Supporting Information

Synthesis of phosphatase-stable, cell-permeable peptidomimetic pro-drugs that target the

SH2 domain of Stat3

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Experimental Methods

Fmoc-Haic was purchased from ChemImpex or was synthesized as described.¹ Fmoc-Gln-NHBn was prepared as described.² Other amino acids were from ChemImpex or NovaBiochem. Rink amide resin was from Advanced Chemtech. Solid phase peptide synthesis was carried out as described.^{2,3} Peptides and prodrugs were tested for purity by reverse phase HPLC on an Agilent 1100 HPLC using a Phenomenex Luna 5μ C18(2) 4.6 × 250 mm column. For both peptide intermediates and prodrugs the gradient was 10 – 80% MeCN/30 min each solvent containing 0.1% TFA. Peptides or prodrugs were purified by reverse phase HPLC on a Varian Dynamax HPLC using a Phenomenex Luna 10μ C18(2) 21.2 × 250 mm column. Gradients of MeCN in H₂O at 20 mL/min were employed. For peptides 0.1% TFA was included in the eluents. For prodrugs no TFA was used in the mobile phase. NMR spectra were obtained on either a Bruker DPX 300 MHz spectrometer or a Bruker DRX 500 MHz spectrometer.

MDA-MB-468 breast tumor cells were acquired from the American Type Culture Collection and were maintained as monolayers in DMEM medium supplemented with 10% fetal bovine serum, 100 mM L-glutamine, 100 μ M streptomycin and 100 U/mL penicillin. Cells were grown at 37°C in an atmosphere of 95% humidified air and 5% CO₂. Antibodies against Stat3 (Cat# 9132) and pStat3^{Y705} (Cat# 9145) were purchased from Cell Signaling Technology (Beverly, MA).

Synthesis of tert-butyl (E)-3-(4-iodophenyl)prop-2-enoate (2): A solution of tert-butyl diethylphosphonoacetate (10.0 g, 39.6 mmol), 4-iodobenzaldehyde (9.20g, 39.6 mmol) and cesium carbonate (19.3 g, 59.4 mmol) in dry THF (15 mL) was stirred for 4 h. The solvent was removed *in vacuo*, the residue dissolved in 300 mL EtOAc, and the organic phase was washed with water (2×30 mL) followed by brine (1×30 mL) and dried (MgSO₄). After filtration and

concentration, the crude product was purified by silica gel column chromatography eluting with 10% EtOAc-hexanes. A white solid was obtained (**2**, 11.1 g, 86%). ¹H NMR (CDCl₃, 300 MHz) δ 1.45 (s, 9H), 6.28 (d, 1H, J =16.0 Hz), 7.15 (d, 2H, J = 8.4 Hz), 7.41 (d, 1H, J=16.0 Hz), 7.62 (d, 1H, J = 8.4 Hz).

Synthesis of tert-butyl (E)-3-(4-diethoxyphosphoryldifluoromethyphenyl)prop-2-enoate

(3): To a solution of diethyl bromodifluoromethylphosphonate (6.45 g, 24.1mmol) in dry DMF (100 mL) was added cadmium powder (5.41 g, 48.2 mmol). The suspension was stirred for 12 h under argon. Excess cadmium was removed by filtration under argon and the filtrate was treated with CuCl (2.86 g, 28.9 mmol) and **2** (5.00 g, 15.1 mmol) for 24 h at rt. Et₂O (300 mL), was added and the mixture stirred for 5 min and filtered. The organic layer was washed with saturated NH₄Cl (2×40 mL) and water (4×40 mL), dried (MgSO₄) and evaporated to give an oily residue. The crude product was purified by silica gel column chromatography with 40% EtOAc-hexanes as the eluant to give 4.92 g (83%) of **3** as colorless oil. HRMS (M + 1) calcd 391.1486, found 391.1441; ¹H NMR (CDCl₃, 300 MHz) δ 1.32 (t, 6H, J = 7.2 Hz), 1.54 (s, 9H), 4.11-4.27 (m, 4H), 6.43 (d, 1H, J = 16.0 Hz), 7.56-7.64 (m, 5H). ¹⁹F NMR (CDCl₃, 282.0 MHz) δ -108.8 (d, J = 115.0 Hz, 2F). ¹³C NMR (CDCl₃, 75 MHz) δ 16.2, 16.3, 28.1, 64.8, 64.9, 80.8, 122.2, 126.8, 127.8, 137.1, 142.1, 165.8.

