Supplemental Data. Hashiguchi et al. (2009). Loss-of-function mutations of retromer large subunit genes suppress the phenotype of an *Arabidopsis zig* mutant that lacks Qb-SNARE VTI11.



Supplemental Figure 1. Morphological phenotype of *zip3-1 zig-1*.

We determined three morphological features to evaluate the suppression effect of zip3-1 on the zig-1 phenotype. (A) The angles between two adjacent internodes of the primary shoot in the second metamer. N=12. (B) The internodal length in the second metamer. 7-8 week-old plants were used for measurement. N=12. (C) Leaf morphology. The fifth and sixth leaves from 4-week-old plants were used for the measurement. N=16. For all three features, the zip3-1 zig-1 double mutant exhibited an intermediate phenotype between wild type and zig-1.



Supplemental Figure 2. Phenotypes of alleles of *zip3 zig-1* double mutants.

(A) Schematic structure of the *ZIP3* (*At2g17790*) gene and the position of mutations and T-DNA insertions. Boxes indicate exons and white boxes represent untranslated regions. Morphological phenotypes of 5-week-old plants of *zip3-1 zig-1* (B), *zip3-2 zig-1* (C), *zip3-3 zig-1* (D), and *zip3-4 zig-1* (E). Scale bars; 3 cm. (F) Gravitropic phenotypes. The gravitropic response of inflorescence stems of 5-week-old plants was gravi-stimulated by being placed horizontally at 23° C under dim non-directional light. 15 individuals of each genotype were examined. Bars represent SE.



Supplemental Figure 3. Immunoblot analyziz of ZIP3/VPS35A protein

Proteins were extracted from 2-week-old seedlings from each genotype and subjected to SDS-PAGE, followed by immunoblot analysis using anti-VPS35A antibody. An approximately 90-kDa protein band was detected only in extracts prepared from the wild type plant. Lower panel shows CBB staining to indicate that a similar amount of protein was applied to each lane of the gel.



Supplemental Figure 4. Phenotypes of alleles of *zip3* single mutants.

5-week-old plants of zip3-1 (A), zip3-2 (B), zip3-3 (C), and zip3-4 (D). Scale bars; 3 cm. (E) Gravitropic phenotypes. Gravitropic response of inflorescence stems of 5-week-old plants was gravi-stimulated by being placed horizontally at 23° C under dim non-directional light. 15 individuals of each genotype were examined. Bars represent SE.



Supplemental Figure 5. Morphological phenotypes of *mag1-1* and *mag1-1 zig-1* mutants.

5-week-old plants of mag1-1 (A), mag1-1 zig-1 (B). Scale bars; 3 cm.



Supplemental Figure 6. Phenotypes of *vps26* single mutants.

The 5-week-old plants of vps26a (A) and vps26b (B). Scale bars; 3 cm. (C) Gravitropic phenotypes. Gravitropic response of inflorescence stems of 5-week-old plants were gravistimulated by being placed horizontally at 23° C under dim non-directional light. 15 individuals of each genotype were examined. Bars represent SE.



Supplemental Figure 7. Phenotypes of *vps35b* and *vps35c* single mutants.

5-week-old plants of vps35b (A) and vps35c (B). Scale bars; 3 cm. (C) Gravitropic phenotypes. Gravitropic response of inflorescence stems of 5-week-old plants was gravistimulated by being placed horizontally at 23° C under dim non-directional light. 15 individuals of each genotype were examined. Bars represent SE.



Supplemental Figure 8. Localization of GFP-VTI12 and ARA6-mRFP in endodermal cells

Confocal images of GFP-VTI12 and ARA6-mRFP in endodermal cells. (A-C) wild type, (D-F) zig-1, (G-I) zip3-1 zig-1, (J-L) zip3-1. VTI12 and Ara6 were observed with GFP (A, D, G, H) and RFP fluorescence (B, E, H, K), respectively. zip3-1 zig-1/VTI12pro:GFP-VTI12 and WT/SCRpro:ARA6-mRFP lines were crossed, and four genotypes of wild type, zig-1, zip3-1 zig-1 and zip3-1 bearing both constructs homozygously were isolated. (M) Quantitative analysis of GFP-, RFP- and both signal- positive dots. The number of dots in images of endodermal cell as shown in A to L was counted. Values represent means \pm SE. Fifty cells of each genotype were examined.

