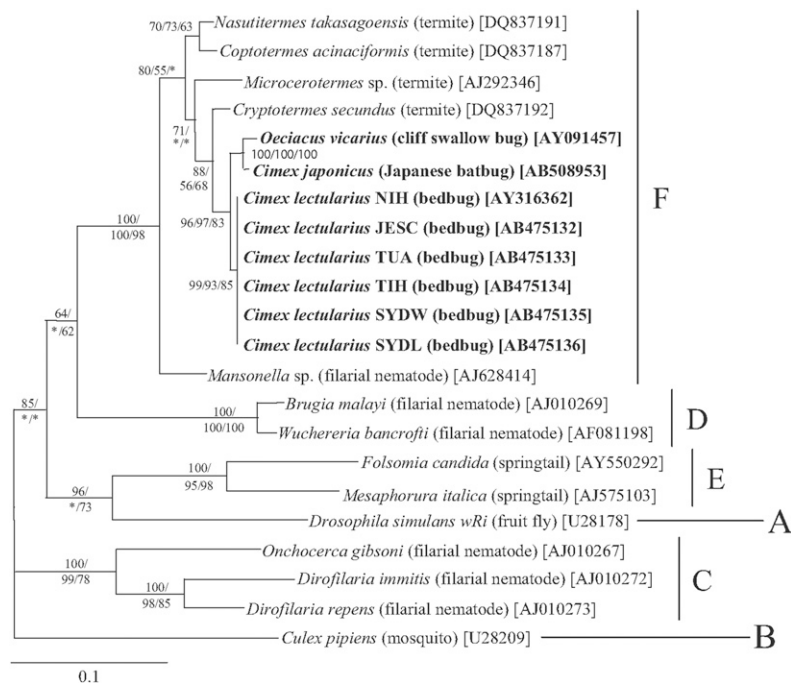
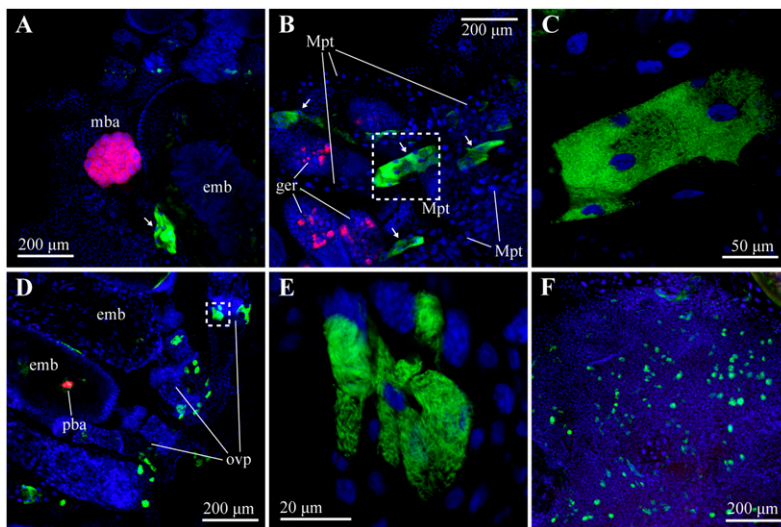


# Supporting Information

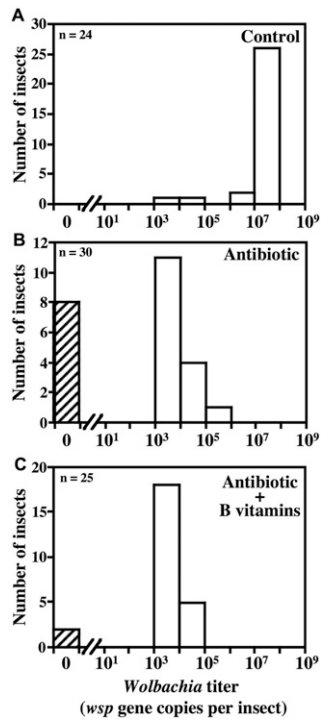
Hosokawa et al. 10.1073/pnas.0911476107



**Fig. S1.** Phylogenetic placement of *Wolbachia* from *C. lectularius* and allied bugs on the basis of 722 aligned nucleotide sites of *ftsZ* gene sequences. A Bayesian (BA) phylogeny is shown. Host insect names in *italics*, accession numbers in brackets, and *Wolbachia* supergroups A–F on the right side are shown. Posterior probabilities for BA analysis and bootstrap probabilities for maximum parsimony (MP) and maximum likelihood (ML) analyses greater than 50% are indicated at the nodes in the order of BA/MP/ML.



**Fig. S2.** Localization of  $\gamma$ -proteobacterial symbiont in *C. lectularius*. (A) A Malpighian tubule infected with  $\gamma$ -proteobacterial symbiont (arrow) nearby a maternal bacteriome and an embryo. (B) Infected regions (arrows) and uninfected regions (Mpt) of Malpighian tubules nearby ovarioles. (C) An enlarged image of the infected Malpighian tubule. (D) A number of infected cells in ovariole pedicels. (E) An enlarged image of infected cells in the ovariole pedicel. (F) Infected cells scattered in mesospermalage, a hemocyte-containing female organ into which sperm are introduced via traumatic insemination. Green, red, and blue signals indicate  $\gamma$ -proteobacteria, *Wolbachia* and insect nuclei, respectively. Abbreviations: emb, embryo; ger, germarium; mba, maternal bacteriome; Mpt, Malpighian tubule; pba, embryonic primordial bacteriome; ovp, ovariole pedicel.



