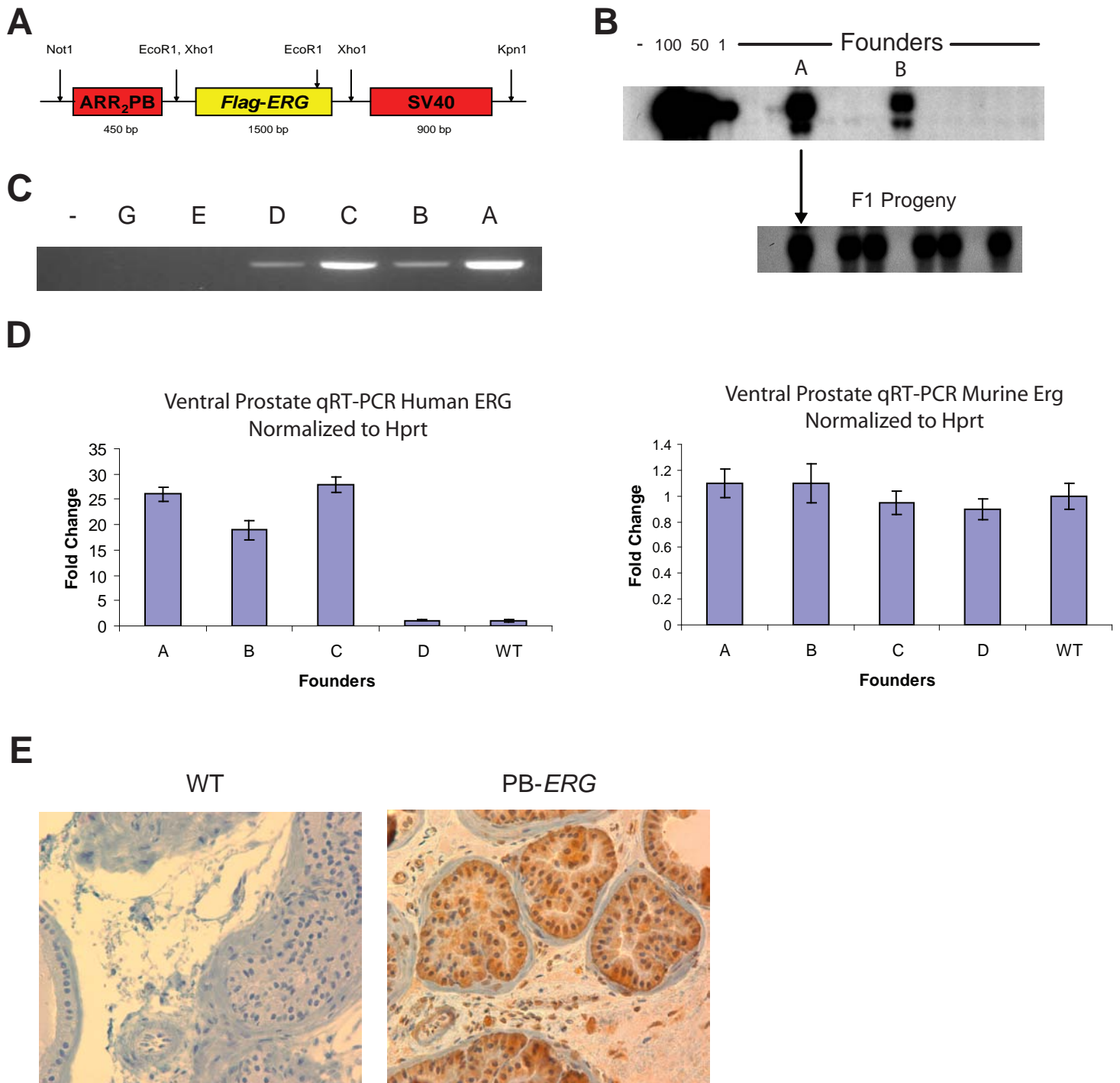


# **Aberrant ERG expression cooperates with loss of PTEN to promote cancer progression in the prostate**

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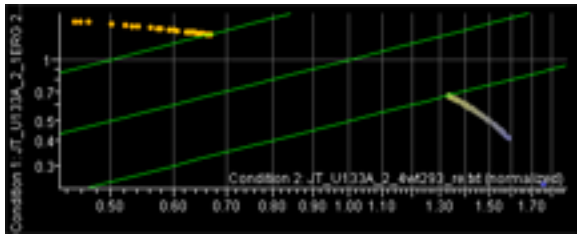
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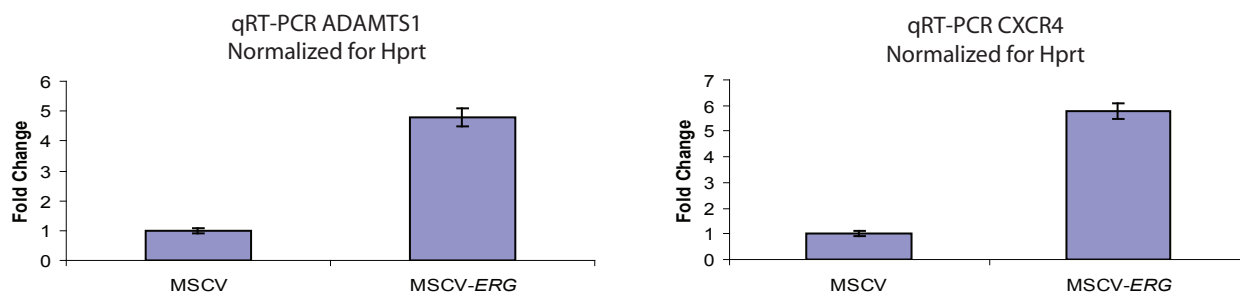
### Supplementary Figure 1. Generation of probasin-ERG mouse model

Our *Flag-ERG* construct was subcloned into the probasin-SV40 construct utilizing a *Xho1* multicloning site and linearized with *Not1* and *Kpn1* (A). Founders were identified using Southern blot analysis with *EcoR1* restriction enzyme digest, demonstrating a positive band corresponding to the *ERG* transgene at 1500 bp, and F1 offspring were analyzed for equivalent germ line transmission of the transgene (B). mRNA expression analysis was performed in triplicate using quantitative RT-PCR for both the human and murine *ERG* transcripts (D). The bar graph demonstrates the fold change in mRNA level following normalization to *Hprt* and our lowest value, and the mean and standard deviations from 3 experiments are shown. Immunohistochemistry was performed on representative sections of paraffin embedded prostate glands with the *Flag* polyclonal antibody, demonstrating protein expression of the *ERG* transgene (E).

