

Supplementary Data

A New Usage of Functionalized Oligodeoxynucleotide Probe for Site-Specific Modification of a Guanine Base within RNA

Kazumitsu Onizuka, Yosuke Taniguchi, Shigeki Sasaki,*

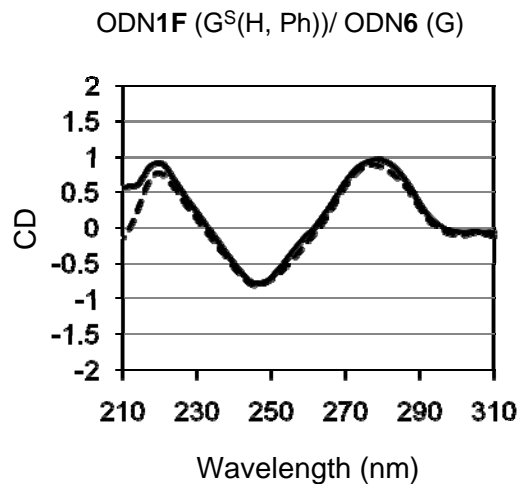
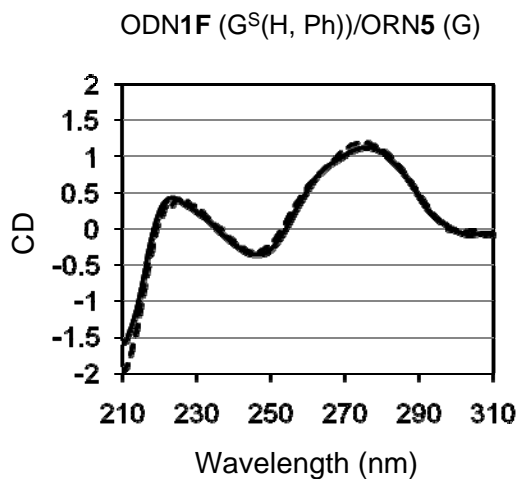
Graduate School of Pharmaceutical Sciences, Kyushu University, 3-1-1 Maidashi,
Higashi-ku, Fukuoka 812-8582, Japan, and CREST, Japan Science and
Technology Agency.

sasaki@phar.kyushu-u.ac.jp

5'-d (CTTT~~X~~TTCTCCTTTCT)
3'-r (GAAA~~Y~~AAGAGGAAAGA)

5'-d (CTTT~~X~~TTCTCCTTTCT)
3'-d (GAAA~~Y~~AAGAGGAAAGA)

CD spectra



UV melting curves

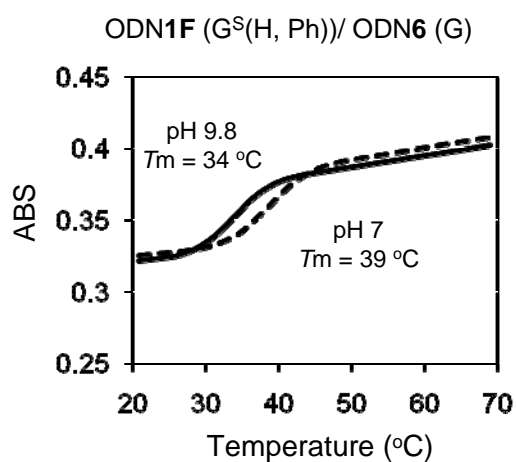
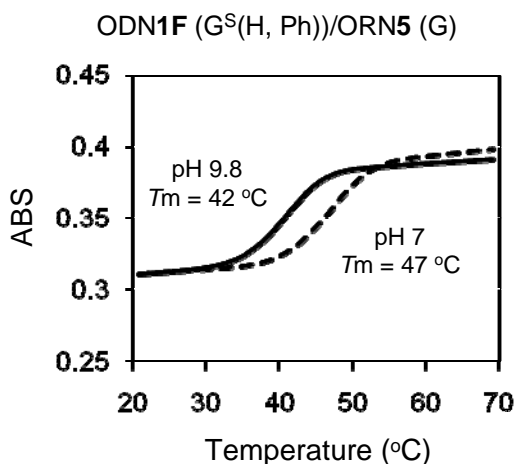


