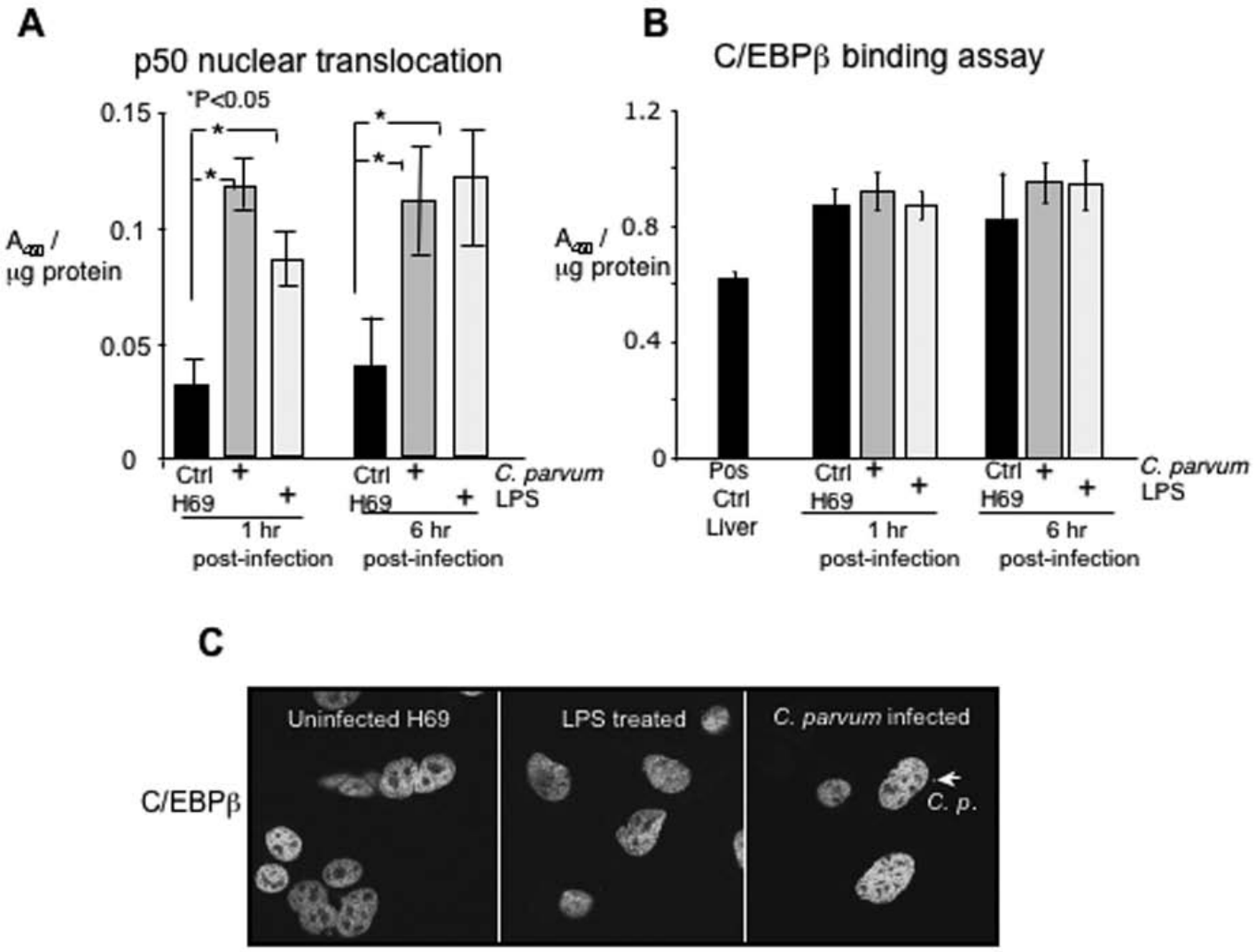
Supplemental Fig. 1. Cellular localization of the transcription factors p50 and C/EBPβ̃. **A.**̃ An ELISA based method, utilizing isolated nuclei, was used to demonstrate the nuclear translocation of the NF-κB p50 subunit following microbial stimulus. Both *C. parvum* infection and LPS treatment induced the nuclear translocation of p50 within one hour. The nuclear localization of p50 was also detected six hours after microbial stimulus. **B.** An ELISA-based C/EBPβ consensus sequence DNA binding assay was also performed on isolated nuclei from control uninfected and microbial-stimulated H69 cells. Isolated nuclei from total liver were used as the positive control. Cholangiocyte C/EBPβ could interact with the C/EBPβ consensus sequence in all conditions tested. **C.** Immunofluoresence was utilized to define the cellular localization of C/EBPβ in uninfected or microbial stimulated culture H69 cells. C/EBPβ was confined to the nucleus in both control, uninfected and in microbe-stimulated cholangiocytes. Data are represented as mean +/- SEM.