Synthesis of pentachlorophenyl (E)-3-(4-diethoxyphosphoryldifluoromethyphenyl)prop-2-enoate (4): A solution of 7 (4.00 g, 10.2 mmol) in 5 mL dry dichloromethane was treated with 20 mL of trifluoroacetic acid for 1 h at room temperature. The TFA was removed *in vacuo* and residual acid was removed by addition and evaporation of toluene (2 × 10 mL). ¹H NMR (DMSO-d₆, 300 MHz) δ 1.31 (t, 6H, 6.9 Hz), 3.97-4.1 (m, 4H), 6.54 (d, 1H, J = 16.0 Hz), 7.47-7.57 (m, 3H), 7.76 (d, J = 8.1Hz, 2H). ¹⁹F NMR (DMSO-d₆, 282.0 MHz) δ -107.7 (d, J = 122.8

Hz, 2F). ¹³C NMR (DMSO-d₆, 75 MHz) δ 16.5, 16.6, 65.1, 65.2, 121.9, 126.8, 126.9, 128.8, 133.5, 133.7, 137.4, 142.9, 167.7. The crude cinnamic acid derivative (3.5 g, 10.5 mmol), pentachlorophenol (3.1 g, 11.5 mmol), DCC (2.6 g, 12.6 mmol) and DMAP (1.3 g, 1.05 mmol) in 100 mL of EtOAc was stirred at room temperature for 24 h. The mixture was filtered through celite and the solvent removed *in vacuo*. The crude product was purified by silica gel chromatography eluting with 25% EtOAc-hexanes to give 5.1 g (84%) of **4** as a white solid. HRMS (M + 1) calcd 582.9195, found 582.9135; ¹H NMR (CDCl₃, 300 MHz) δ 1.26 (t, 6H, J = 7.2 Hz), 4.1-4.2 (m, 4H), 6.67 (d, 1H, J = 16.0 Hz), 7.63 (s, 4H), 7.9 (d, J = 16.0 Hz, 1H). ¹⁹F NMR (CDCl₃, 282.0 MHz) δ -109.06 (d, J = 114.2 Hz, 2F). ¹³C NMR (CDCl₃, 75 MHz) δ 16.3, 16.4, 64.9, 65.0, 116.7, 127.1, 127.8, 128.5, 131.6, 132.1, 135.9, 144.2, 147.6, 162.2.

Synthesis of Pentachlorophenyl (E)-3-(4-phosphoryldifluoromethyphenyl)prop-2-enoate (*5*): lodotrimethylsilane (2.0 mL, 3.7 mmol) in 5 mL of dry CH₂Cl₂ was added dropwise to a solution of **4** (2.0 g, 3.43 mmol) and bis(trimethylsilyl)trifluoroacetamide (1.8 mL, 6.8 mmol) in 20 mL of dry CH₂Cl₂ at 0°C under argon. Stirring was continued for 1 h at 0°C and 1 h at room temperature. The solution was concentrated *in vacuo*. The residue was taken up in 20 mL MeCN/H₂O/AcOH (8:1:1), stirred for 45 min and concentrated *in vacuo*. Toluene (5 mL) was added and evaporated twice. On addition of ether solids separated, which were collected by filtration and washed with the same solvent to give 1.6 g of **5** as a white powder (89%). HPLC *t*_R 25.22, HRMS (M + H) calcd 526.8569, found 526.8542. 1H NMR (DMSO-d₆, 300 Hz) δ 7.13 (d, J = 15.9 Hz,1H), 7.60 (d, J = 8.1 Hz, 2H), 8.0 (d, J = 8.1 Hz, 1H), 8.08 (d, J = 15.9 Hz, 2H). ¹³C NMR (DMSO-*d*₆, 125 MHz) δ 116.4, 127.2, 127.8, 129.3, 131.2, 131.7, 135.4, 144.3, 148.9, 162.8. ¹⁹F NMR (DMSO-*d*₆, 282 MHz) δ -108.52 (d, 2F, J= 104.3 Hz).