Figure S1. CD spectra and UV melting curves. CD spectra were measured using 2.5 μM duplexes in 50 mM MES buffer (pH 7, broken line) or 50 mM carbonate buffer (pH 9.8, solid line) containing 100 mM NaCl. Melting curves were measured using 1.3 μM duplexes in 50 mM MES buffer (pH 7) or 50 mM carbonate buffer (pH 9.8) containing 100 mM NaCl.

pH dependency (DNA)

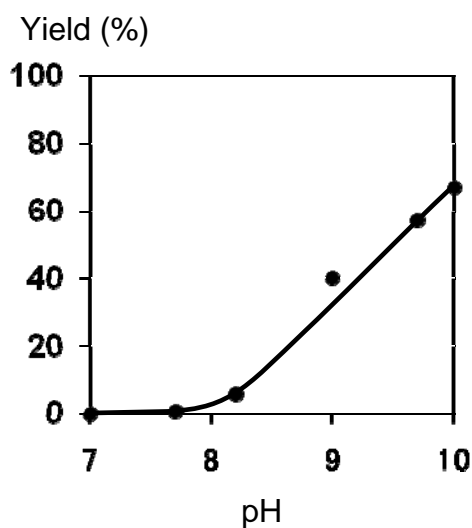
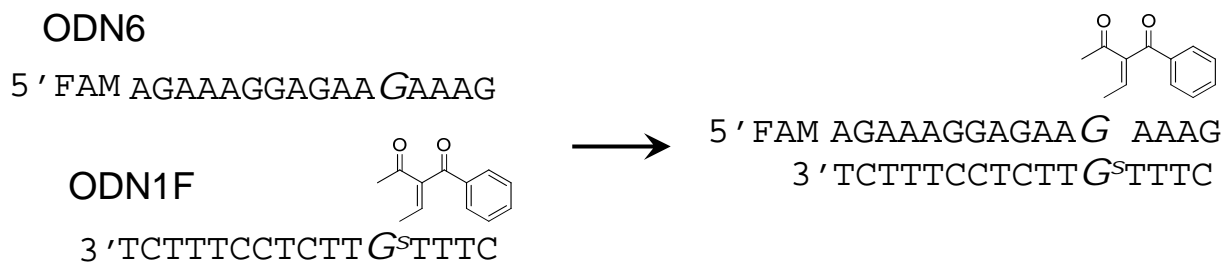


Figure S2. The pH dependency of the transfer reaction with DNA substrate. The transfer reactions were performed by using 1.5 μM of S-functionalized ODN1F (G^{S} (Me, Ph)) and 1.0 μM of the target ODN6 (G) in 50 mM phosphate or carbonate buffer (pH > 9) at 25°C for 1h, and followed by HPLC (Column: SHISEIDO C18, 4.6 x 250 mm; Solvent: A: 0.1 M TEAA Buffer, B: CH_3CN , B: 10% to 30% /20 min, linear gradient; Flow rate: 1.0 ml/min; monitored by fluorescence detector at 518 nm with emission at 494 nm).

Base selectivity (DNA)

