

## Supporting information

### Toward Palau' amine; Hg(OTf)<sub>2</sub>-Catalyzed Synthesis of Cyclopentane Core

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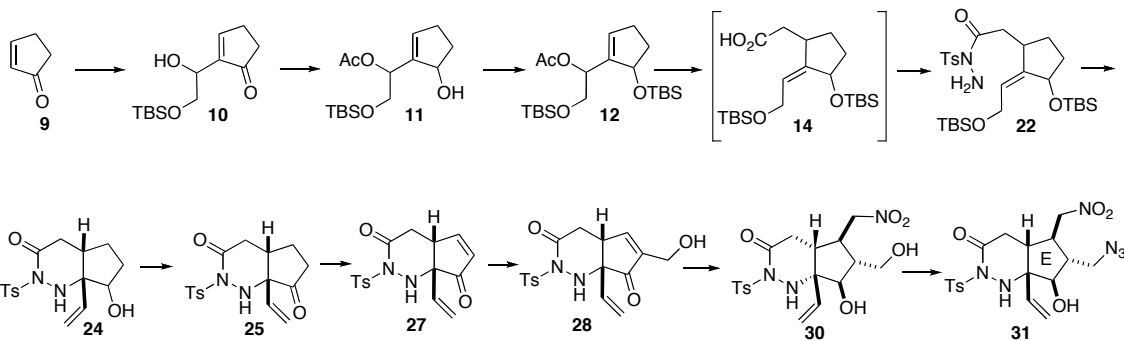
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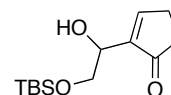


## General Procedures and Methods

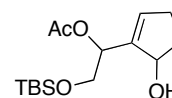
FTIR spectra were measured on a JASCO FT/IR-410 infrared spectrophotometer. NMR spectra were recorded on a Varian Mercury plus-300-4N spectrometer. Chemical shifts are reported in parts per million (ppm). For  $^1\text{H}$  NMR spectra ( $\text{CDCl}_3$ ), the residual solvent peak was used as the internal reference (7.24 ppm), whereas the central solvent peak as the reference (77.03 ppm) for  $^{13}\text{C}$  NMR spectra ( $\text{CDCl}_3$ ). Mass spectra were recorded on a JEOL the Mstation JMS-700. Analytical thin layer chromatography (TLC) was performed with E. Merck pre-coated TLC plates, silica gel 60F-254, layer thickness 0.25 mm. Flash chromatography was performed on Kanto Chemical 60 (63-210) mesh silica gel. Reagents and solvents are commercial grade and were used as supplied. Mercury triflate was prepared by the following procedure; to a suspension of  $\text{HgO}$  (541.4 mg, 2.5 mmol) in acetonitrile (5 mL) was dropwise added  $\text{Tf}_2\text{O}$  (705.3 mg, 2.5 mmol) at  $0\text{ }^\circ\text{C}$ , and the mixture was stirred at  $0\text{ }^\circ\text{C}$  until the yellow color disappear. The resulting colorless solution was transferred to 25 mL of messflask and diluted with anhydrous acetonitrile to give 0.1M solution.

### 2-(2-(*tert*-butyldimethylsilyloxy)-1-hydroxyethyl)cyclopent-2-enone (**10**)

To a solution of 2-cyclopentene-1-one **9** (4.71 g, 57.4 mmol) in THF (57 mL) was added (*tert*-Butyldimethylsilyloxy)-acetaldehyde (10.0 g, 57.4 mmol) and *n*-tributylphosphine (1.43 mL, 5.74 mmol). The mixture was stirred for 4 h at room temperature and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (elution with hexane/ethyl acetate = 4/1 to 2/1) to give **10** (10.3 g, 70%) as colorless amorphous. IR (neat) 3444, 2928, 2857, 1698, 1252, 1120,  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.63 (td,  $J = 2.8, 1.6$  Hz, 1H), 4.48-4.55 (m, 1H), 3.84 (dd,  $J = 10.0, 3.6$  Hz, 1H), 3.52 (dd,  $J = 10.0, 6.4$  Hz, 1H), 3.11 (d,  $J = 4.8$  Hz, 1H), 2.58-2.68 (m, 2H), 2.41-2.47 (m, 2H), 0.89 (s, 9H), 0.06 (s, 3H), 0.05 (s, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  208.8, 159.9, 145.1, 68.3, 65.7, 35.2, 26.8, 25.8, 18.2, -5.4, -5.5; HRMS (CI)  $m/z$   $[\text{M}+\text{H}]^+$  calcd for  $[\text{C}_{13}\text{H}_{24}\text{O}_3\text{Si}+\text{H}]^+$  257.1573, found 257.1575.



