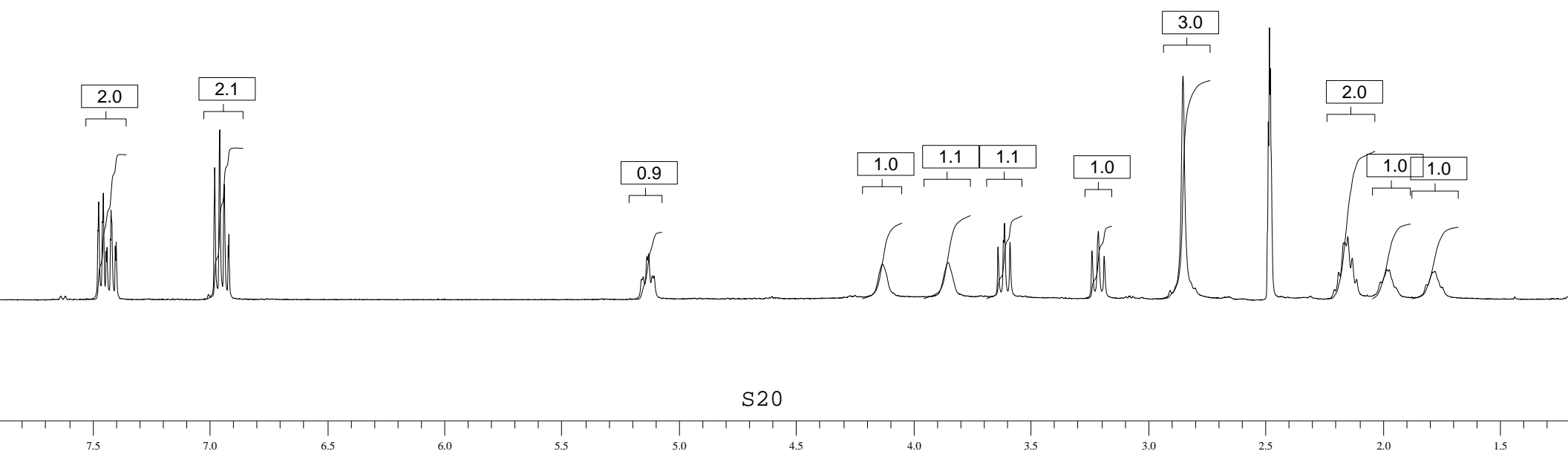
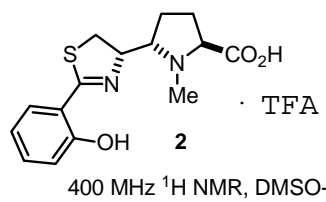
68.609
65.985