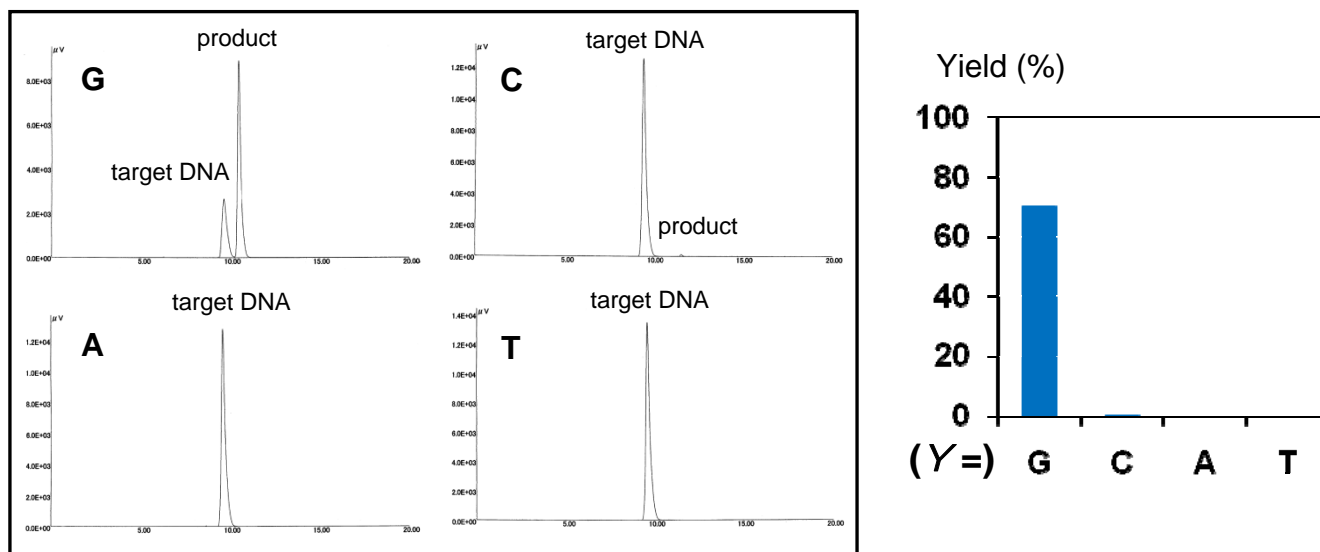
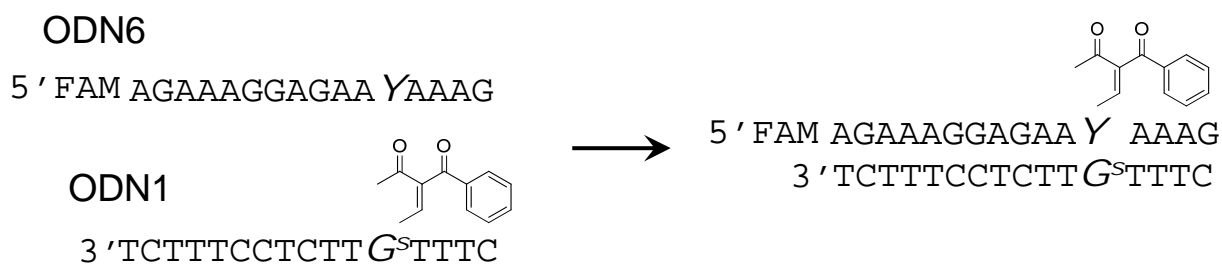
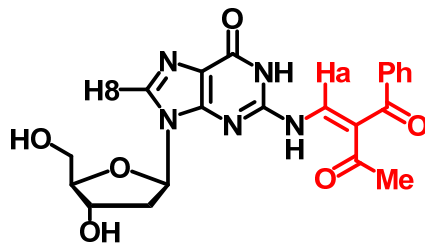
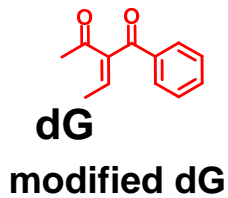