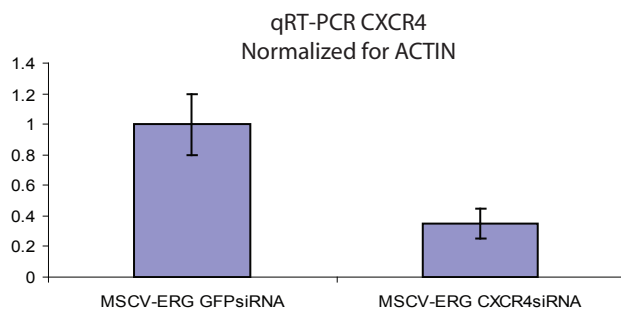
A



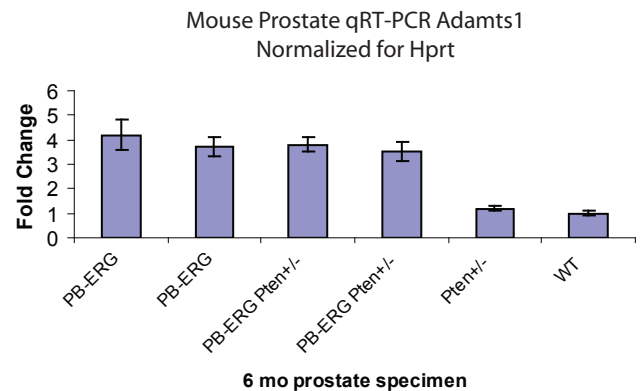
B



C



D



### Supplementary Figure 2. CXCR4 and ADAMTS1 regulation by ERG

Microarray expression analysis of ERG over-expression in 293HEK cells compared to vector control demonstrated differential expression of 167 genes (A). Candidate genes, *CXCR4* and *ADAMTS1*, were further evaluated and found to be up-regulated in primary MEF over-expressing ERG compared to controls, validating our findings across species (B). The bar graphs demonstrate the fold change in mRNA level following normalization to Hprt and our lowest sample value, and the mean and standard deviations from 3 experiments are shown. Knock-down of *CXCR4* with pooled siRNA against *CXCR4* and control (C). Further evaluation revealed that the mRNA expression of murine *Adamts1* was up-regulated in the prostate specimens of *ERG* transgenic mice compared to controls (D). The bar graphs demonstrate the fold change in mRNA level following normalization to Hprt and our lowest sample value, and the mean and standard deviations from 3 experiments are shown.

Supplementary Table 1. Primer sequences used for cloning, genotyping, RNA, and DNA analyses.

<b><u>Primer Name</u></b>	<b><u>Primer Sequence</u></b>
<b>Cloning</b>	
<i>ERG</i> forward	CTT GAT CGC ATT ATG GCC
<i>ERG</i> reverse	CCG CCA GGT CTT TAG TAG
<b>Genotyping</b>	
ARR2PB promoter forward	AGC AGG AAG CTA CTC TGC ACC
<i>ERG</i> 5' reverse	TTT GGC CAC ACT GCA TTC ATC
<b>qRT-PCR</b>	
human <i>ACTIN</i> forward	CAC GAG ACC ACC TTC AAC TC
human <i>ACTIN</i> reverse	CTT GAT CTT CAT GGT GCT GG
human <i>ERG</i> forward	TTA TCA GTT GTG AGT GAG GAC CA
human <i>ERG</i> reverse	AAG TCT GTC CAT AGT CGC TGG
human <i>CXCR4</i> forward	TAC ACC GAG GAA ATG GGC TCA
human <i>CXCR4</i> reverse	AGA TGA TGG AGT AGA TGG TGG G
human <i>ADAMTS1</i> forward	ACG AGG ACG AAG GGA CTG AG
human <i>ADAMTS1</i> forward	ATC GCT TCT TTC TTA TGC TTC CA
murine <i>Erg</i> forward	ACC TCA CCC CTC AGT CCA AA
murine <i>Erg</i> reverse	TGG TCG GTC CCA GGA TCT G
murine <i>Hprt</i> forward	CAC AGG ACT AGA ACA CCT GC
murine <i>Hprt</i> reverse	GCT GGT GAA AAG GAC CTC T
murine <i>Cxcr4</i> forward	GCT TCC GGG ATG AAA ACG TC
murine <i>Cxcr4</i> reverse	ACC AAT CCA TTG CCG ACT ATG
murine <i>Adamts1</i> forward	GAC CCG AGA GCC AGA ACA C
murine <i>Adamts1</i> reverse	CAC AAA TCG CTT CTT CCT TAT GC
<b>ChIP</b>	
<i>ADAMTS1</i> promoter forward	CTT TCC TCA TAT GTT CTC
<i>ADAMTS1</i> promoter reverse	AAC ATT GAC ATC CGC ACA CC
<i>CXCR4</i> promoter forward	GGC TGC GTC TGC TGA AAG C
<i>CXCR4</i> promoter reverse	GCT AGG AAC GCG TCT CTC