

Supporting Information

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SI Methods

Quantification of mBST-2 Expression. 43D and 17-5 cells were treated with different concentrations of mouse IFN- α for 24 h. Total cellular RNAs (1 μ g) were reverse-transcribed with the qScript cDNA synthesis kit (Quanta Biosciences). The cDNAs were subjected to real-time PCR with Power SYBR Green PCR Master Mix (Applied Biosystems) and primers for mBST-2 (forward primer 5'-tcaggagtcctggagaaga and reverse primer 5'-atggagctgccagagttcac). GAPDH cDNA with appropriate primers was also amplified for normalization. The results are expressed as relative mBST-2 RNA levels compared to those in the untreated cells.

Confocal Microscopy. Confocal microscopy was performed as described in ref. 1 with slight modifications. The 43D and 17-5 cells were plated on glass coverslips 1 day before fixation. The cells were incubated with ice-cold PBS or 0.5% Triton X-100 for 2 min at 4 °C with gentle shaking. Afterward the cells were immediately overlaid with 4% paraformaldehyde and incubated for 20 min at RT. After washing with PBS and blocking with 3% BSA, 1% Triton X-100 in PBS, antibodies were added. The cells were stained with fluorescent-conjugated secondary antibodies followed by mounting with Vectashield mounting medium (Vector Laboratories). The images were analyzed on a confocal laser scanning microscope LSM710 with the image analyzing software Zen 2008 (Carl Zeiss).

1. Beer C, Pedersen L, Wirth M (2005) Amphotropic murine leukaemia virus envelope protein is associated with cholesterol-rich microdomains. *Virology* 2:36.
2. Kuznetsov YG, et al. (2002) Atomic force microscopy investigation of fibroblasts infected with wild-type and mutant murine leukemia virus (MuLV). *Biophys J* 83: 3665–3674.
3. Low A, et al. (2007) Mutation in the glycosylated gag protein of murine leukemia virus results in reduced in vivo infectivity and a novel defect in viral budding or release. *J Virol* 81:3685–3692.

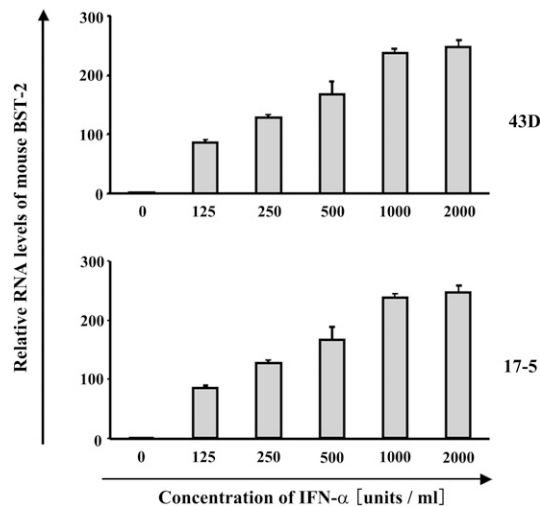


Fig. S1. IFN induces mouse BST-2 in both 43D and 17-5 cells. 43D and 17-5 cells were treated with different concentration of mouse IFN- α for 24 h. Total RNA extracted from the cells was subjected to real-time RT-PCR for quantification of mouse BST-2 and GAPDH. Expression levels of mouse BST-2 was normalized by GAPDH, and the results were expressed as relative RNA levels of mBST-2 to those in the untreated cells.

