Reciprocal negative regulation of 3-phosphoinositide-dependent protein kinase-1 (PDK1) and apoptosis signal-regulating kinase 1 (ASK1) signaling by direct interaction and phosphorylation

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Generation of Stable Cell Lines for Overexpression and Knockdown of Endogenous PDK1 To generate HEK293 cells [PDK1 (OE)] stably expressing PDK1, we performed a PCR reaction using the full-length PDK1 cDNA as the template. The primers used for PCR were as follows: sense (5'-GCGAATTCATGGCCAGGACCACCAGCCAG-3', EcoRI site underlined) and antisense (5'-GCCAGCTGTCACTGCACAGCGGCGTCCGG-3', Sall site underlined). The amplified PCR products were cut with EcoRI plus SalI and cloned into pcDNA3.1-HisC vector, cut with EcoRI and XhoI, to generate pcDNA-His-PDK1. HEK293 cells were then transfected with pcDNA-His-PDK1. To prepare HEK293 cells (PDK1 shRNA) stably expressing PDK1specific shRNA, a double-stranded oligonucleotide (forward, 5'-GATCCCCGAGACCT CGTGGAGAAACTTTCAAGAGAAGTTTCTCCACGAGGTCTCTTTTTGGAAA-3'; reverse, 5'-AGCTTTTCCAAAAAGAGACCTCGTGGAGAAACTTCTCTTGAAAGTTTCTCCACGA GGTCTCGGG-3'; PDK1 sequence underlined) was cloned into the pSuper vector. For inducible knockdown of endogenous PDK1 expression, a double-stranded oligonucleotide (forward, 5'-TCGAGGGAGACCTCGTGGAGAAACTTTCAAGAGAAGTTTCTCCACGAG GTCTCCTTTTTTA-3'; reverse, 5'-AGCTTAAAAAAGGAGACCTCGTGGAGAAACTTCT CTTGAAAGTTTCTCCACGAGGTCTCCC-3'; PDK1 sequence underlined) was cloned into the pSingle-tTS-shRNA vector as described previously (38).

# **Construction of PDK1 Mutants**

PDK1 mutants were generated by PCR mutagenesis, as described previously (17), on PDK1-C (amino acids 249 to 556) or wild-type PDK1 using the following primers: PDK1-C forward primer 5'-GC<u>GAATTC</u>GTTTCTCCAGAGCTGCTCACG-3' (*EcoR*I site underlined), reverse primer 5'-GC<u>CAGCTG</u>TCACTGCACAGCGGCGTCCGG-3' (*Sal*I site underlined); wild-type

PDK1 forward primer 5'-GC<u>GAATTC</u>ATGGCCAGGACCACCAGCCAG-3' (*EcoR*I site underlined), reverse primer 5'-GC<u>CAGCTG</u>TCACTGCACAGCGGCGTCCGG-3' (*Sal*I site underlined); for S258A/S262A, sense 5'-ACGGAGAAGGCCGCCTGTAAGGCTTCAG ACCTT-3', antisense 5'-AAGGTCTGAAGCCTTACAGGCGGCCTTCTCCGT-3'; for S394A /S398A, sense 5'-TCCTCCTCCGCACACTCCCTGGCAGCCTCCGAC-3', antisense 5'-GTCGGAGGCTGCCAGGGAGTGTGCGGAGGAGGAGA-3'; for T518A/T522A, sense 5'-TTT GTCCACGCGCCTAACAGGGCGTATTATCTG-3', antisense 5'-GTCGGAGGCTGCCAGG GAGTGTGCGGAGGAGA-3'.

## Small Interfering RNA (siRNA)

The PDK1 siRNA (#1) oligonucleotide (5'-GGACUCGAACUCCUUUGAATT-3') targeting the coding region (amino acids 420 to 425) on human PDK1 (GenBank<sup>TM</sup> accession number Y15056) and the PDK1 siRNA (#2) oligonucleotide (5'-GAGACCUCGUGGAGAAACU-3') corresponding to the coding region (amino acids 310 to 316) were used for the siRNA experiments.

### **Immuno-depletion of Akt**

HEK293 cell extracts expressing Myc-PDK1 were incubated with approximately 4  $\mu$ g of anti-Akt antibody for 1 h at 4 <sup>0</sup>C, and protein A-Sepharoses were subsequently added and incubated for an additional 3 h at 4 <sup>0</sup>C, as described previously (38). After immuno-depletion twice with an anti-Akt antibody, the immuno-depletion of Akt was confirmed by immunoblot analysis using an anti-Akt antibody.