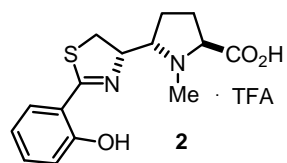
36.743
33.005
28.451
28.352
27.016



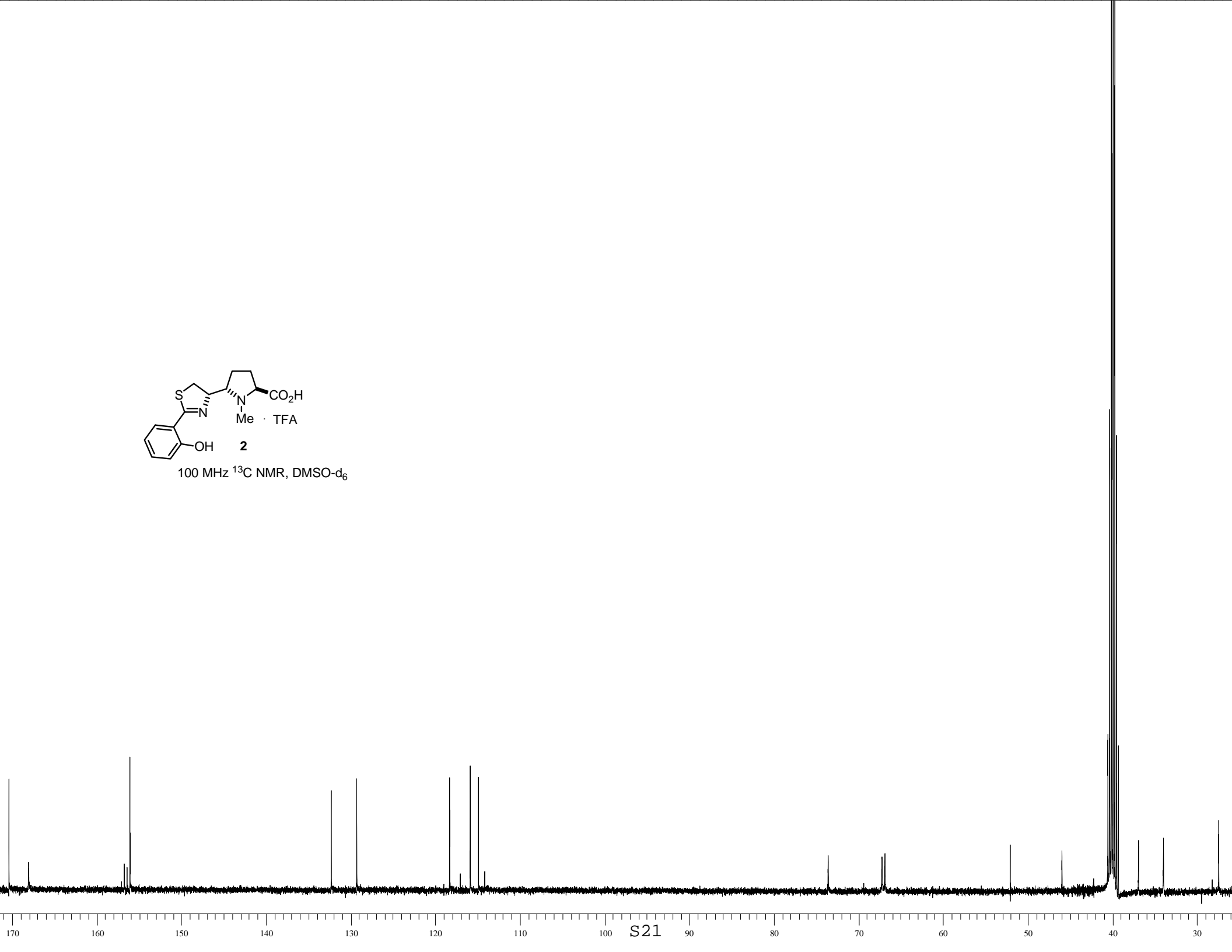
100 Mhz ¹³C NMR, CDCl₃

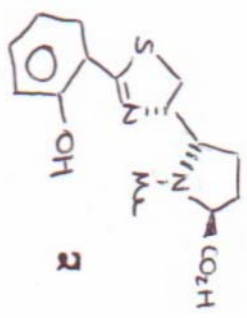
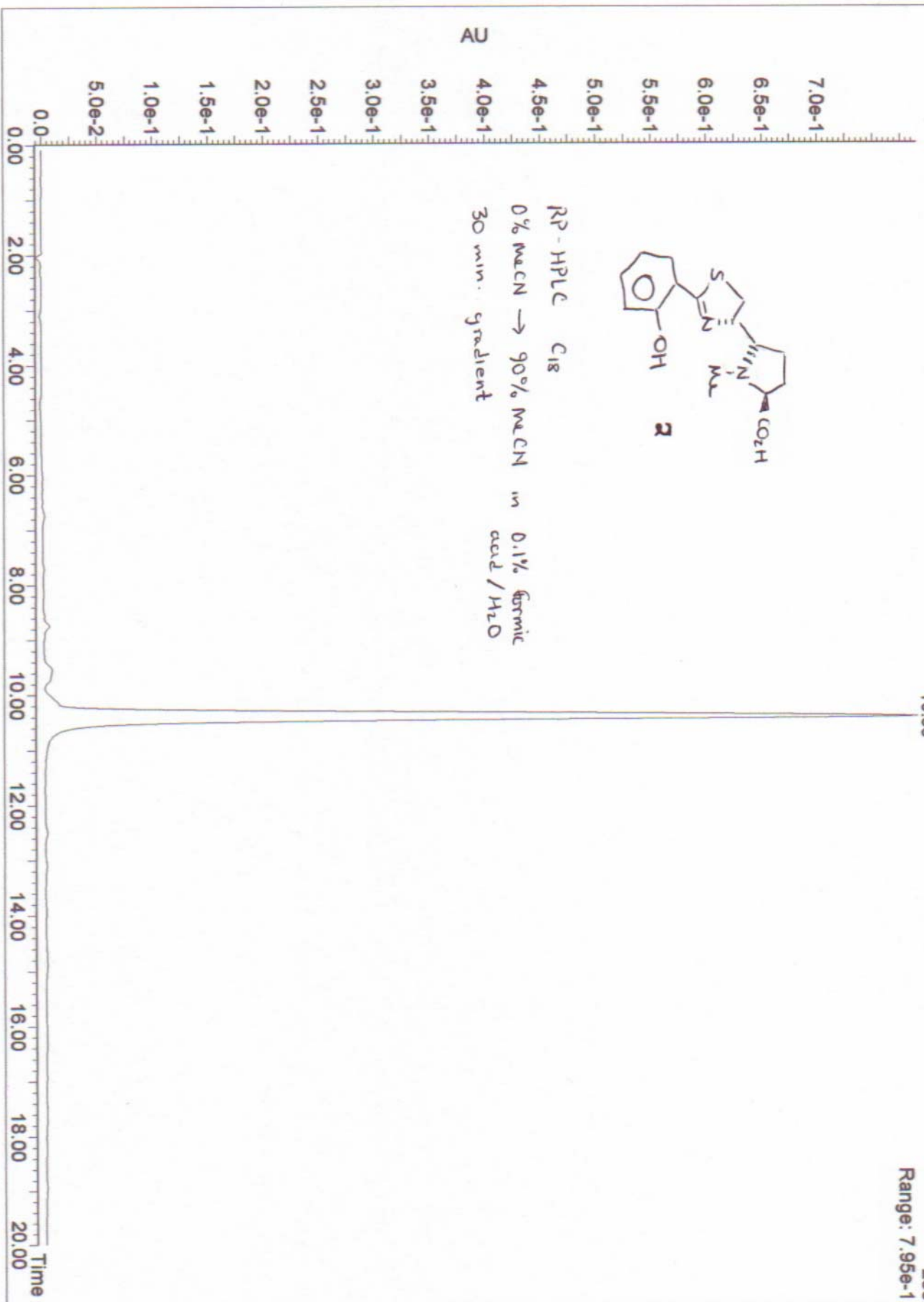