**2-(*tert*-butyldimethylsilyloxy)-1-(5-hydroxycyclopent-1-enyl)ethyl acetate (**11**)**

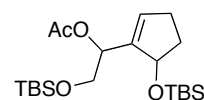


To a solution of **10** (4.0 g, 15.6 mmol) in pyridine (15 mL) was added acetic anhydride (7.37 mL, 78 mmol) at 0 °C. The mixture was stirred for 15 h at 0 °C and concentrated under reduced pressure. The residue was subjected to short-pass flash chromatography on silica gel (elution with hexane/ethyl acetate = 20/1 to 4/1) to give crude acetylated **11** which was used without further purification. To a solution of crude acetylated **11** in methanol (67 mL) was added NaBH<sub>4</sub> (1.0 g, 26.7 mmol) at -20 °C. The mixture was stirred for 1 h, quenched with saturated NH<sub>4</sub>Cl (20 mL), and extracted with diethyl ether (40 mL x 3). The combined organic layers were washed with brine, dried over anhydrous MgSO<sub>4</sub>, filtered, concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (elution with hexane/ethyl acetate = 6/1) to give **11** (2.56 g, 55%) as colorless oil (1.5 : 1 diastereomeric mixture). IR (neat) 3452, 2930, 1740, 1372, 1254, cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 5.80 (br s, 1Ha+1Hb), 5.42 (t, *J* = 4.5 Hz, 1Ha), 5.36 (t, *J* = 5.7 Hz, 1Hb), 4.69 (br s, 1Ha), 4.67 (br s, 1 Hb), 3.79 (dd, *J* = 11.4, 3.9 Hz, 1Ha), 3.78 (d, *J* = 5.7 Hz, 2Hb), 3.71 (dd, *J* = 11.4, 4.8 Hz, 1Ha), 3.50 (br s, 1Ha), 3.11 (br s, 1Hb), 2.32-2.50 (m, 1Ha + 1Hb), 2.04-2.24 (m, 2Ha + 2Hb), 2.01 (s, 3Hb), 2.00 (s, 3Ha), 1.64-1.82 (m, 1Ha + 1Hb), 0.82 (s, 9Ha + 9Hb), 0.01 (s, 6Ha + 6Hb); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 142.7, 142.4, 133.2, 131.8, 76.6, 75.6, 72.6, 71.9, 64.7, 64.4, 33.3, 33.0, 29.9(x2), 25.7(x2), 21.1, 21.0, 18.2, 18.1, -5.4, -5.5; HRMS (CI) *m/z* [M-H]<sup>-</sup> calcd for [C<sub>15</sub>H<sub>28</sub>O<sub>4</sub>Si-H]<sup>-</sup> 299.1678, found 299.1679.

The modified condition was found after the above examination. However, a large scales synthesis of modified condition was not tried, because the sufficient amount of **11** for the cyclopentane core was already obtained. The modified condition is next; to a solution of **10** (45.6 mg, 0.18 mmol) in pyridine (0.5 mL) was added acetic anhydride (84.1 μL, 78 mmol) at 0 °C. The mixture was stirred for 25 h at room temperature and concentrated under reduced pressure. The residue was subjected to short-pass flash chromatography on silica gel (elution with hexane/ethyl acetate = 9/1) to give crude acetylated **11** which was used without further purification. To a solution of crude acetylated **11** in methanol (19 mL) was added NaBH<sub>4</sub> (215 mg, 5.69 mmol) at -78 °C. The mixture was stirred for 1 h, quenched with saturated NH<sub>4</sub>Cl (5 mL), and extracted with diethyl ether (10 mL x 3). The combined organic layers were washed with brine,

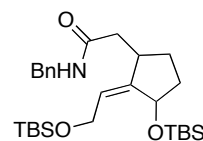
dried over anhydrous MgSO<sub>4</sub>, filtered, concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (elution with hexane/ethyl acetate = 6/1) to give **11** (1.0 g, 70%) as colorless oil (1.5 : 1 diastereomeric mixture).