Synthesis of pentachlorophenyl (E)-3-(4-dipivaloyloxymethylphosphoryl-

difluoromethyphenyl)prop-2-enoate (6): NaOH (144 mg, 3.6 mmol) in 2 mL of water was

added dropwise to a stirred suspension of **5** (1 g, 1.9 mmol) in 5 mL of water. When the mixture became clear (pH~10), AgNO₃ (807 mg, 4.75 mmol) was added. After 2 h at 4°C the gray precipitate was collected by filtration, dried, and pulverized in a mortar and pestle. The powder was suspended in dry toluene (10 mL) and pivyloxymethyl iodide (1.4 g, 5.7 mmol) was added and stirred for 24 h at room temperature. After filtration the solvent was removed *in vacuo* and the crude product was purified by silica gel column chromatography eluting with 30% EtOAchexanes to give a colorless sticky liquid of **6** (0.90 g, 64%) which solidified on storage at 4°C. ¹H NMR (CDCl₃, 500 MHz) δ 1.15 (s, 18H), 5.58 (dd, J = 12.3, 5.0 Hz, 2H), 5.67 (dd, J = 12.3, 5.0 Hz, 2H), 6.68 (d, J = 16.0 Hz, 1H), 7.6-7.64 (m, 4H), 7.9 (d, J = 16.0 Hz, 1H). ¹³C NMR (CDCl₃, 125 MHz) δ 26.7, 38.7, 82.4, 82.5, 117.0, 127.2, 127.8, 128.6, 131.6, 132.1, 136.3, 144.1, 147.4, 162.2, 176.5. ¹⁹F NMR (CDCl₃, 282 MHz) δ -109.54 (d, J = 123.4 Hz, 2F). ³¹P NMR (CDCl₃ 202 MHz) δ 4.65 (m, 1P). Proton Decoupled ³¹P NMR (CDCl₃ 202 MHz) δ 4.64 (t, J = 123.2 Hz, 1P). Anal. Calcd for C₂₈H₂₈Cl₅F₂O₉P: C, 44.56; H, 3.74; Cl, 23.49; F, 5.03. Found: C, 44.60; H, 3.76; Cl, 23.46; F, 4.95. HRMS (M + Na) calcd 776.9750, found 776.9718.

Synthesis of TFA-H-Haic-GIn-NHBn (7): Fmoc-Haic-GIn-NHBn was prepared on solid phase using 0.300 g (0.36 mmol) of rink resin as described by Mandal et al.³ The Fmoc group was removed by treatment with 20% piperidine/DMF and the resin then treated with 3×10 mL of trifluoroacetic acid: triisopropylsilane: H₂O (TFA:TIS:H₂O) (95:2.5:2.5 v/v/v)⁴ and filtered. The combined filtrates were evaporated to ca 5 mL and the product was isolated by precipitation in Et₂O and collected by centrifugation. The crude peptide was purified by reverse phase HPLC, using 0.1% TFA-water/acetonitrile, lyophilized, and dried *in vacuo* over P₂O₅ at 37°C to get 106 mg of a white solid. HPLC *t*_R16.28, (Purity, 100% at 230 nm, 95% at 275 nM). LC-MS (M + H) calcd 464.2298, found 464.1441. ¹H NMR (DMSO-d₆ 500 MHz) δ 1.78-1.83 (m, 1H), 1.86-1.94 (m, 1H), 2.03-2.19 (m, 4H), 2.96 (d, J = 15.0 Hz, 1H), 3.14 (m, 2H), 3.44 (m, 1H), 4.22 (m, 2H),

4.28 (t, J = 6.0 Hz, 1H), 5.17 (dd, J = 2.5, 11.0 Hz, 1H), 6.8 (s, 1H), 7.02 (t, J = 7.5Hz, 1H), 7.08 (d, J = 7.5 Hz, 1H), 7.12 (d, J = 7.5 Hz, 1H), 7.21 -7.25 (m, 3H), 7.28-7.30 (m, 2H), 8.36 (m, 2H), 8.43 (m, 2H). ¹³C NMR (DMSO-d₆ 125 MHz) δ 28.5, 31.2, 31.9, 32.3, 52.9, 60.8, 124.4, 127.3, 127.5, 128.7, 171.1, 171.52.