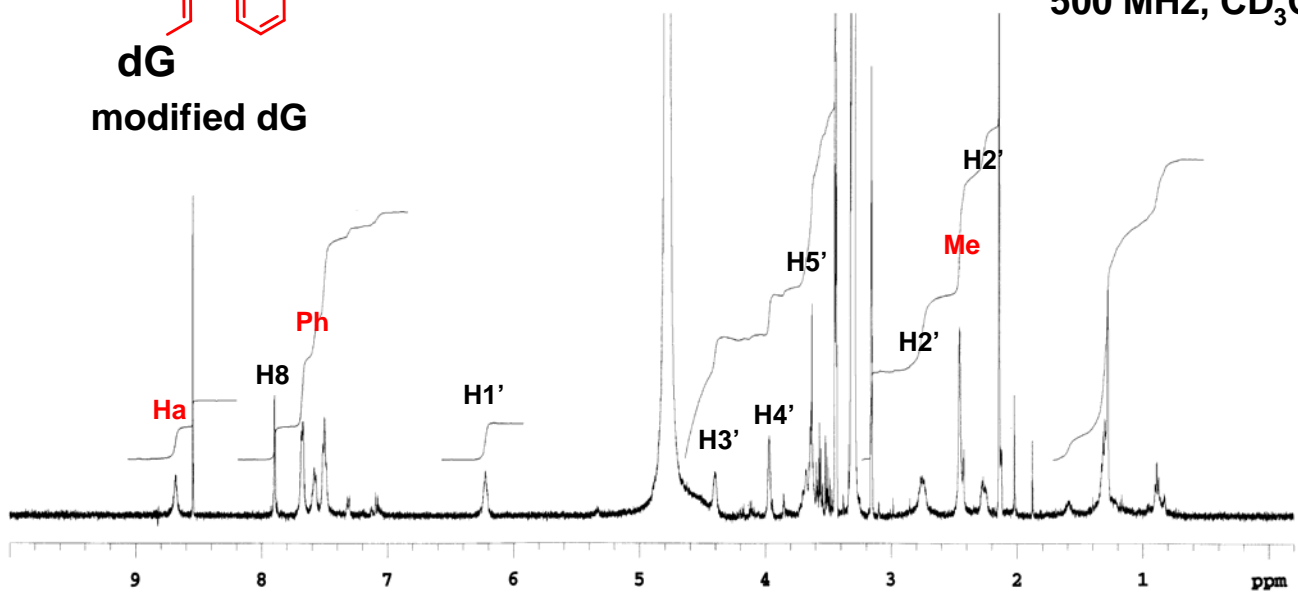
Figure S3. Selectivity to the guanine base in the DNA substrates. The transfer reactions were performed by using 1.5 μ M of S-functionalized ODN1F(G^S(Me, Ph)) and 1.0 μ M of the target ODN6 (Y) in 50 mM carbonate buffer containing 100 mM NaCl at pH 10 for 2h, and followed by HPLC (Column: SHISEIDO C18, 4.6 x 250 mm; Solvent: A: 0.1 M TEAA Buffer, B: CH₃CN, B: 10% to 30% /20 min, linear gradient; Flow rate: 1.0 ml/min; monitored by fluorescence detector at 518 nm with emission at 494 nm).