Supplemental Table 1. List of primers used for PCR-based approaches. Underlined sequences in the "Forward Primer" column correspond to inserted XhoI restriction endonuclease sites, while the underlined sequences in the "Reverse Primer" column correspond to inserted BglII restriction endonuclease sites used for subsequent cloning into the pGL4.22 Luciferase vector.



<Supplemental Figure 1>

Table 1. List of primers.

Method		
Let-7i Race PCR	**Forward Primer**	**Reverse Primer**
5' RACE		
Gene Specific Primer	5'-TAGCAAGGCAGTAGCTTGCGCAGTTA	Supplied by manufacturer
Nested	5'-ATCTCCACAGCGGGCAATGTCACAA	Supplied by manufacturer
3' RACE		
Gene Specific Primer	5'-CTGGCTGAGGTAGTAGTTTGTGCTGTT	Supplied by manufacturer
Nested	5'-TTGTGACATTGCCCGCTGTGGAGAT	Supplied by manufacturer
RT-PCR		
PPM1H	5'-TGAAGCCTTAAGCAGTGGAAA	5'-TGTGGAAGGAAAACCAGTGAC
MON2	5'-TGACTGGCCATCTACTTGAGAA	5'-GATCATTTCTGATTGCTCCCA
C12ORF61	5'-AGCTCTACAGGAGGCGAAAAC	5'-CCTCTTTCAGGGCTGATTTCT
Let-7i promoter ChIP	5'-CGTTTTCGCCTCCTGTAGAGC	5'-CGCCAAACAGCAGCCAAGGCA
Luciferase Constructs		
Full-length (-2461)	5'-CCGCTCGAGCGCAGTCAATGAGCAGTGAGGG	5'-GGAAGATCTAGCAAAGCGGCTCCGGCG
Δ1(-1991)	5'-CCGCTCGAGAATATGTCAGGCAGGCTGAT	5'-GGAAGATCTAGCAAAGCGGCTCCGGCG
Δ2(-1767)	5'-CCGCTCGAGTCTTACTTTGCCAGAAAGGGAAT	5'-GGAAGATCTAGCAAAGCGGCTCCGGCG
Δ3(-1167)	5'-CCGCTCGAGCTGGTTGGGAAAGGAGGTGCC	5'-GGAAGATCTAGCAAAGCGGCTCCGGCG
Δ4(-1110)	5'-CCGCTCGAGCTTAGGCGGCGATATGCG	5'-GGAAGATCTAGCAAAGCGGCTCCGGCG
Δ5(-961)	5'-CCGCTCGAGCTGATTTCTGGTAGCACGCTGA	5'-GGAAGATCTAGCAAAGCGGCTCCGGCG
Luciferase Mutants		
Δ1M-1**		
Distal	5'-CCGCTCGAGAATATGTCAGGCAGGCTGAT	5'CCACCCGTTCCCCAGCACCCTTCAGCTCTACAGGAGGCGA
Proximal	5'TCGCCTCCTGTAGAGCTGAAGGGTGCTGGGGAACGGGTGG	5'-GGAAGATCTAGCAAAGCGGCTCCGGCG
Δ1M-2**		
Distal	5'-CCGCTCGAGAATATGTCAGGCAGGCTGAT	5'-CAGCAGCCAAGGCATAGCAGGCCACCCGTTCCCCAGCACC
Proximal	5'-GGTGCTGGGGAACGGGTGGCCTGCTATGCCTTGGCTGCTG	5'-GGAAGATCTAGCAAAGCGGCTCCGGCG

** Distal and proximal amplicons were used as promoter and template to generate final mutant promoters