

**Supplemental Figure 1.** pdTPI protein localizes to the chloroplast. **(A)** Alignment of *pdTPI* amino acid sequence with proteins from Arabidopsis (At cytoTPI), Petunia (gi602589), Solanum chacoense (gi38112661), *Coptis japonica* (gi340520), *Oryza sativa* (gi169820; gi115440976), *Triticum aestivum* (gi11124571), *Glycine max* (gi77540215), *Phaseolus vulgaris* (gi57283984), *Secale cereale* (gi407524), *Hordeum, vulgare* (gi1785947), *Pteris vittata* (gi84626306). **(B)** Subcellular localization of the pdTPI:GFP fusion protein in leaf cells. Transgenic Arabidopsis leaf protein expressing pdTPI-GFP was fractionated and detected by immunoblotting. Plastid PDH  $\alpha$ -subunit was used to determine plastid enrichment; antibody to the mitochondrial PDH  $\alpha$ -subunit was used to assay mitochondria enrichment. **(C)** Leaves from the transgenic pdTPI-GFP lines were observed by confocal microscopy. Transgenic lines transformed with vector only were used as a control.



**Supplemental Figure 2.** Expression of alternatively spliced *pdTPI* transcripts.

Tissues were harvested from *Col* wild-type. RT-PCR was applied to detect transcript abundance. Primers S21170F and S21170R were used to amplify 122 bp and 95 bp fragments from splice variant 1 and splice variant 2, respectively. *UBQ* was used as loading control.



Supplemental Figure 3. CytoTPI mutant identification.

(A) Schematic representation of T-DNA insertion positions in At3g55440 gene. LB, T-DNA left border.

(B) *cytoTPI* gene expression was not affected in Salk-106806 and Salk-003991 T-DNA insertion lines. (C) Salk-106806 and Salk-003991 appeared similar to wild type when grown for 7 weeks under long-day conditions.



Supplemental Figure 4. Characterization of *pdtpi* mutant.

(A) Schematic representation of T-DNA insertion positions in At2g21170 gene. LB, T-DNA left border. The primer locations and directions were indicated here.

**(B)** T-DNA insertion in Salk-152526 was verified by PCR by combining T-DNA primer and plant gene-specific primers.

(C) RT-PCR results indicated that T-DNA insertion in Salk-152526 line reduced gene expression. *pdTPI* gene expression in Salk-022963 and CS829061, however, was not affected. *UBQ* was used as a control.



Dark to light

**Supplemental Figure 5.** Metabolite augmentation does not rescue the *pdtpi* mutant phenotype. (A) Plants germinated on MS agar plates with the addition of 1 mM of sucrose, glucose, fructose-1, 6bisphosphate (F-1,6-P), dihydrohyacetone phosphate (DHAP), DL-glyceraldehyde 3-phosphate (GAP) and D(-)3-phosphoglyceric acid (PGA) for 5 d.

(B) Plants were first germinated on regular MS plates under darkness for 5 d, and then were transferred onto MS plates enriched with glycolytic intermediates and grown for an additional 2 d under continuous white light.



**Supplemental Figure 6.** Complementation of *pdtpi* mutant by transgenic expression of *pdTPI* cDNA. **(A)** PCR genotyping of individual transgenic T1 lines.

(B) Wild type and complementation lines grew under long-day condition for 20 days; the T2 generation of rescued lines 3, line 4 and line 6 are shown here.



**Supplemental Figure 7.** *pdtpi* mutant can not be rescued by the application of myo-inositiol. Wild type and *pdtpi* mutant were germinated on myo-inositol enriched MS plates for 5 d under continuous white light.



**Supplemental Figure 8.** Rubisco large subunit is down-regulated in the *pdtpi* mutant.

(A) Total protein from 5-d old seedlings was isolated and separated by SDS-PAGE. L: Rubisco large subunit, S: Rubisco small subunit. Anti-cytoTPI polyclonal antibody cross-reacts with pdTPI protein.

**(B)** Different amounts of purified, recombinant cytoTPI and ΔpdTPI protein were separated by SDS-PAGE, transformed to nitrocollulose membrane and probed with anti-outeTPI antibody.

transferred to nitrocellulose membrane and probed with anti-cytoTPI antibody.