Preparation of BP-PM6: To a stirred solution of 7 (0.050 g, 0.086 mmol), NMM (0.03 mL, 0.26 mmol) and DMAP (0.005 g, 0.04 mmol) in 2 mL of anhydrous NMP was added a solution of 6 (0.070 g, 0.091 mmol) in 2 mL of dry CH₂Cl₂ under argon. The reaction was monitored by HPLC. After completion, about 2 h, the reaction mixture was concentrated under vacuum and triturated with hexane-ether. The solid residue (0.102 g) was taken up in 2 mL of MeCN and purified by reverse phase HPLC using a gradient of MeCN in H_2O (no TFA in the eluant). Yield: 0.060 g (72%). HPLC t_{R} 25.80 (purity>98%), HRMS (M + H) calcd 952.3709, found 952.3412. ¹⁹F NMR (Acetone- d_6 282 MHz) δ -108.0 (d, J =123.0, 2F). Proton Decoupled ³¹P NMR (Acetone- d_{6} 202 MHz) δ 4.1 (tr, J = 123.0 Hz, 1P). ¹H NMR (Acetone- d_{6} 500 MHz) δ 1.2 (s, 18H), 1.95-2.00 (m, 1H), 2.1-2.2 (m, 1H), 2.24-2.34 (m, 4H), 3.13-3.22 (m, 3H), 3.5 (m, 1H), 4.4 (d, J = 5.5Hz, 2H), 4.44 (m, 1H), 4.65-4.7 (m, 1H), 5.24 (m, 1H), 5.72-5.8 (m, 4H), 6.95-7.02 (m, 2H), 7.1 (m, 1H), 7.2-7.21 (m, 0.8H), 7.25-7.28 (m, 3H), 7.6-7.66 (m, 3H), 7.8 (d, J = 8.0 Hz, 1H), 8.2 (d, J = 8.0Hz, 1H), 8.28 (t, J = 6.0 Hz, 1H), 8.4 (d, J = 7.0 Hz, 0.2 H). 13 C NMR (Acetone-d₆ 125 MHz) δ 26.1, 27.9, 30.8, 31.3, 38.4, 42.5, 52.8, 53.7, 61.6, 82.7, 123.2, 123.6, 124.3, 126.5, 126.9, 127.3, 128.1, 128.4, 129.5, 132.8, 138.5, 138.9, 139.0, 165.0, 170.4, 171.5, 171.7, 175.5, 176.3.

Synthesis of TFA-H-Leu-Pro-Gln-NHBn (8): Fmoc-Leu-Pro-Gln-NHBn was prepared on solid phase using 0.200 g (0.24 mmol) of rink resin as described by Mandal et al.³ The Fmoc group was removed by treatment with 20% piperidine/DMF and the peptide was cleaved and isolated as in the case of TFA-H-Haic-Gln-NHBn. The crude peptide was purified by reverse phase

HPLC, using a gradient of 0.1%TFA-water/acetonitrile, to get 88 mg of white solid after drying *in vacuo* over P₂O₅ at 37°C for 24 hr. HPLC t_R 9.17, (Purity, 95%) HRMS (M + H) calcd 446.2767, found 446.3924. ¹H NMR (DMSO-d₆ 500 MHz) δ 0.9-0.95 (m, 7H), 1.55 (m, 2H), 1.78-1.98 (m, 7H), 2.01-2.13 (m, 3H), 3.42 (m, 1H), 3.72 (m, 1H), 4.12 (m, 1H), 4.22 (m, 1H), 4.28-4.3 (m, 2H), 4.42-4.45(m 1H), 6.8 (s, 1H), 7.23-7.33 (m, 7H), 8.15-8.2 (m, 4H), 8.37 (m, 1H).¹³C NMR (DMSO-d₆ 125 MHz) δ 21.6, 23.6, 23.8, 25.1, 28.4, 29.5, 31.9, 42.5, 47.3, 49.9, 53.0, 60.1, 127.2, 127.5, 128.7, 139.8, 158.4, 158.7,168.2, 171.3, 171.6, 174.3.

