Supporting Information

A Short, Strong Hydrogen Bond in the Active Site of Human Carbonic Anhydrase II

Balendu Sankara Avvaru,^a Chae Un Kim^b, Katherine H. Sippel,^a Sol M. Gruner^{b,c}, Mavis Agbandje-McKenna^a, David N. Silverman^{a,d,*}, Robert McKenna^{a,*}

Department of Biochemistry and Molecular Biology^a, Department of Pharmacology and Therapeutics^d, College of Medicine, University of Florida, Gainesville, Florida 32610, USA and Cornell High Energy Synchrotron Source (CHESS)^b, Physics Department^c, Cornell University, Ithaca, NY 14853, USA.

*Corresponding authors: D.N.S. Department of Pharmacology, College of Medicine, University of Florida, Box 100267, Gainesville, Florida 32610; Phone: (352) 392 3556; Fax (352) 392 9696; e-mail: silvrmn@ ufl.edu; R.M. Department of Biochemistry and Molecular Biology, College of Medicine, University of Florida, Box 100245, Gainesville, Florida 32610; Phone: 352-392-5696; Fax: 352-392-3422; e-mail: rmckenna@ufl.edu.

Materials and methods

Expression and Purification of HCA II. The plasmid encoding HCA II was transformed into E.coli BL21 cells through standard procedures and the transformed cells were expressed at 37 °C in LB medium containing 100 μ g/ml ampicillin (*1*). HCA II production was induced by the addition of isopropyl thiogalactoside to a final concentration of 1mM at an O.D₆₀₀ of 0.6 AU. The cells were harvested 4 hrs after induction. The cell pellets were lysed and HCA II was purified through affinity chromatography using pAMBS resin as has been described elsewhere (*2*).

Crystallization and X-ray data collection of HCA II. Crystals of HCA II were obtained using the hanging drop vapor diffusion method (*3*). Ten µl drops of equal amounts of protein and precipitant were equilibrated against precipitant solution (1.3 M sodium citrate; 50mM Tris-HCl; pH 7.0) at room temperature (~20 °C) (*4*). A crystal was cryoprotected by quick immersion into 20% glycerol precipitant solution and flashcooled by exposing it to a gaseous stream of nitrogen at 100K. X-ray diffraction data were collected at the Cornell High Energy Synchrotron Source (CHESS) F1 Station with the wavelength of 0.9173 Å. Quantum 270 was used and the distance was 100 mm to allow 1 degree oscillation without spot overlapping. To cover high resolution spots, the detector was offset (moved upwards). Indexing, integration, and scaling were performed using HKL2000 (*5*).

Structure determination of HCA II. The crystal structure of HCA II (PDB accession code: 2ILI) (4) was used to obtain initial phases of the apo-structure using SHELX97 (6). The zinc and all solvent molecules were removed to avoid model bias. 5% of the unique reflections were selected randomly and excluded from the refinement

data set for the purpose of R_{free} calculations (7). Structural refinement proceeded using SHELXL initially with data from 50.0 to 2.0 Å resolution. The protein geometry was defined using the default constrains of conjugate-least squares (CGLS) mode in SHELXL. Each round of CGLS comprised of 15 cycles of refinement. 2Fo-Fc and Fo-Fc electron density Fourier difference maps were calculated after each successive round of CGLS and manually inspected by the graphics program COOT (8) for further fine-tuning of the model and the incorporation of solvent molecules. After some initial rounds of CGLS refinement, the resolution was extended to 0.9 Å and subsequently after several more cycles of refinement, the model was further subjected to several cycles of full anisotropic refinement and hydrogen riding which led a convergence of R_{crvst} and R_{free} to 12.5 and 13.1, respectively. The geometry of the final model was checked using the PROCHECK algorithm (9). The RMSD for bond lengths and bond angles were found to be within accepted limits of 0.004 Å and 1.0°, respectively. It was observed that 89 % of the dihedral angles were in the most favored region while the rest were in the allowed region with the exception of 0.5 % which were in the generously allowed region. The geometry and statistics of the final model are summarized in Table S1.

Unit cell dimensions,	
<i>a</i> , <i>b</i> , <i>c</i> (Å), β (°)	42.2,41.3,72.2,104.2
Resolution (Å)	50 - 0.9 (0.92-0.9)
Number of unique	164840 (6751)
Reflections	
Completeness (%)	92.3(76.1)
Redundancy	6.1 (2.8)
R _{symm} ^a	7.8 (58.0)
Ι/σ(Ι)	25.0 (2.0)
$R_{\rm cryst}^{\rm b}/R_{\rm free}^{\rm c}$	12.5 / 13.1
Ramachandran statistics (%)	
Most favored, additionally allowed and	89.0, 10.5, 0.5
generously allowed regions	
Number of protein/solvent atoms	2327/487
Average B factors(Å ²)	
main/side chain	10.9/15.2
Zn/solvent	6.5/26.0

Table S1. Refinement and model statistics for HCA II

Values in parentheses refer to the highest resolution bin. ^{*a*} $R_{symm} = \Sigma |I - \langle I \rangle / \Sigma \langle I \rangle x$

100. ${}^{b}R_{cryst} = \Sigma |Fo| - |Fc| / \Sigma |F_{obs}| \times 100$. ${}^{c}R_{free}$ is calculated in same manner as R_{cryst} ,

except that it uses 5% of the reflection data omitted from refinement.



Figure S1. Two dimensional representation of Figure 1. Bond lengths of the oxygen atoms in their respective hydrogen bonding partners are given. Note: The figure is not to scale. Figure created using Ligplot.

Figure S2. Two dimensional representation of Figure 1. Bond angles of the oxygen atoms to their respective hydrogen bonding partners are given. Note: The figure is not to scale. Figure created using Ligplot.

References

- Forsman, C. A., Behravan, G., Osterman, A., and Jonsson, B. H. (1988) Production of active human carbonic anhydrase II in E. coli, *Acta. Chem. Scand.* 42, 314–318.
- Khalifah, R. G., Strader, D. J., Bryant, S. H., and Gibson, S. M. (1977) Carbon-13 nuclear magnetic resonance probe of activesite ionizations in human carbonic anhydrase, *Biochemistry* 16, 2241–2247.
- McPherson, A. (1982) Preparation and Analysis of Protein Crystals, 1st Ed., Wiley, New York.
- Fisher, S. Z., C. M. Maupin, M. Budayova-Spano, L. Govindasamy, C. K. Tu, M. Agbandje-McKenna, D. N. Silverman, G. A. Voth, R. McKenna. (2007) Atomic Crystal and Molecular Dynamics Simulation Structures of Human Carbonic Anhydrase II: Insights into the Proton Transfer Mechanism, *Biochemistry* 46, 2930-2937.
- Otwinowski, Z., and Minor, W. (1997) Processing of x-ray diffraction data collected in oscillation mode, *Methods Enzymol.* 276, 307–326.
- Sheldrick, G. M. (2008) A Short History of SHELX. Acta Crystallogr. Sect. A 64, 112–122.
- Brunger, A. T. (1992) Free R value: A novel statistical quantity for assessing the accuracy of crystal structures, *Nature* 355, 472–475.
- Emsley, P., and Cowtan, K. (2004) Coot: model-building tools for molecular graphics, *Acta Crystallogr. Sect. D* 60, 2126–2132
- 9. Laskowski, R. A., MacArthur, M. W., Moss, D. S., and Thornton, J. M. (1993) PROCHECK: a program to check the stereochemical quality of protein structures, *J. Appl. Cryst.* 26, 283–291.