100 MHz ^{13}C NMR, DMSO- d_6



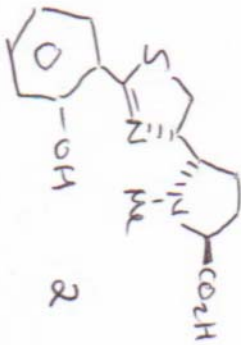


RP-HPLC C18
0% MeCN → 90% MeCN in 0.1% formic acid / H₂O
30 min. gradient

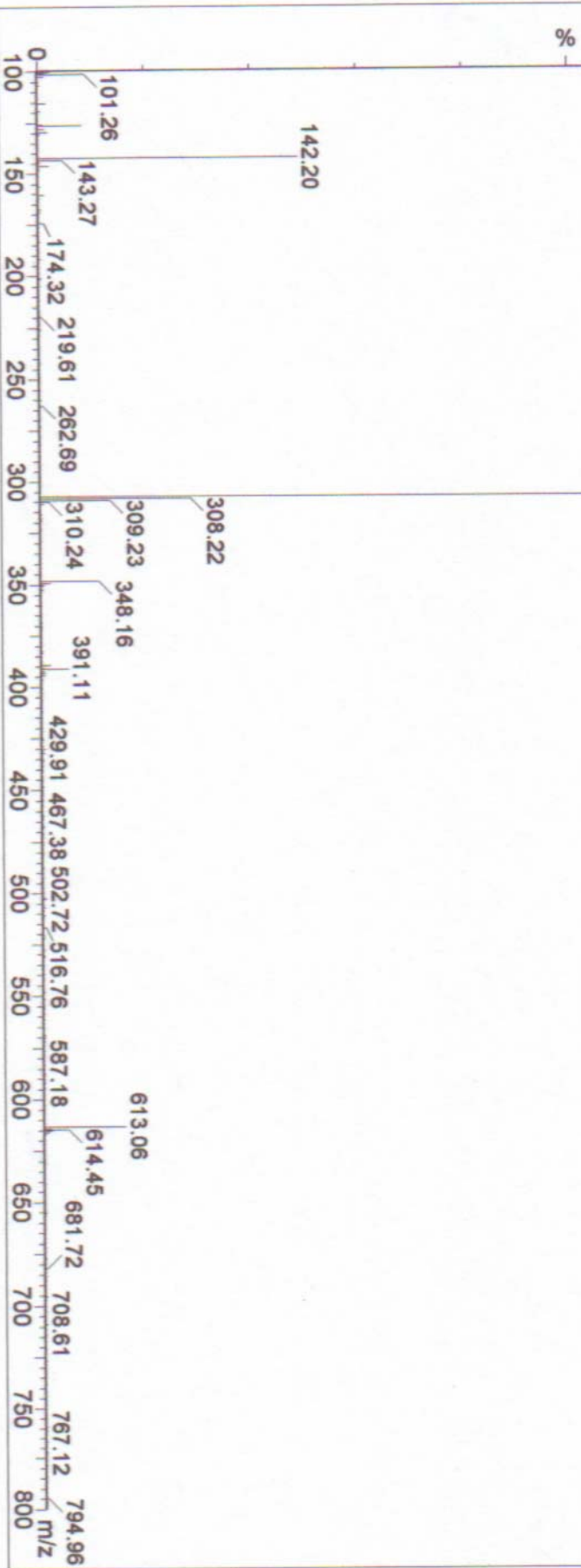
JD-02-30B 1030 (10.386)