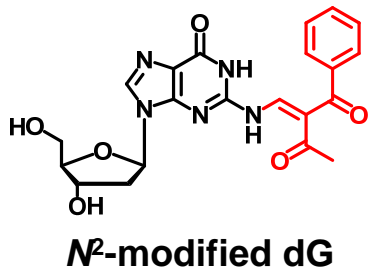
Hydrolysate of the modified DNA



500 MHz, CD₃OD



Synthesized compound



500 MHz, CD₃OD

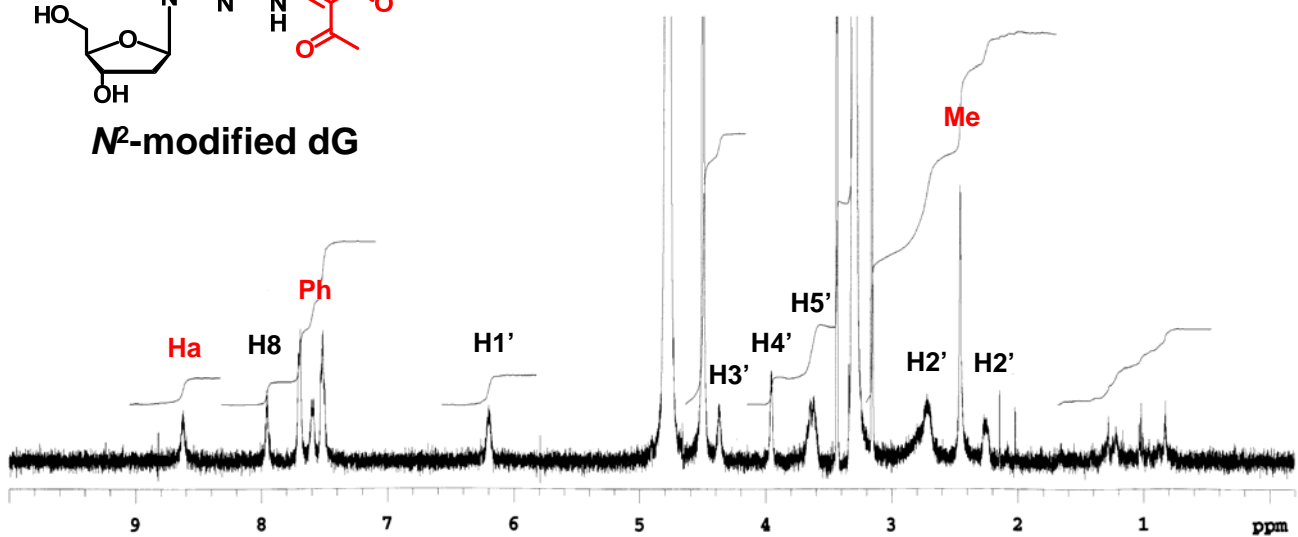
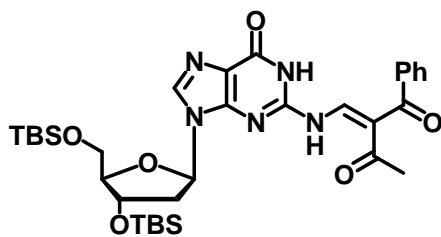


Figure S4. ¹H-NMR measurements



K01071-2,h1

Pulse Sequence: s2pu1
 DATE Jan 16 2009
 SOLVENT cdcl3
 OBSERVE H1
 FREQUENCY 399.873 MHz
 SPECTRAL WIDTH 11999.4 Hz
 ACQUISITION TIME 2.731 sec
 RELAXATION DELAY 2.269 sec
 PULSE WIDTH 6.0 usec
 TEMPERATURE 30.0 deg. C.
 NO. REPETITIONS 16
 DOUBLE PRECISION ACQUISITION
 DATA PROCESSING
 RESOLUTION ENHANCEMENT -0.0 Hz
 FT SIZE 65536
 TOTAL ACQUISITION TIME 1 minutes