Position	WT	zig-1	zip3-1 zig-1	zip3-1
Тор	0.06±0.03	3.2±0.35	0.22±0.07	0.04±0.03
Middle	0.18±0.08	2.0±0.51	0.36±0.12	0.24±0.09
Bottom	5.16±0.24	2.94±0.28	3.78±0.29	4.8±0.33

Supplemental Table 1. Localization of amyloplasts in the endodermal cell.

Localization of amyloplasts in the endodermal cell. An endodermal cell is divided into three areas (top, middle, bottom), and then the number of amyloplasts (mean Å} SD) located at each area was counted, based on the image of longitudinal sections shown in Figure 3 A -D. Fifty cells of each genotype were examined.

	WT	zig-1	zip3-1 zig-1	zip3-1
Öü	6.42±0.46	4.12±0.39	5.66±0.48	6.6±0.63
Ö†	3.04±0.34	0.16±0.07	1.78±0.23	4.44±0.49
Ö°	0.46±0.21	4.14±0.39	2.1±0.30	0.38±0.14
Ö¢	35	5	29	33
Ö£	0	13	6	0

Supplemental Table 2. Quantitative analysis of the cytological phenotype of living endodermal cells based on the observation as shown in Figure 3 H-K.

Quantitative analysis of the cytological phenotype of living endodermal cells based on the observation as shown in Figure 3 H-K. Time-lapse imaging for 5 min with 15 seconds-intervals were performed. Five characteristics of amyloplasts and vacuolar membrane were quantitatively observed in living endodermal cells (I to V). I; Average number of amyloplasts in an endodermal cell in a focal plane at a timepoint (time 0). II; Average number of amyloplasts that appeared to be enclosed by the vacuolar membrane within I. III; Average number of amyloplasts that hardly move during a 5 min-observation within I. IV; Number of cells in which transvacuolar strands arose during 5 min-observation, indicating vacuolar membrane dynamics. V; Number of cells in which abnormal clusters of vesicles were found during 5 min-observation. Fifty cells were observed for each genotype and then analyzed.

Supplemental Table 3.

Primer sets for cloning and RT-PCR

Primer name	Sequence		
gZIP3-F	5ÕCCCGGGAAATGTTAGATGCTTC-3Õ		
gZIP3-R	5ÕCCCGGGTTTACATTTCGAGCAGC-3Õ		
cZIP3-F	5ÕGTTCCCGGGATGATCGCAGACGGATCAGA-3Õ		
cZIP3-R	5ÕGTTCCCGGGTACTTTGATCGCCTGGTATC-3Õ		
cVPS35B-F	5ÕCTTGGATCCATGAGAACGCTCGCCGGAGTAG-3Õ		
cVPS35B-R	5ÕCTTGGATCCTCACAGCTTGATAGGGTCATA-3Õ		
cVPS35C-F	5ÕCTTGGATCCATGATCGCCGACGACGATGAG-3Õ		
cVPS35C-R	5ÕCTTGGATCCTCATTCAAACCATTCCATTTTG-3Õ		
cVPS35B-F2	5ÕAAGAGTTCTTCTCTGTTTAAGACG-3Õ		
cVPS35B-R2	5ÕTGTGTCTCGGATTTAAGTCACAGC-3Õ		
cVPS35C-F2	5ÕATTCTGCAAGACTCCTAAGGAAGC-3Õ		
cVPS35C-R2	5ÕTCTTCTTGCTGTCTCTGAAACTCG-3Õ		
cVPS26A-F1	5ÕGTCTCTCCCGATTGAAATAAACC-3Õ		
cVPS26A-R2	5ÕTGCATACAGAAAGTCTTGAAAAGG-3Õ		
cVPS26B-F1	5ÕATTACGCTCTTAACATCAGCTTGG-3Õ		
cVPS26B-R2	5ÕATCTGAGTACACAAAGTCTAAAGC-3Õ		
ACT8 Df	5ÕGAGAGATTCAGGTGCCCAG -3Õ		
ACT8 Dr	5ÕAGAGCGAGAGCGGGTTTTCA -3Õ		
pSCR(-30)D	5ÕTCTTACCTTATTTATAACCTAGGC-3Õ		
cVPS35B-R1	5ÕGATACATTCTGGGCAATATGTTGC-3Õ		
cZIP3-R2	5ÕATACAGTCTATTAGGTAATATTGG-3Õ		
cVPS35C-R1	5ÕATTCATGTGTGCATCACCATCTCC-3Õ		