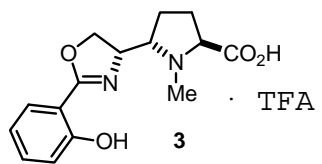
307.15

1: Scan ES+
3.11e6

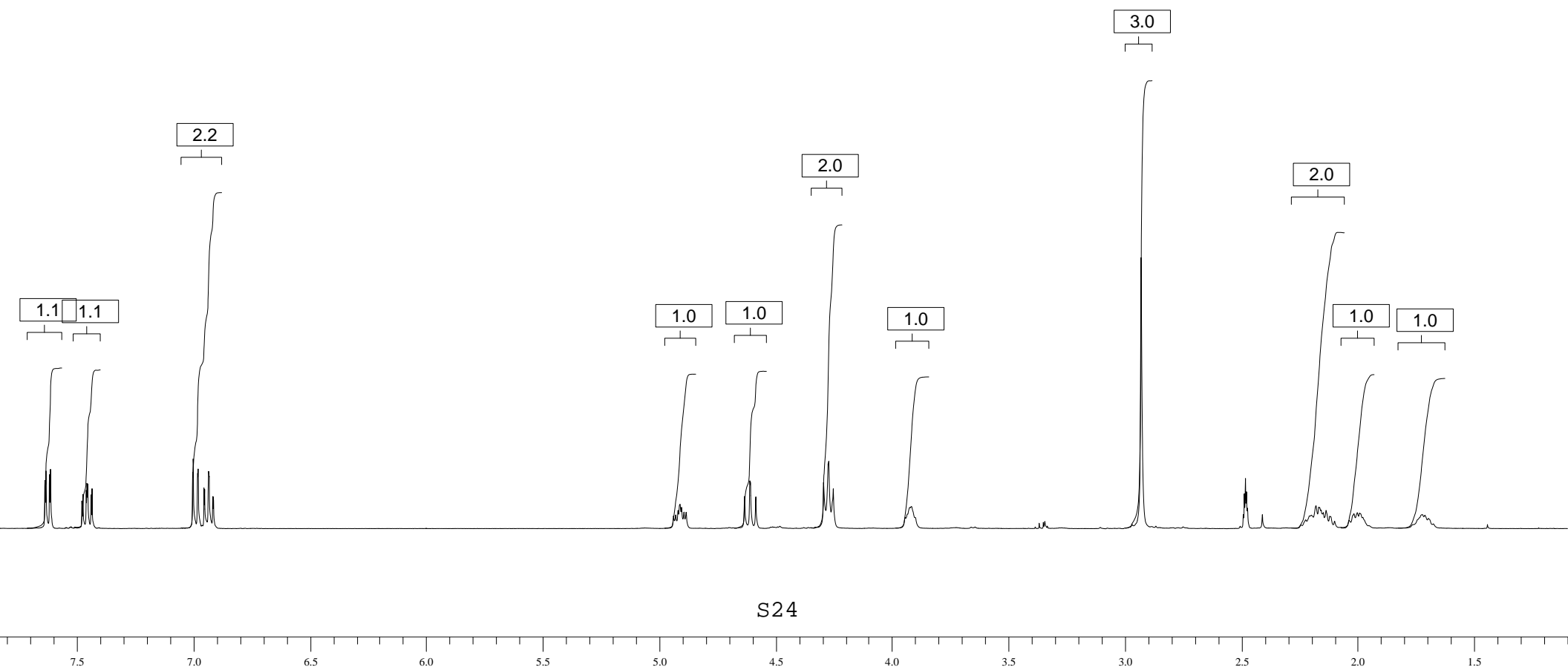


ESI-MS (+) For RT = 10.386 min.

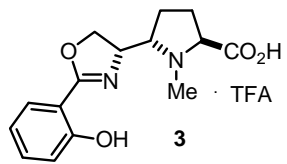




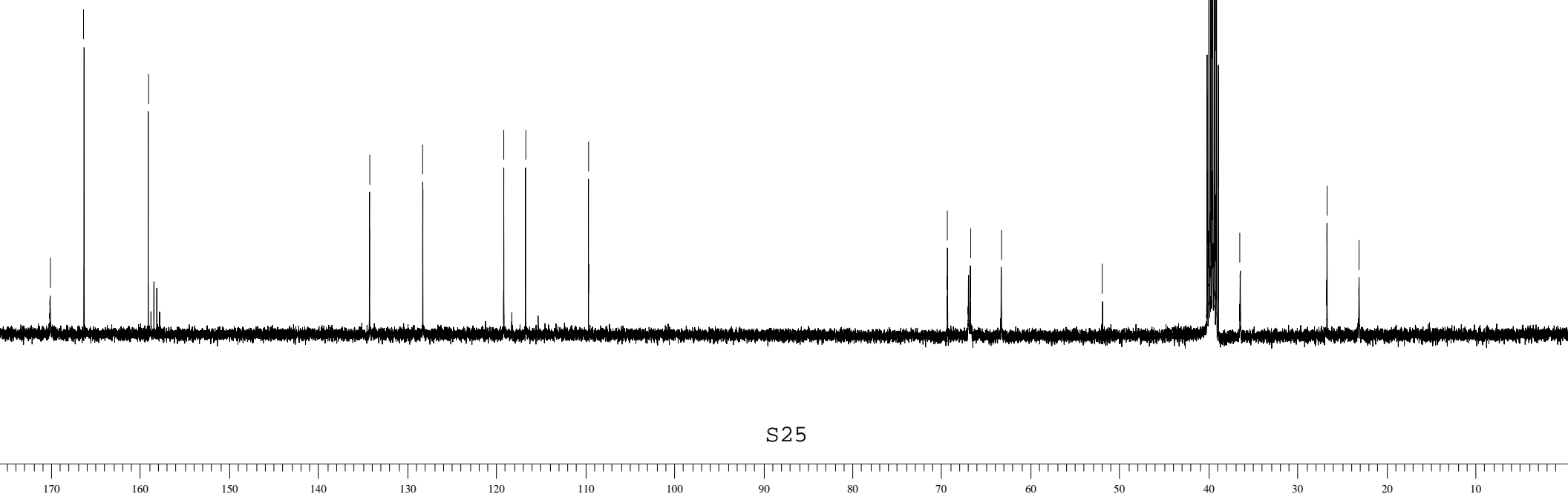
400 MHz ¹H NMR, DMSO-d₆



170.096
166.276
159.075
134.220
128.237
119.137
116.694
109.625
69.305
66.737
63.270
51.875
36.402
26.670
23.068