Preparation of BP-PM279G: To a stirred solution of **8** (0.042 g, 0.075 mmol), NMM (0.025 mL, 0.22 mmol) and DMAP (0.005 g, 0.04 mmol) in 5 mL of 50:50 anhydrous MeCN/CH₂Cl₂, was added 6 (0.060 g, 0.078 mmol) under inert atmosphere. The reaction was monitored by HPLC. After completion, about 2 h, the reaction mixture was concentrated under vacuum and triturated with ether. The solid residue (0.092 g) was taken up in 2 mL of MeCN and purified by reverse phase HPLC using a gradient of MeCN in H_2O (no TFA in the eluant). Yield: 53 mg (75%). HPLC $t_{\rm R}$ 26.29 (purity >98%), HRMS (M + H) calcd 934.4179, found 934.4190. ¹⁹F NMR (Acetone-d₆ 282 MHz) δ -109.43 (d, J =124.0, 2F). Proton Decoupled ³¹P NMR (Acetone-d₆ 202 MHz) δ 4.3 (t, J = 121.0 Hz, 1P). ¹H NMR (Acetone-d₆ 500 MHz) δ 0.93 (d, J = 6.0 Hz, 3H), 0.97 (d, J = 6.0 Hz, 3H), 1.22 (s, 18H), 1.53-1.58 (m, 1H), 1.67--1.74 (m, 2H), 1.93-2.0 (m, 2H), 2.18-2.23 (m, 1H), 2.3-2.4 (m, 2H), 3.85 (m, 1H), 3.94 (m, 1H), 4.3-4.37 (m, 3H), 4.43-4.49 (m, 1H), 4.93 (m, 1H), 5.72-5.81 (m, 4H), 6.5 (s, 0.5H), 6.9 (d, J = 16.0 Hz, 1H), 7.14 (s, 0.5), 7.23 (t, J = 7.0 Hz, 1H), 7.27-7.35 (m, 4H), 7.57 (d, J = 7.5 Hz, 1H), 7.62-7.7 (m, 4H), 7.77 (d, J = 8.0 Hz, 2H), 7.86 (m, 1H), 8.21(d, J = 7.0 Hz, 1H). ¹³C NMR (Acetone-d₆ 125 MHz) δ 21.1, 22.8, 24.5, 24.9, 26.1, 26.7, 31.5, 38.4, 40.4, 42.4, 47.4, 49.7, 53.5, 61.6, 82.6, 123.6, 126.6, 126.8, 127.2, 127.3, 127.9, 128.2, 138.2, 138.7, 139.7, 164.6, 171.1, 171.5, 172.4, 175.5, 176.0.

Synthesis of F2PmCinn-Haic-Gin-NHBn (NP-PM6): Fmoc-Haic-Gin-NHBn was assembled on 0.2 g (0.12 mmol) of Rink resin using the general procedures described above. Compound **5** (0.095 g, 0.18 mmol), HOBt (0.027 g, 0.18 mmol) and DIEA (31 μL, 0.18 mmol) in 5 mL of DMF was added. After 3 hr the ninhydrin test was negative. The peptide was cleaved from the resin with trifluoroacetic acid: triisopropylsilane: H₂O (95:2.5:2.5 v/v/v) and purified as above to give 43 mg of final peptide. T_R (min.) (0.1%TFA/MeCN-H₂O): 13.64 (Purity: > 99%). HRMS (M+H) Cald: 724.2348. Found 724.2620. ¹H NMR (DMSO-*d*6 500 MHz) δ 1.38-1.48 (m, 2H), 1.67 (m, 1H), 1.80 (m, 1H), 1.98-2.04 (m, 4H), 2.87 (d, J = 16.0 Hz, 1H), 2.95-3.06 (m, 2H), 3.32 (m, 2H), 4.12-4.22 (m, 3H), 4.42 (m, 1H), 5.03 (d, J = 8.5 Hz, 1H), 6.66 (s, 1H), 6.8 (d, J = 15.5 Hz, 1H), 6.88 (t, J = 7.0 Hz, 1H), 6.99 (m, 1H), 7.11-7.2 (m, 5H), 7.36 (d, J = 16.0 Hz, 1H), 7.48 (s, 4H), 8.18 (d, J = 8.0 Hz, 1H), 8.32 (m, 1H), 8.50 (d, J = 7.0 Hz, 1H). ¹³C NMR (DMSO-*d*6 125 MHz) δ 22.4, 22.7, 28.5, 29.5, 31.5, 31.9, 32.1, 42.5, 44.1, 52.9, 53.4, 61.0, 123.0, 124.0, 125.9, 127.1, 127.2, 127.5, 128.7, 129.7, 133.1, 135.7, 139.1, 139.4, 139.7, 164.9, 169.6, 171.0, 171.6, 174.1. ³¹P NMR (DMSO-*d*6 202 MHz) δ 0.59 (t, J = 90.9 Hz, 1P).