FIGURE 1. **PDK1 phosphorylates ASK1** *in vitro*. GST-tagged wild-type PDK1, precipitated using glutathione-Sepharose beads from HEK293 cells transiently expressing GST-PDK1, was assayed for its kinase activity in the presence of kinase buffer containing approximately 3 to 4 µg of recombinant GST-tagged ASK1(K709R) or ASK1(T838A) substrates.



B



A

FIGURE 2. **PDK1 directly phosphorylates ASK1 at Ser<sup>967</sup>.** *A*, GST-tagged PDK1(WT and KD), precipitated using glutathione-Sepharose beads from HEK293 cells expressing GST-PDK1(WT and KD) treated or untreated with specific inhibitors for PKA (H-89; 10  $\mu$ M, 30 min) and PKC (staurosporine; 100 nM, 30 min), was analyzed for its kinase activity by an *in vitro* kinase assay using approximately 3 to 4  $\mu$ g of recombinant GST-tagged ASK1(K709R) as a substrate. *B*, Myc-PDK1, immunoprecipitated using an anti-Myc antibody from control cell extracts (-) or Akt-depleted extracts (*Akt depletion*) of HEK293 cells expressing Myc-PDK1, was assayed for its kinase activity in the presence of kinase buffer containing recombinant GST-tagged ASK1(K709R) and one of its substitution mutants, ASK1(S83A), ASK1(T838A), ASK1(S967A), or ASK1(S1034A), as substrates (each 4  $\mu$ g). The level of immuno-depleted Akt in total cell lysates was determined by immunoblot analysis using an anti-Akt antibody (*5th panel*).



FIGURE 3. Determination of the expression levels of ASK1 and its downstream targets used in immunoblot analyses. The expression levels of ASK1, MKK3, p38, and ATF2 in total cell lysates were evaluated by immunoblot analyses using the indicated antibodies (see Fig. 4*B* and *C*, *lower right*). The overexpression or knockdown level of endogenous PDK1 was determined by anti-PDK1 immunoblotting.

Α



FIGURE 4. **Effect of PDK1 on caspase-3 activity and PARP cleavage.** HEK293 cells were transiently transfected with increasing amounts of PDK1(WT and KD) (2 and 4  $\mu$ g), wild-type ASK1 (2 and 4  $\mu$ g), or PDK1 and ASK1 siRNAs (100 and 200 nM) in the presence or absence of H<sub>2</sub>O<sub>2</sub>. Cell lysates were then analyzed by immunoblot analysis using an anti-caspase-3 antibody (Calbiochem) to determine the degradation of caspase-3 (*A*). PARP degradation was also measured by immunoblotting with an anti-PARP antibody (Santa Cruz Biotechnology) (*B*).



FIGURE 5. Effect of ASK1-mediated phosphorylation of PDK1 at Ser<sup>394</sup> and Ser<sup>398</sup> on PDK1-mediated apoptosis and cell cycle progression. Assays were done using 293T (*upper*) or HaCaT (*lower*) cells transfected with the indicated expression vectors, as described in Fig. 8A and B.



B

Α



9

FIGURE 6. **Regulation of the association between PDK1 or ASK1 and its regulators by H<sub>2</sub>O<sub>2</sub> and insulin.** *A*, Modulation of the association between PDK1 and its regulators, STRAP and 14-3-3 protein, by H<sub>2</sub>O<sub>2</sub> and insulin. HEK293 cells were transfected with expression vectors encoding FLAG-STRAP, FLAG-14-3-3 protein and GST-PDK1, as indicated. After 48 h of transfection, the cells were incubated with 100 nM insulin for 20 min. For H<sub>2</sub>O<sub>2</sub> treatment, the cells were incubated with 2 mM H<sub>2</sub>O<sub>2</sub> for 30 min. The cell lysates were subjected to precipitation with glutathione-Sepharose beads (*GST purification*), followed by immunoblotting with an anti-FLAG antibody to determine the association between PDK1 and STRAP or 14-3-3 protein. *B*, Modulation of the association between ASK1 and its regulators, including 14-3-3 protein, Trx, MKK3, and ASK1, by H<sub>2</sub>O<sub>2</sub> and insulin. HEK293 cells, transfected with the plasmids indicated, were treated with or without H<sub>2</sub>O<sub>2</sub> or insulin as described in *A*, and the cell lysates were immunoprecipitated with an anti-FLAG antibody (*α-FLAG*), followed by immunoblotting with an anti-HA antibody to determine the association between ASK1 and its regulators.