Supplementary Material

No specific short haplotype was found in the vicinity of SNP rs1801552

Material and Methods: To address whether a shorter haplotype could be identified in the vicinity of rs1801552, we chose five SNPs (rs9940010, rs13330170, rs9935563, rs9925080, rs9925161) with MAFs between 0.008 and 0.438 that were genotyped in six probands displaying high AI. Primer sequences and amplicon sizes are listed in the Supplementary Table 6 (Haplotype Analysis 2).

Results and discussion: In order to address whether a shorter haplotype could be found in the vicinity of SNP rs1801552, we chose five SNPs (rs9940010, rs13330170, rs9935563, rs9925080, rs9925161) with MAF between 0.008 and 0.438 that were genotyped in 6 probands displaying high *CDH1* AI. We registered no differences between our series and the HapMap control's haplotypes, and therefore we did not proceed with further analysis.

SNPs variation at miRNA target sites in the CDH1 3'UTR do not explain AI

Material and Methods: To assess whether genetic variation within or in the vicinity of microRNAs binding sites, at the 3'UTR of the *CDH1* gene, could modulate the binding of specific microRNAs, we genotyped a polymorphism rich region encompassing six SNPs overlapping several miRNA binding sites predicted by the following bioinformatic tools: PicTar (http://pictar.mdc-berlin.de/), DIANA-MicroT (http://diana.cslab.ece.ntua.gr/), MicroInspector (http://mirna.imbb.forth.gr/microinspector/) and TargetScan (http://www.targetscan.org/ and the Sanger miRNA database http://microrna.sanger.ac.uk/). Searching the Patrocles database (http://www.patrocles.org/) we were able to list two putative microRNA-target interactions near

polymorphic sites in the 3'-UTR of *CDH1* (rs33956791 and rs34182664). We genotyped these SNPs in our control and proband's cohorts, but these two SNPs were not represented in these individuals. Primers used and amplicon size are listed in Supplementary Table 7.

Results and discussion: Although rs1801552_C allele was apparently not in phase with SNPs downstream from the CDH1 3'UTR region, we still tested whether genetic variation occurring at miRNA binding sites could exist resulting in modulation of CDH1 ASE. To accomplish this task, we have applied a number of algorithms and databases to identify putative microRNA binding sites at the 3'UTR of the gene (See Material and Methods), at the same time that we spotted, using HapMap, all SNPs that localized at selected microRNA binding sites. Searching the Patrocles database, we were able to list two putative microRNA-target interactions near polymorphic sites at the CDH1 3'UTR (rs33956791 and rs34182664), nevertheless genotyping of these SNPs, both in control and probands' cohorts, demonstrated high degree of homozygosity in all cases excluding the possibility of an allele specific effect. One possible explanation for high *CDH1* AI arises from recent studies showing that the affinity of microRNAs (miRNAs) for a target mRNA sequence is potentially modulated by the presence of a SNP, as some SNPs result in alteration of the minimum free energy hybridization between the miRNA and mRNA. Nevertheless and although SNPs at predicted miRNAs binding sites and adjacent regions were identified, at the CDH1 3'UTR sequence, the high degree of homozygosity, both in controls and probands, hampered this possibility.

Supplementary Tables

SNP	DNA primers	Primer sequence	PCR product size
ng1901552	F	TGGCCTTAGAGGTGGGTGAC	262 bp
r\$1801552	R	CCAGGAAATAAACCTCCTCCAT	
rs33964119	F	CTCTCAACACTTGCTCTGTC	206 bp
	R	AGAGATCACCACTGAGCTAC	
rs1801026	F	AGACCCATGTGCTGGGAAAT	280 hp
	R	CTGGGTGAACCTTCTGATGC	289 bp

Supplementary table 1: Genotyping of Cancer-Free Individuals and Patients.

Supplementary table 2: ASE Analysis.

SNP	Primers	Primer sequence	PCR product size
	F	GGATAACCAGAATAAAGACCA	178 bp
RNA	R	CAGCAAGAGCAGCAGAATCA	
rs1801552	SBE1 (F)	TGACTGTGAAGGGGGCCGC	NA
	SBE2 (R)	CTGTGCCTTCCTACAGACGCC	NA
	F	TGCTGCTCTTGCTGTTTCTTC	144 bp
RNA rs33964119	R	GCAGCTGGCTCAAGTCAAAG	
	SBE	GAGGATGACACCCGGGACAA	NA
	F	CTACTTGAACGAATGGGGCAAT	174 bp
RNA rs1801026	R	TTCCCCAGAAACTCATCTCAAG	
	SBE	GTGCTGGGAAATGCAGAAATCA	NA
gDNA rs1801552	F	TGGCCTTAGAGGTGGGTGAC	262 bp
	R	CCAGGAAATAAACCTCCTCCAT	
	SBE	CTGTGCCTTCCTACAGACGCC	NA

Note: NA, Not applicable.

Supplementary (table 3: CDH1	Germline Promoter	Methylation Analysis.
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<i>CDH1</i> primers	CpG	flanking	Primer sequence	PCR product size
F			GGTAGGTGAATTTTTAGTTAATTAG	226 hr
R			ACTCCAAAAACCCATAACTAACC	220 Op

Supplementary table 4: Arms PCR.