**Fig. 53.** *Wolbachia* infection titers in adult insects of *C. lectularius* after 8 weeks of experimental treatments. (A) Control insects. (B) Antibiotic-treated insects fed on a rifampicin-containing blood meal. (C) Antibiotic-treated and B vitamin-supplemented insects fed on a rifampicin- and B vitamin-containing blood meal.

**Table S1. The insects used in this study and their infection with the *Wolbachia* symbiont and the  $\gamma$ -proteobacterial symbiont**

	Insect origin and description	<i>Wolbachia</i> infection rate*	$\gamma$ -proteobacterial infection rate*	<i>Wolbachia</i> 16S rRNA gene	<i>Wolbachia</i> ftsZ gene	$\gamma$ -proteobacterial 16S rRNA gene	Insect elongation factor 1 $\alpha$ gene
The common bedbug <i>Cimex lectularius</i>	JESC: A laboratory strain of unknown origin maintained on mice at the Japan Environmental Sanitation Center for over 10 years. Provided by A. Muto. <sup>†</sup>	100% <sup>†</sup> (46/46)	0% <sup>†</sup> (0/46)	AB475122	AB475132	—	AB475141
	TUA: Insects collected in 2005 and 2006 from a colony at a quail coop in the Tokyo University of Agriculture, Japan. Provided by T. Ishikawa. <sup>‡</sup>	100% <sup>†</sup> (48/48)	96% <sup>†</sup> (46/48)	AB475123	AB475133	AB475137	AB475142
	TIH: A laboratory strain maintained on mice at the Toyama Institute of Health for 7 years since collected at Toyama, Japan. Provided by T. Yamauchi. <sup>‡</sup>	100% <sup>†</sup> (3/3)	67% <sup>‡</sup> (2/3)	AB475124	AB475134	AB475138	—
	SYDW: Insects collected in 2007 at Sydney, Australia. Provided by S.L. Doggett. <sup>§</sup>	100% <sup>†</sup> (4/4)	100% <sup>‡</sup> (4/4)	AB475125	AB475135	AB475139	—
	SYDL: A laboratory strain maintained on rats at the Institute of Clinical Pathology and Medical Research (ICPMR) for 4 years since collected at Sydney, Australia. Provided by S.L. Doggett. <sup>§</sup>	100% <sup>†</sup> (4/4)	100% <sup>‡</sup> (4/4)	AB475126	AB475136	AB475140	—
	Total	100% (105/105)	53% (56/105)				
The Japanese batbug <i>Cimex japonicus</i>	KTCH: Insects collected in 2008 from a colony of the bat <i>Vespertilio sinensis</i> at Kutchan, Hokkaido, Japan. Provided by D. Fukui. <sup>¶</sup>	100% <sup>§</sup> (5/5)	0% <sup>§</sup> (0/5)	AB508951	AB508953	—	—

\*Infection rates estimated by diagnostic PCR.

<sup>†</sup>First-instar nymphs were inspected.

<sup>‡</sup>Adult insects were inspected.

<sup>§</sup>Developmental stages were uncertain.

<sup>¶</sup>Affiliations: A. Muto, Japan Environmental Sanitation Center, Kawasaki, Japan; T. Ishikawa, Tokyo University of Agriculture, Setagaya, Japan; T. Yamauchi, Toyama Institute of Health, Imizu, Japan; S. L. Doggett, Institute of Clinical Pathology and Medical Research, Westmead, Australia; D. Fukui, Forestry and Forest Products Research Institute, Sapporo, Japan.

**Table S2. Summary of cloning and genotyping of 16S rRNA gene clones from adult insects of *C. lectularius***

Strain	Sex and ID	Tissue	Number of genotyped clones	16S rRNA genotypes*	
				<i>Wolbachia</i>	$\gamma$ -proteobacterium
TUA	Male 1	Bacteriome	12	11	1
TUA	Male 2	Bacteriome	15	15	0
TUA	Female 1	Bacteriome	15	15	0
TIH	Male 1	Bacteriome	5	5	0
TIH	Male 2	Bacteriome	6	6	0
TIH	Female 1	Bacteriome	8	8	0
JESC	Male 1	Bacteriome	16	16	0
JESC	Male 2	Bacteriome	11	11	0
JESC	Male 3	Bacteriome	14	14	0
JESC	Female 1	Bacteriome	10	10	0
JESC	Female 2	Bacteriome	10	10	0
SYDW	Male 1	Abdomen	8	3	5
SYDW	Male 2	Abdomen	7	3	4
SYDW	Female 1	Abdomen	8	3	5
SYDW	Female 2	Abdomen	7	1	6
SYDL	Male 1	Abdomen	8	3	5
SYDL	Male 2	Abdomen	8	0	8
SYDL	Female 1	Abdomen	6	0	6
SYDL	Female 2	Abdomen	6	3	3
		Total	180	137	43

\*Sequencing data and restriction fragment length polymorphism data were combined.

**Table S3. List of B vitamins supplemented to the blood meal for *Wolbachia*-eliminated insects**

Compounds*	Final concentration*
Thiamine	100 $\mu$ g/mL
Riboflavin	20 $\mu$ g/mL
Nicotinic acid	100 $\mu$ g/mL
Pantothenic acid	100 $\mu$ g/mL
Pyridoxine	100 $\mu$ g/mL
D-biotin	1 $\mu$ g/mL
Folic acid	30 $\mu$ g/mL
Cobalamin	1 $\mu$ g/mL
Choline chloride	185 $\mu$ g/mL
Meso-inositol	118 $\mu$ g/mL

\*Prescription according to a previous study on the kissing bug *Rhodnius prolixus* (1)

1. Lake P, Friend WG (1968) The use of artificial diets to determine some of the effects of *Nocardia rhodonii* on the development of *Rhodnius prolixus*. *J Insect Physiol* 14:543–562.