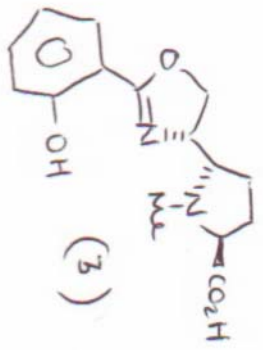
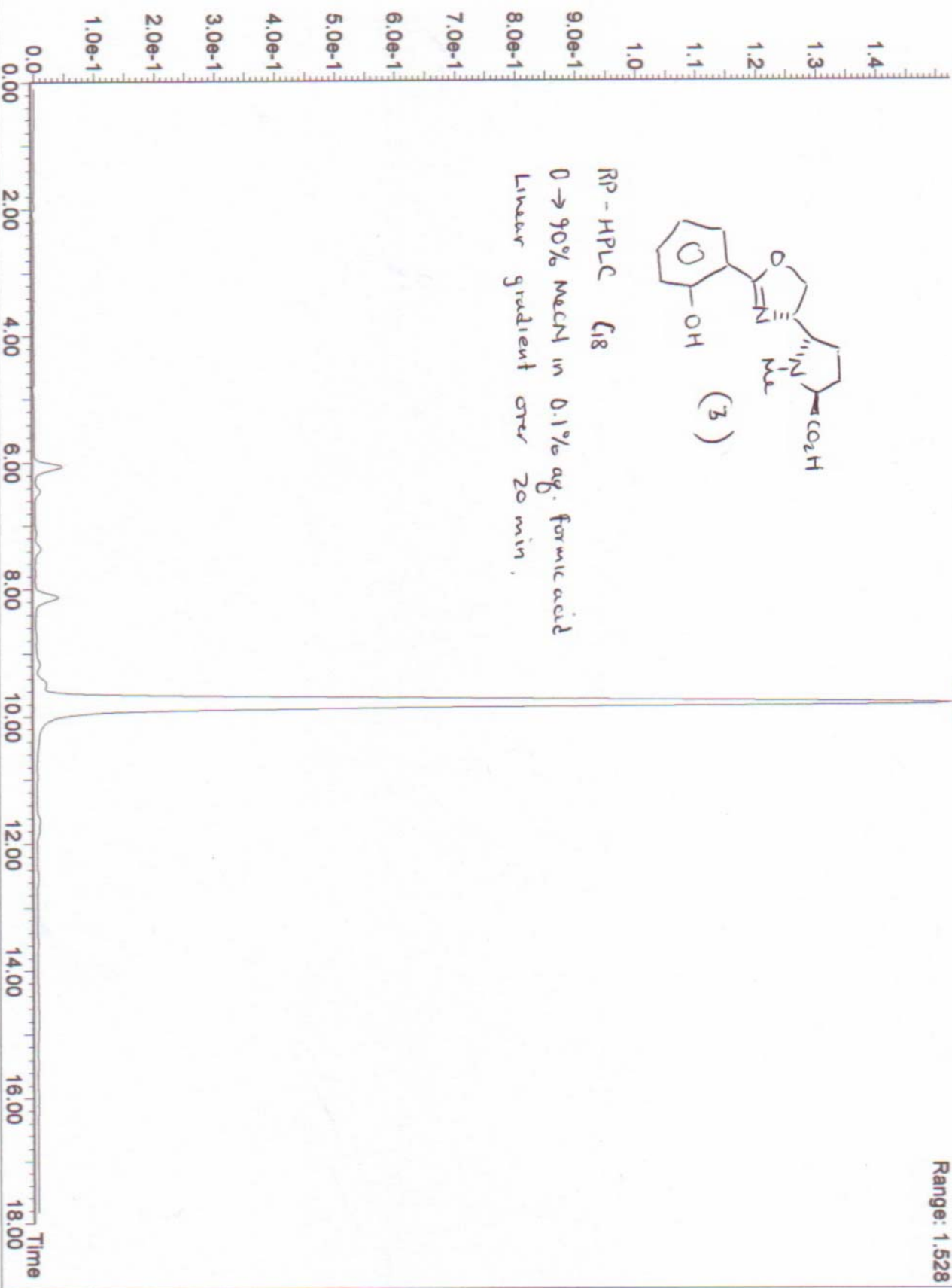


100 MHz ¹³C NMR, DMSO-d₆



JD-02-31

2: Diode Array
280
Range: 1.528



RP-HPLC (18)

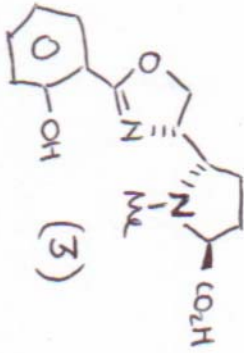
0 → 90% MeCN in 0.1% aq. formic acid
Linear gradient over 20 min.