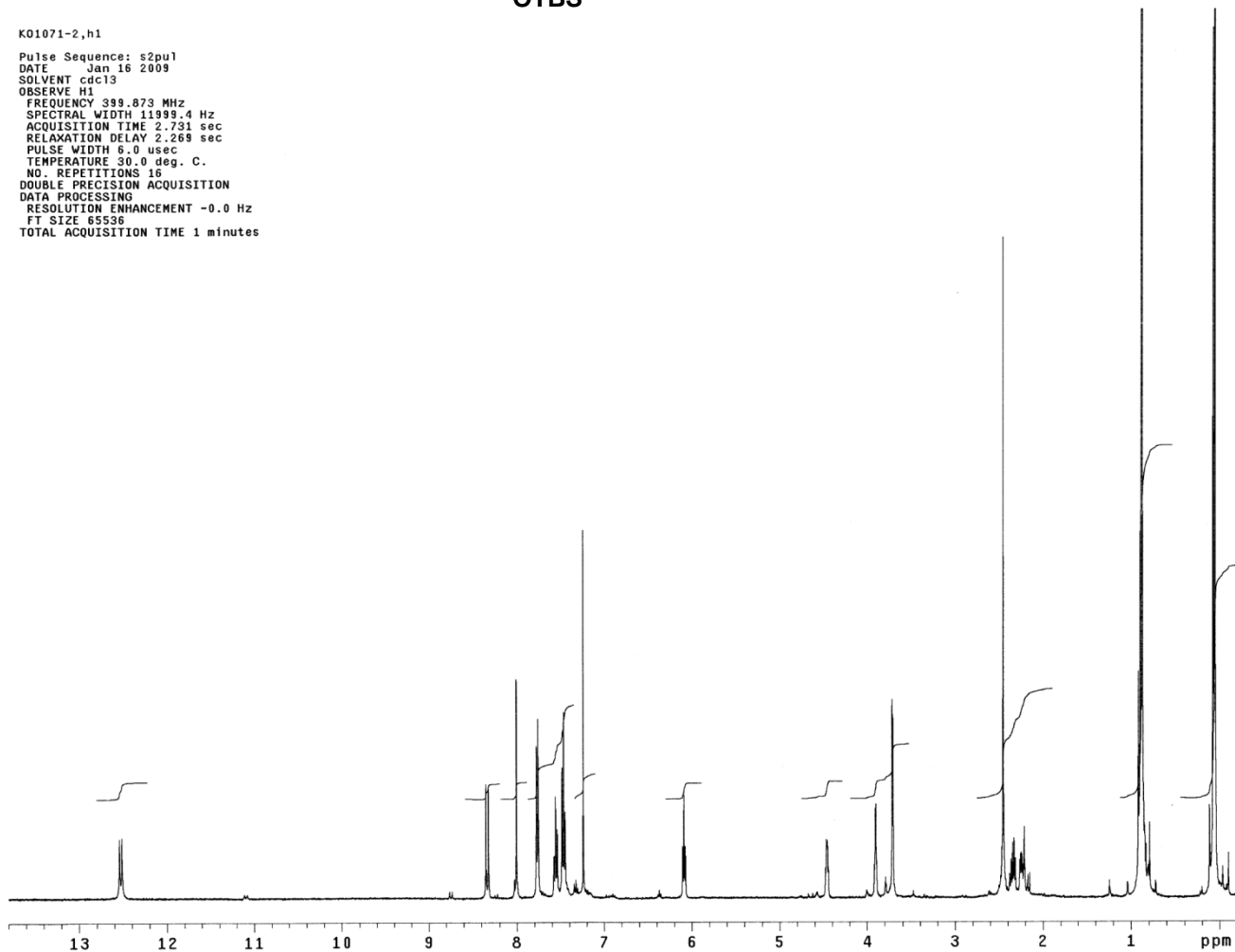


Table S1. UV Melting temperatures of the duplexes formed with.

ODN	Complementary strand	Y =	pH	T _m (°C)
ODN1-G ^S	ORN5	rG	9.8	39
ODN1F-G ^S (H, Ph)	ORN5	rG	9.8	42
ODN1F-G ^S (H, Ph)	ORN5 (FAM)	rG	9.8	43
ODN1F-G ^S (H, Ph)	ORN5 (FAM)	rC	9.8	42
ODN1F-G ^S (H, Ph)	ORN5 (FAM)	rA	9.8	44
ODN1F-G ^S (H, Ph)	ORN5 (FAM)	rU	9.8	43
ODN1F-G ^S (H, Ph)	ORN5	rG	7	47
ODN1-G ^S (H, Ph)	ORN5 (FAM)	rG	7	49
ODN1F-G ^S (H, Ph)	ORN5 (FAM)	rC	7	49
ODN1F-G ^S (H, Ph)	ORN5 (FAM)	rA	7	49
ODN1F-G ^S (H, Ph)	ORN5 (FAM)	rU	7	48
ODN1F-G ^S (H, Ph)	ODN6 (FAM)	dG	9.8	35
ODN1F-G ^S (H, Ph)	ODN6 (FAM)	dC	9.8	37
ODN1F-G ^S (H, Ph)	ODN6 (FAM)	dA	9.8	35
ODN1F-G ^S (H, Ph)	ODN6 (FAM)	dT	9.8	35
ODN1F-G ^S (H, Ph)	ODN6	dG	9.8	34
ODN1F-G ^S (H, Ph)	ODN6	dHx	9.8	33
ODN1F-G ^S (H, Ph)	ODN6	d2AP	9.8	34
ODN1F-G ^S (H, Ph)	ODN6	dG	7	39
ODN1F-G ^S (H, Ph)	ODN6	dHx	7	38
ODN1F-G ^S (H, Ph)	ODN6	d2AP	7	39
ODN2F-G ^S (H, Ph)	ORN7 (FAM)		9.8	45
ODN2F-G ^S (H, Ph)	ORN8 (FAM)		9.8	45
ODN3F-G ^S (H, Ph)	ORN7 (FAM)		9.8	42
ODN3F-G ^S (H, Ph)	ORN9 (FAM)		9.8	45
ODN4F-G ^S (H, Ph)	ORN10 (FAM)	rG	9.8	36
ODN4F-G ^S (H, Ph)	ORN10(FAM)	rC	9.8	37
ODN4F-G ^S (H, Ph)	ORN10(FAM)	rA	9.8	36
ODN4F-G ^S (H, Ph)	ORN10 (FAM)	rU	9.8	36