Synthesis of MP-PM6: From failed solid phase synthesis of **BP-PM6**. 22 mg of desired material was obtained after HPLC purification. T_R (min) (0.1%TFA/MeCN-H₂O): 19.78 (Purity: > 97%). HRMS (M+H) Cald: 838.3029. Found 838.3032. ¹H NMR (DMSO-*d*6 500 MHz) δ1.06 (s, 9H), 1.68 (m, 1H), 1.80 (m, 1H), 1.94-2.08 (m, 4H), 2.87 (d, J = 16.5 Hz, 1H), 2.95-3.1 (m, 2H), 3.35 (m, 1H), 4.11-4.22 (m, 3H), 4.43 (t, J = 8.0 Hz, 1H), 5.04 (m, 1H), 5.38 (d, J = 11.5 Hz, 2H), 6.84 (d, J = 16.0 Hz, 1H), 6.9 (t, J = 7.0 Hz, 1H), 7.00 (t, J = 8.0 Hz, 1H), 7.11-7.21 (m, 5H), 7.38 (d, J = 16.0 Hz, 1H), 7.47 (d, J = 8.0 Hz, 2H), 7.55 (d, J = 8.0 Hz, 2H), 8.15 (d, J = 8.0 Hz, 1H), 8.31 (t, J = 6.0 Hz, 1H), 8.51 (d, J = 7.5 Hz, 1H). ¹³C NMR (DMSO-*d*6 125 MHz) δ 27.1, 28.5, 29.5, 31.5, 31.8, 38.6, 42.5, 52.8, 53.4, 54.6, 61.0, 83.4, 117.9, 123.5, 125.9, 127.2, 127.5, 128.7,

129.7, 133.1, 136.7, 138.7, 139.4, 139.7, 169.6, 171.6, 174.1, 176.8, 197.7. ³¹P NMR (DMSO*d*6 202 MHz) δ 0.61 (t, J = 101.0 Hz, 1P).

Inhibition of Stat3 tyrosine 705 phosphorylation in MDA-MB-468 cells.

MDA-MB-468 breast tumor cells (0.4×10^6) were plated onto 35 mm culture dishes and were allowed to grow overnight. Prodrugs were prepared as 10 mM stock solutions in DMSO and aliquots were added to the culture media to give the correct final concentrations. After 2 h the cells were washed with ice cold phosphate buffered saline. Washed cells were treated with lysis buffer (50 mM Hepes, pH 7.4, 150 mM NaCl, 1.5 mM MgCl₂, 1 mM EGTA, 100 mM NaF, 10 mM sodium pyrophosphate, 10% glycerol, 1% Triton X-100, 1 mM PMSF, 1 mM Na₃VO₄, 10 µg/mL leupeptin and 10 µg/mL aprotinin). Cell-free detergent extracts were centrifuged at 15,000 rpm in a microcentriguge for 30 min at 4°C and the protein concentrations of the supernatants determined. Aliquots containing 12 µg of protein were separated on 8% SDS-PAGE and were transferred to PVDF filters. The filters were blocked with 5% bovine serum albumin and were probed with pStat3^{Y705} antibody followed by secondary antibody, whose signal was detected with an enhanced chemiluminescence kit (ECL, Amersham, Chicago, IL). Filters were stripped with stripping buffer (62.5 mM Tris, pH 6.8, 2% SDS, and 0.1 M 2mercaptoethanol) at 50°C for 30 min. Filters were then probed with total Stat3 antibody and visualized with chemiluminescence as above.

