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Supplementary Table: Sequences of primers used to amplify candidate regions in the blue tit.

Gene name	Forward primer 5'->3'	Reverse primer 5'->3'	T_m* (ºC) forward/ reverse	Expected length (bp)	PCR conditions[d]	
					$MgCl_2$ (endconcentration)	$T_{annealing}$(ºC)
AANAT	CRGCRCTGACCCTRCACA	GTGCTGCATCTCSRYGAAG	59/58	260[b]	2.0 mM	57
CKIδ	ATTGCTGCWGGMGARGAGGTT	TCCWCCCTGCATCATYTTGT	59/56	100[b]	2.0 mM	56
CKIε	GCAAGARGTGTCACGGAT	CTAAGCAAACACTGGTCC	54/53	460[b]	2.0 mM	49
CKIε (tau)	GCTGGTGTGGAGGGTTAAAT	TCCCAGGTGGGTGTTGAT	57/56	435[b]	1.5 mM	55
CKIε (dbt)	ATGATCTTCTCAGCAGGGGA	GAGAGTAGGCACAAATGCTTCC	57/60	250[b]	1.0 mM	55
PERIOD2	CTCTACTGTGTTGAAGKCATCTG	CTAATTCAGGTTGTGGYTTTTTG	59/56	170[b]	2.0 mM	55
ADCYAP1	GATGTGAGTAACCAGCCACT	ATAACACAGGAGCGGTGA	57/53	166[c]	1.5 mM	51
CLOCK[a]	TTTTCTCAAGGTCAGCAGCTTGT	CTGTAGGAACTGTTGYGGKTGCTG	58/64	285[c]		
CREB1	GGTCAGGCAGTTAAGATATTG	GTCTTACCAGTGGTTCCTTTAR	55/57	556[c]	2.0 mM	53
NPAS2	CTGTGGTAAATTTGATGATTCTGA	ACACCAAGTTCTTTGCACAATG	55/56	184[c]	2.0 mM	55

* approximate melting temperature
[a] primers according to Johnson et al. 2007
[b] approximate length
[c] contains maximum number of repeats obtained in the sample of 148 blue tit individuals
[d] PCRs were conducted in a final volume of 20 µl containing 1 µl of genomic DNA and 0.5 U Taq DNA polymerase (Fermentas), and a final concentration of 200 µM dNTPs, 0.5 µM of each of the forward and reverse primers, varying $MgCl_2$ concentrations (see table) and 1X Taq buffer with $(NH_4)_2SO_4$ (Fermentas). PCR cycling profiles for all markers began with an initial denaturation at 95ºC for 3 min and then proceeded with 30-35 cycles of 95ºC for 30 sec, annealing temperatures according to the table for 30 sec, and 72ºC for 1 min, followed by a final extension of 72ºC for 7 min.