Table S2. MALDI-TOF.MS Data of all ODN and ORN compounds used in this study.

DNA or RNA	5' Label	X or Y	Calcd ([M-H] ⁻)	Found
ODN1		X = G ^S	4767.8	4766.5
ODN1F		X = G ^S (Me, Ph)	4939.7	4941.5
ODN1F		X = G ^S (Me, Pyrene)	5063.9	5064.4
ODN1 F		X = G ^S (H, Ph)	4897.8	4898.8
ODN2		X = G ^S	4742.7	4742.8
ODN2 F		X = G ^S (Me, Ph)	4914.7	4914.2
ODN2 F		X = G ^S (Me, Pyrene)	5038.8	5037.8
ODN2F		X = G ^S (H, Ph)	4872.7	4872.9
ODN3		X = G ^S	4742.7	4742.4
ODN3F		X = G ^S (Me, Ph)	4914.7	4914.0
ODN3F		X = G ^S (Me, Pyrene)	5038.8	5038.4
ODN3F		X = G ^S (H, Ph)	4872.7	4872.7
ODN4		X = G ^S	6719.1	6719.1
ODN4 F		X = G ^S (Me, Ph)	6891.1	6890.5
ODN4 F		X = G ^S (H, Ph)	6849.1	6849.0
ORN5		Y = rG	5297.8	5297.8
ORN5	FAM	Y = rG	5835.9	5835.9
ORN5	FAM	Y = rC	5795.9	5795.1
ORN5	FAM	Y = rA	5819.9	5819.5
ORN5	FAM	Y = rU	5796.9	5796.5
ORN5		Y = rG(Me, Ph)	5469.8	5469.6
ORN5		Y = rG(Me, Pyrene)	5593.9	5593.8
ODN6	FAM	Y = dG	5579.1	5579.5
ODN6	FAM	Y = dC	5539.1	5538.7
ODN6	FAM	Y = dA	5563.1	5563.5
ODN6	FAM	Y = dT	5554.1	5553.7
ODN6		Y = dG	5041.9	5091.6
ODN6		Y = dHx	5026.9	5026.7
ODN6		Y = d2AP	5026.0	5025.4
ODN6		Y = dG(Me, Ph)	5214.0	5213.6
ODN6		Y = d2AP(Me, Ph)	5198.0	5198.3
ORN7	FAM		5851.9	5851.3
ORN8	FAM		5811.9	5811.3
ORN9	FAM		5811.9	5811.2
ORN10	FAM	Y = rG	9145.1	9145.4
ORN10	FAM	Y = rC	9105.1	9105.9
ORN10	FAM	Y = rA	9129.1	9129.0
ORN10	FAM	Y = rU	9106.1	9106.3