AU

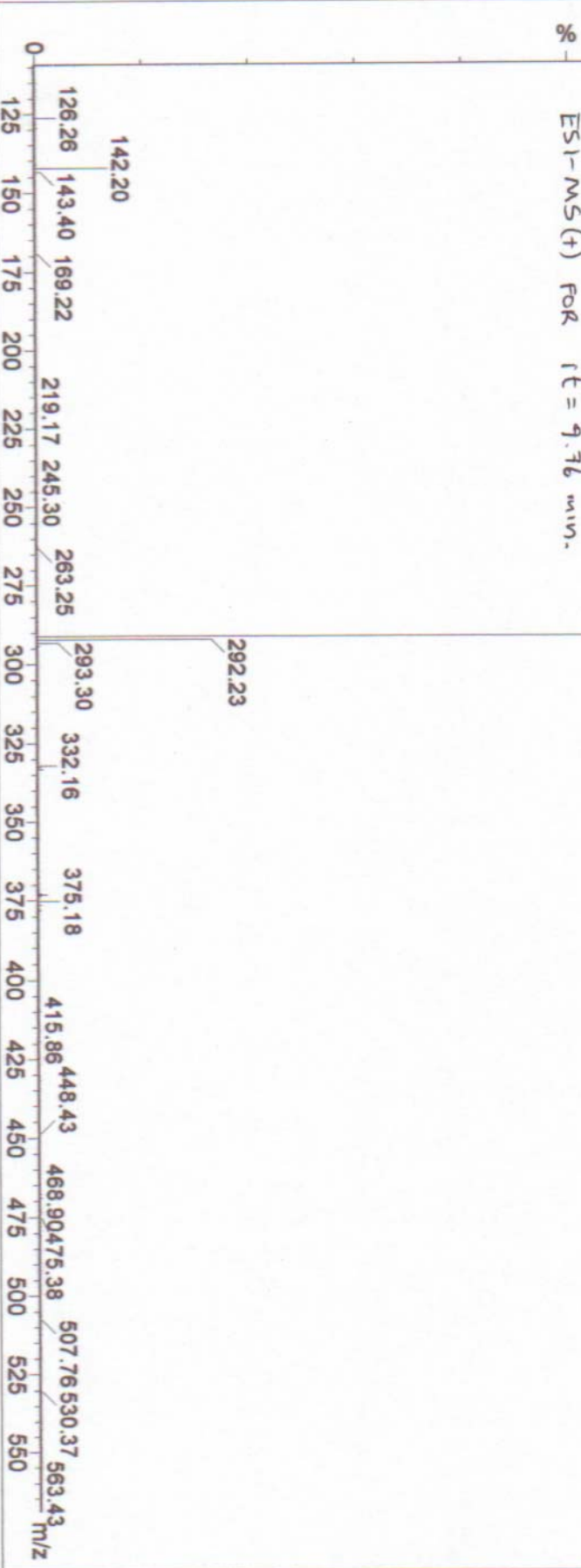
JD-02-31 968 (9.761)