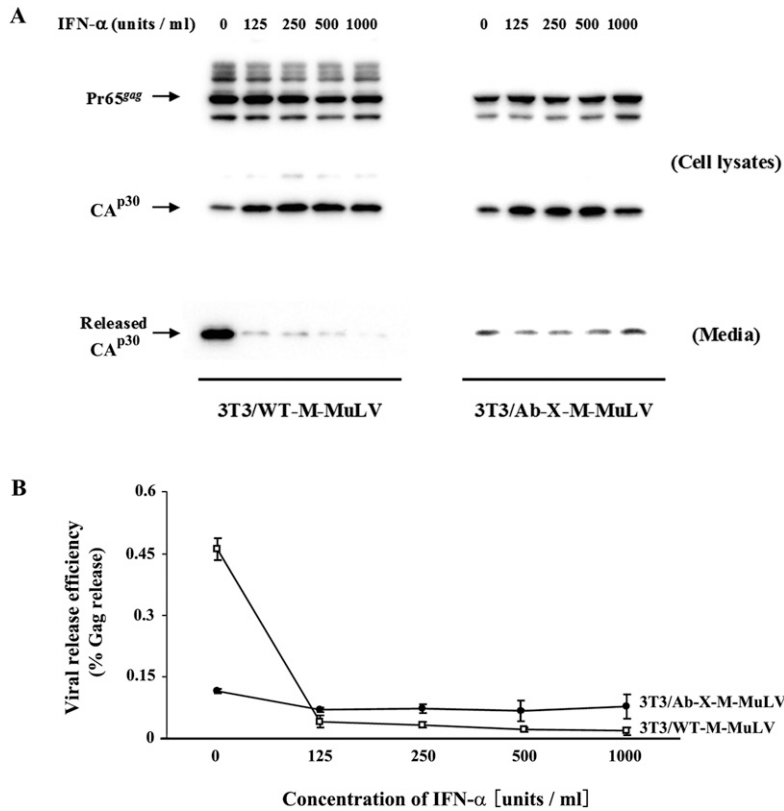


Fig. S2. Effect of IFN- α on virus release from the NIH 3T3 cells freshly infected with WT-M-MuLV and Ab-X-M-MuLV. (A) NIH 3T3 cells were infected with WT-M-MuLV (3T3/WT-M-MuLV) or Ab-X-M-MuLV (3T3/Ab-X-M-MuLV) isolated from each 43D and 17-5 cells. The cells were treated with different concentrations of mouse IFN for 24 h, after which media were replaced, and the cells and released viruses were collected 6 h after incubation. The same portions of cells and released viruses were subjected to the Western blots with anti-CA^{p30}. (B) Virus release efficiency from 3T3/WT-M-MuLV and 3T3/Ab-X-M-MuLV cells was calculated (means \pm SD from two replicate experiments).

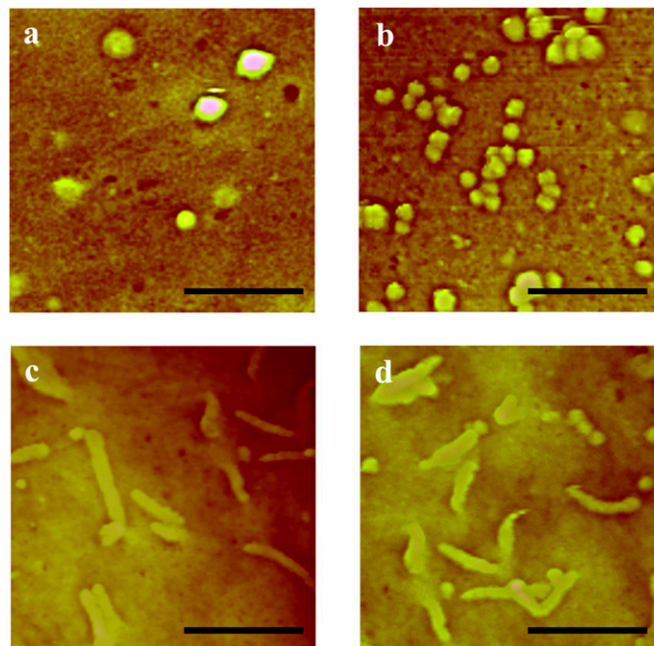


Fig. S3. Analyses of viral particles on the cell surface by atomic force microscopy (AFM). AFM procedures for virus-infected cells were described in ref. 2. Cells were grown on glass coverslips, fixed with 1% glutaraldehyde in PBS and postfixed with a 1% solution of osmium tetroxide in double-distilled H₂O. 43D cells (A and B) and 17-5 cells (C and D) with (B and D) or without (A and C) 500 U/mL IFN- α for 24 h. 43D cells show exclusively spherical virus particles on the cell surface (2, 3); some 17-5 cells show spherical particles, some show tube-like structures, and some show a combination of both (shown here). (Scale bars: 1 μ m.)