References for the Supporting Information

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S11

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25						
-30						
mdd		F12 - Proc	CPDPRG2 NUC2 PCPD2 PL2 PL12 PL13 SFO2	======= NUC1 P11 PL1 SF01	F2 - Acqu Date Time INSTRUM PROBHD PULPROG SOLVENT NS SOLVENT NS SWH FIDRES AQ RG DW DE TE DU DE TE D1 d11 DELTA	Current D NAME EXPNO PROCNO
	S38	essing parameters 32768 202.4563350 MHz EM 0 1.00 Hz 1.40	CHANNEL f2 ====== waltz16 1H 90.00 usec -2.00 dB 18.50 dB 20.00 dB 20.00 dB 20.00 dB	CHANNEL fl ====== 31P 15.00 usec -4.50 dB 202.4462121 MHz	11 sition Parameters 20090525 10.43 spect 5 mm TXI 1H/D- zgpg30 65536 Acetone 16 4 80645.164 Hz 1.230548 Hz 0.4063794 sec 20642.5 6.200 usec 7.50 usec 2.00000000 sec 1.89999998 sec 1	Nata Parameters 675_3-1 5

BP-PM6
F2 - Acquisition Parameters

Date
2009022

Time
11.15

INSTRUM
spect

PROMID
mm QNP 1H/19

POLIPRG
Sgfhidgn

SOLVENT
CDC13

SOLVENT
CDC13

SWH
0.515500

FIDRES
67567.570

AQ
0.515500

PODE
0.50500

DE
0.515500

DE
0.509928

DE
0.000208

DE
0.0002000

DE
0.0022

DE
0.00200000

DI
0.00200000

DI
0.0020000

DI
0.00020000

DI
0.00002000

DI
0.00002000

DI
0.00002000

DI
0.00002000

DI
0.00002000

DI
0.00002000

DI
0.00002000
sec

DI
0.00002000
sec

DI
0.00002000
s F2 - Processing parameters SI SF 282.404350 MHz MDW SSB SB SB 0.0 LB 0.0 C B D D D D D D D 1.0 0 1.0 0 CPDPRG2 NUC2 PCPD2 PL2 PL12 SFO2 NUC1 P1 PL1 SF01 = CHANNEL f1 _____ 19F 12.00 usec -4.00 dB 282.3761148 MHz 67567.577 Hz 0.515500 Hz 0.969928 sec 4096 7.400 Usec 6.00 Usec 1.0000000 sec 0.300.2 X 1.0000000 sec 0.3000200 sec S39

Current Data Parameters NAME 67G_1 EXPNO 7 PROCNO 1

-103

-104

-105

-106

-107

-108

-109

110

| _____ ____

122

143

uudd



Sample Name: 67G 1-2

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RUK







								·			DE TE DI DELTA TDO NUC1 P1 SF01 SF01 CPDPRG2 PL12 PL12 PL12 PL12 PL12 PL12 SF02 F2 - PrC SF WDW SSB LB GB PC	298.2 K 298.2 K 2.0000000 sec 1.89999998 sec 1 CHANNEL f1 15.00 usec -4.50 dB 202.4462121 MHz CHANNEL f2 waltz16 18.50 dB 20.00 dB 18.50 dB 20.00 dB 500.1320005 MHz bccessing parameters 32768 202.4563350 MHz EM 0 1.00 Hz 0 1.40
***	25	20	15	10	5 5 546	0	-5		-15	ppm		









Instrument 1 11/19/2007 5:02:09 PM DS



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Sample Name: F2 CINN

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Data File C:\HPCHEM\1\DATA\081-0301.D

Sample Name: 67G 10-15