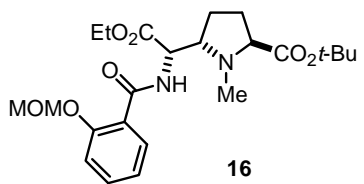
291.16

1: Scan ES+
6.80e6

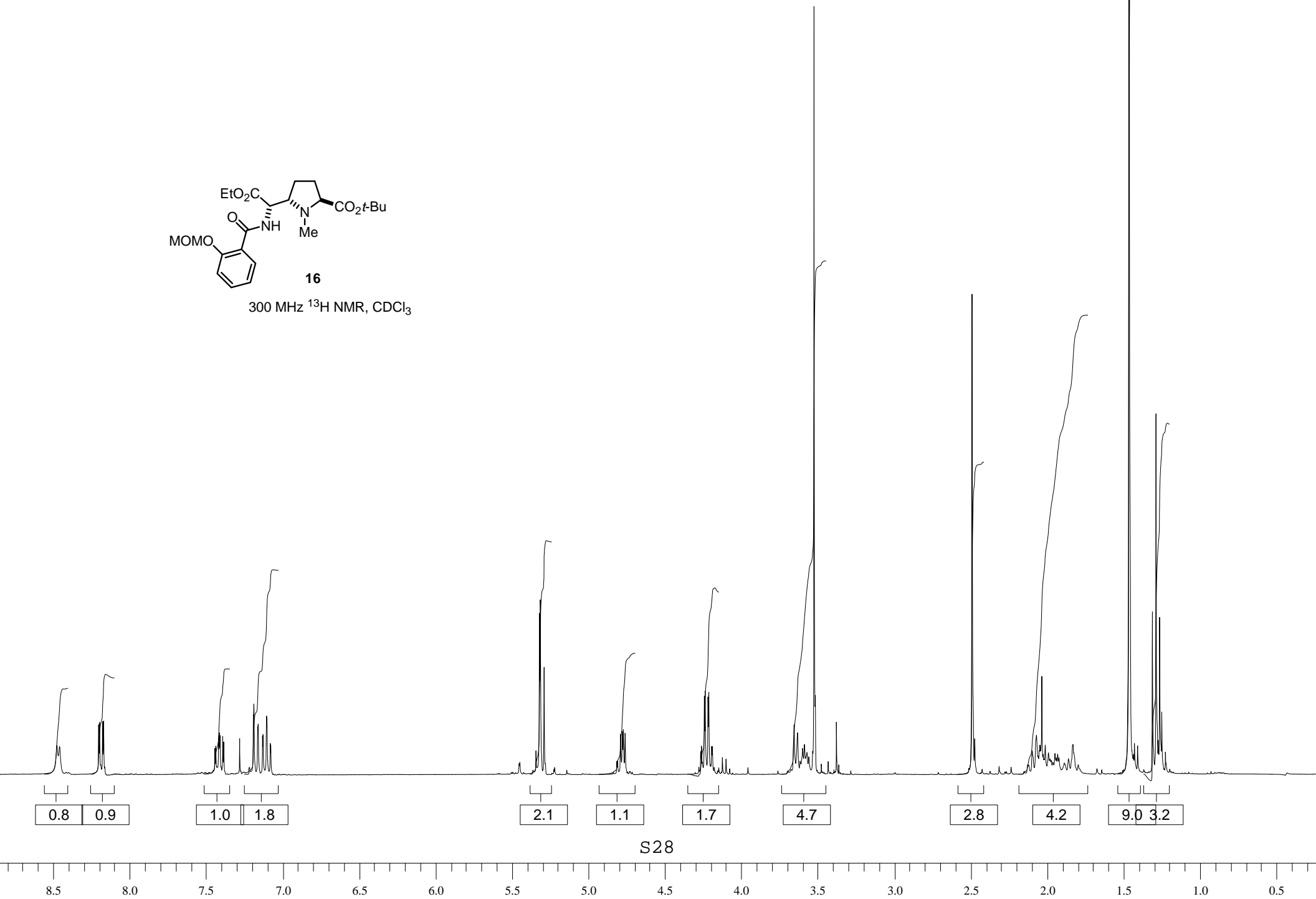


RP-HPLC C₁₈
ESI-MS(+) FOR *t*_r = 9.76 min.