CDH1 ARMS primers	Primer sequence	PCR product size
F	AGATTTTAGTAATTTTAGGTTAGAGGGTT A	339 bp
R	ААААСТАСААСТССАААААСССАТА	
ARMS_A	AGTAATTTTAGGTTAGAGGGTTA A	332 bp
ARMS_C	AGTAATTTTAGGTTAGAGGGTTAT	332 bp

Supplementary table 5: Haplotype Analysis 1.

SNP	Primer	Primer sequence	PCR product size
	F	AATCGACTCCTTGCCCTCAG	100 hr
rs11641611	R	CAGCACTGGGGGAATTTTTGC	- 100 bp
	SBE	CAGTGTACACACGTACACAAA	NA
	F	GGGTCGGGTTTCAGAGGAT	75 hr
rs2862778	R	ACGGTTCCTTCCCCCTAAAA	- 75 Op
	SBE	CACTGAAATTTTTGGGAA	NA
	F	GGAGTTGGAACTGGGAGGAA	114 hp
rs2296409	R	GCAACCACCCCATTGTAGGT	114 op
	SBE	GTGTAGATGGCATCATCCTCATC	NA
	F	AAGGGTGGAGGGATTCAACC	120 hp
rs4783665	R	TCCAAGAATGTCACCATCCCTA	129 op
	SBE	CTTTATCTCCAACTTTCACGC	NA
rs17715450	F	GGAGGTGGTTCTCCGCAAT	140 bp
	R	ATAAAGGGTGGCCCTCGACT	140 UP

	SBE	GACACCCATGTACCGTCCT	NA
rs16260	F	AACCGTGCAGGTCCCATAAC	252 bp
	R	CAAGCTCACAGGTGCTTTGC	
	SBE	GGCCTCGCGTAGACGCG	NA
	F	AGGCATCCTCATGCCACTCT	215 bp
rs12930371	R	AACAGACCCACCCTCAAGGA	
	SBE	AGCTCACATCTATACCCTTGTCA	NA
	F	CCACTTTGGATTCGGTTGGA	222 hp
rs1125557	R	TTTCTTTGGAGAGCAACATGC	232 вр
	SBE	CTTCCCCTATAATAAAGCATAGCTG	NA
	F	TCCTGACCTCAAGCAGTCCA	188 bp
rs10431923	R	TGAACACTGTGCTGGCAAAA	
	SBE	AAGGGAGTCATGCCTGAAAA	NA
rs2276330	F	TCCTCCCCTGGTCTCATCAT	171 hr
	R	CACAGTCACACACGCTGACC	1/1 op
	SBE	CTTGGGCTGGAGAAAGCA	NA
rs1801552	F	TGGCCTTAGAGGTGGGTGAC	262 hr
	R	CCAGGAAATAAACCTCCTCCAT	202 op
	SBE	CTGTGCCTTCCTACAGACGCC	NA
rs7203904	F	GCCACAGAGTGAGGCACAAT	200 bp
	R	CCCCACGGTACATGAGAAAGA	200 0p
	SBE	TGAGCAATAGAGCAACCAATC	NA

Note: NA,

Not

applicable.

Supplementary table 6: Haplotype Analysis 2.

SNP	RNA primers	Primer sequence	PCR product size
rs9940010,	F	TGGGATTACAGGTGTGAGC	434 bp
rs13330170	R	TCAACAAGAACACTGAATTGGC	454 Op
rs9935563, rs9925080, rs9925161	F	TTTGGGCTAACCTAGGAGCC	2011
	R	CACGTGACACTCACATCCCT	384 бр

<i>CDH1</i> MiRNA binding region flanking sites	Primer sequence	PCR product size
F	TTTGTGTGCTTCTGCTCATT	303 hp
R	TACTCTGGGAGGCCAAGATG	505 bp

Supplementary table 7: SNPs in MiRNA Binding Sites at CDH1 3'-UTR.

Supplementary Figure Legends

Supplementary Figure 1. Linkage Disequilibrium (LD) plot of HapMap phase II CEU data centered on *CDH1* and *CDH3* region of chromosome 16. The image was built using Haploview 4.1 software,^{*} and encompasses the region between positions 67.227.572 and 67.420.442 that includes the 12 SNPs chosen for the Haplotype Analysis. The triangular units designate haplotype blocks. The degree of LD between pairs of markers is indicated by the |D'| statistic (|D'| = 1 bright red; |D'| > 1 shades of pink/red). The positions corresponding to the analyzed SNPs are: rs11641611-1; rs2862778-10, rs2296409-34; rs4783665-38; rs17715450-51; rs16260-71; rs12930371-96; rs1125557-106; rs10431923-139; rs2276330-153; rs1801552-154; rs7203904-163 and are underlined in the figure.

Supplementary Figure 2. Graphical overview of linkage disequilibrium regarding the 12 SNPs analyzed using the GOLD application²⁴. **A**, haplotypes from the HapMap database. **B**, haplotypes from *CDH1* high AI probands; **C**, haplotypes from germline mutant and methylated probands; **D**, haplotypes from *CDH1* negative probands. Color code indicates values of the linkage

disequilibrium for |D'| statistic between pairs of two SNPs from dark blue (0-0.10) to bright red (0.90-1.00).

Supplementary Reference

*Barrett JC, Fry B, Maller J, et al. Haploview: analysis and visualization of LD and haplotype maps. Bioinformatics 2005;21:263-265.