300 MHz ¹H NMR, CDCl₃



173.238
171.157
165.349
155.396

132.784
132.250
122.251
121.999
114.616

94.961

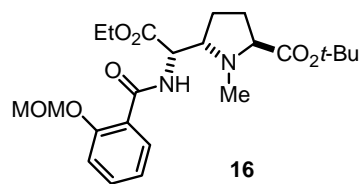
81.056

67.641
62.929
61.089
56.558
54.033

34.987

28.197
27.573
25.420

14.231



75 MHz ^{13}C NMR, CDCl_3

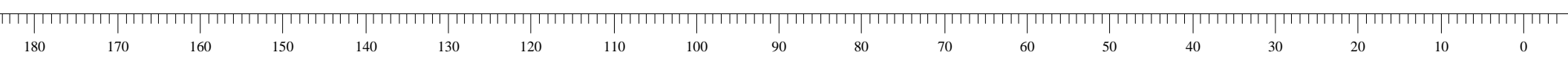
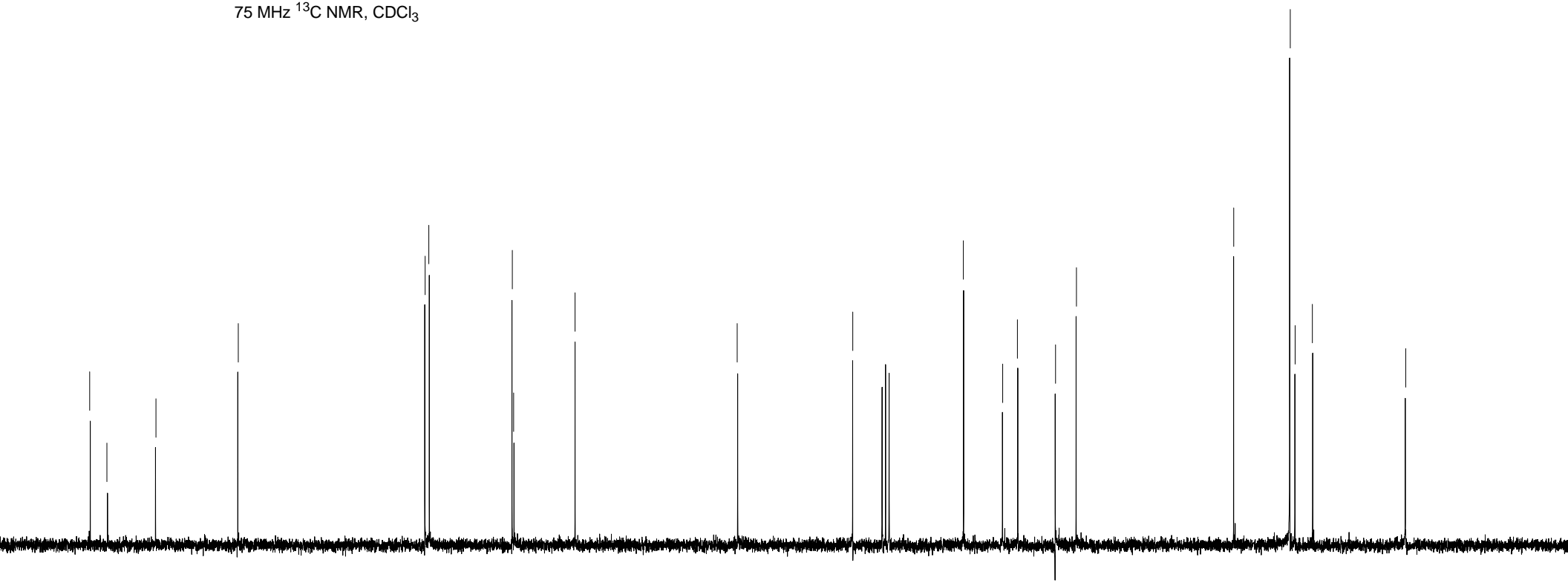
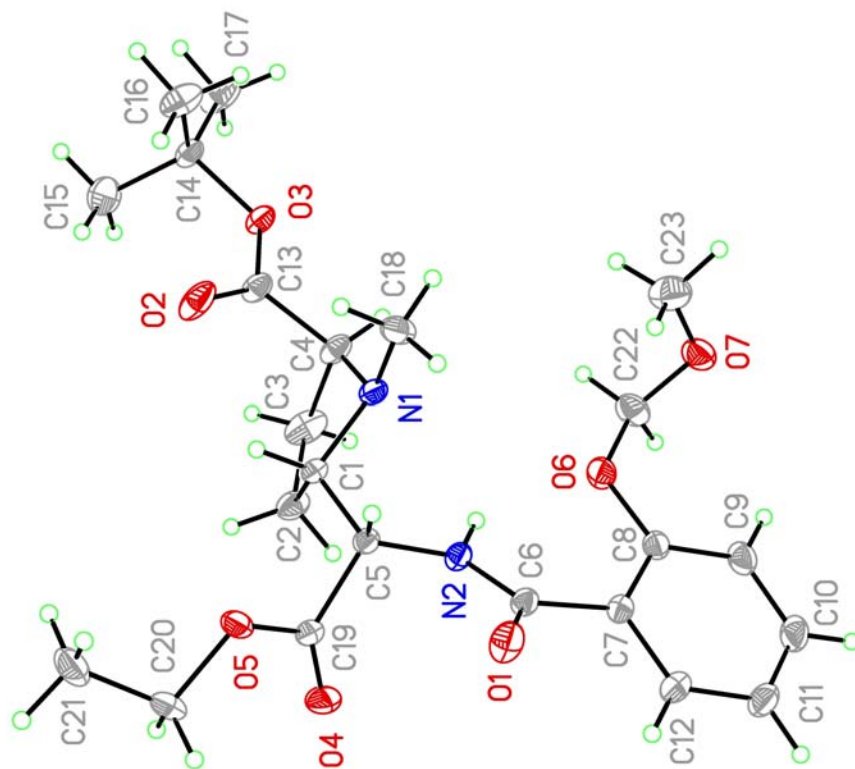


Table 1. Crystal data and structure refinement for **16**.

Identification code	16	
Empirical formula	C ₂₃ H ₃₄ N ₂ O ₇	
Formula weight	450.52	
Temperature	228(2) K	
Wavelength	0.71073 Å	
Crystal system	Orthorhombic	
Space group	P2(1)2(1)2(1)	
Unit cell dimensions	a = 9.0823(6) Å	α = 90°.
	b = 9.2605(7) Å	β = 90°.
	c = 29.233(2) Å	γ = 90°.
Volume	2458.7(3) Å ³	
Z	4	
Density (calculated)	1.217 Mg/m ³	
Absorption coefficient	0.090 mm ⁻¹	
F(000)	968	
Crystal size	0.46 x 0.41 x 0.37 mm ³	
Theta range for data collection	3.07 to 30.50°.	
Index ranges	-12 ≤ h ≤ 12, -13 ≤ k ≤ 13, -41 ≤ l ≤ 41	
Reflections collected	65942	
Independent reflections	4224 [R(int) = 0.0374]	
Completeness to theta = 30.50°	99.8 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.970 and 0.960	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	4224 / 0 / 307	
Goodness-of-fit on F ²	1.038	
Final R indices [I > 2σ(I)]	R1 = 0.0397, wR2 = 0.1024	
R indices (all data)	R1 = 0.0496, wR2 = 0.1106	
Largest diff. peak and hole	0.219 and -0.157 e.Å ⁻³	



Compound **16** single molecule view, all thermal ellipsoids at 20% probability