**2-(*tert*-butyldimethylsilyloxy)-1-(5-(*tert*-butyldimethylsilyloxy)cyclopent-1-enyl)ethyl acetate (**12**)**



To a solution of **11** (3.12 g, 10.4 mmol) in dichloromethane (10.4 mL) was added triethylamine (3.62 mL, 26.0 mmol) and TBSOTf (2.87 mL, 12.5 mmol) successively at 0 °C. The mixture was stirred for 10 min at 0 °C, quenched with sat. NaHCO<sub>3</sub>, and extracted with dichloromethane (x3). The combined organic layers were washed with sat. NaHCO<sub>3</sub>, dried over anhydrous MgSO<sub>4</sub>, filtered, concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (elution with hexane/dichloromethane = 2/1 to 1/1 to 1/4) to give **12** (3.8 g, 88%) as pale yellow amorphous (2 : 1 diastereomeric mixture). **12a**; IR (neat) 2929, 2857, 1741, 1362, 1237 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 5.78 (br s, 1H), 5.26 (br d, *J* = 7.5 Hz, 1H), 4.85 (m, 1H), 3.84 (dd, *J* = 10.8, 3.0 Hz, 1H), 3.63 (dd, *J* = 10.8, 8.1 Hz, 1H), 2.28-2.44 (m, 1H), 2.08-2.26 (m, 2H), 2.00 (s, 3H), 1.52-1.66 (m, 1H), 0.89 (s, 9H), 0.83 (s, 9H), 0.07 (s, 3H), 0.05 (s, 3H), 0.01 (s, 6H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 170.4, 142.8, 130.2, 78.5, 72.6, 64.8, 34.1, 29.9, 25.9, 25.7, 21.2, 18.2, 18.0, -4.3, -4.9, -5.3 (x2); HRMS (CI) *m/z* [M]<sup>+</sup> calcd for [C<sub>21</sub>H<sub>42</sub>O<sub>4</sub>Si<sub>2</sub>]<sup>+</sup> 414.2622, found 414.2623. **12b**; IR (neat) 2929, 2857, 1751, 1362, 1234 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 5.78 (dd, *J* = 7.5, 3.3 Hz, 1H), 5.76 (br s, 1H), 4.73-4.83 (m, 1H), 3.78 (dd, *J* = 11.1, 7.5 Hz, 1H), 3.67 (dd, *J* = 10.8, 3.3 Hz, 1H), 2.34-2.46 (m, 1H), 2.12-2.25 (m, 2H), 2.03 (s, 3H), 1.62-1.74 (m, 1H), 0.88 (s, 9H), 0.83 (s, 9H), 0.07 (s, 3H), 0.06 (s, 3H), 0.01 (s, 6H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 170.1, 142.1, 130.8, 77.2, 73.5, 64.7, 34.4, 29.9, 25.9, 25.7, 21.2, 18.2, 18.0, -4.3, -4.8, -5.3 (x2); HRMS (CI) *m/z* [M-H]<sup>-</sup> calcd for [C<sub>21</sub>H<sub>42</sub>O<sub>4</sub>Si<sub>2</sub>-H]<sup>-</sup> 413.2548, found 413.2547.

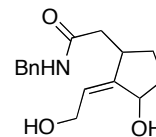
**(Z)-N-benzyl-2-(3-(tert-butyldimethylsilyloxy)-2-(2-(tert-butyl dimethylsilyloxy)ethylidene)cyclopentyl)acetamide (15)**



To a solution of LHMDS (3.46 mL, 1M solution in THF) in THF (10 mL) was added a solution of TBSCl (661 mg, 4.38 mmol) in HMPA (2.97 mL, 17.0 mmol) at -78 °C. The mixture was stirred for 10 min. at -78 °C. To the mixture was added a solution of **12** (784 mg, 1.89 mmol) in THF (2 mL) at -78 °C. The mixture was stirred for 20 min. at 0 °C, quenched with iced water, and extracted with diethyl ether (x2). The combined organic layers were dried over MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure. The solution of residue in toluene (5 mL) was heated to reflux for 1 h, cooled down to room temperature, and concentrated under reduced pressure. To the solution of residue in THF (24 mL) was added H<sub>2</sub>O (7 mL) at room temperature. The mixture was stirred for 3 h, concentrated under reduced pressure to give carboxylic acid **14** which was used without further purification. To a solution of crude **14** in dichloromethane (7 mL) was added benzylamine (125 μL, 1.14 mmol), EDCI (272 mg, 1.42 mmol), and DMAP (23 mg, 0.19 mmol) at room temperature. The mixture was stirred for 25 min., quenched with sat. NH<sub>4</sub>Cl, and extracted with ethyl acetate (20 mL x 3). The combined organic layers were washed with brine, dried over anhydrous MgSO<sub>4</sub>, filtered, concentrated under reduce pressure. The residue was purified by flash column chromatography on silica gel (elution with hexane/ethyl acetate = 10/1 to 8/1 to 6/1 to 4/1 to 2/1) to give **15** (124 mg, 70%) as colorless amorphous (2 : 1 diastereomeric mixture). **15a**: IR (neat) 3286, 2954, 2856, 1644, 1555, 1255, 1008 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.25-7.36 (m, 5H), 5.81 (br s, 1H), 5.38 (ddt, *J* = 6.8, 4.8, 2.4, 1H), 4.62 (t, *J* = 6.8, 1H), 4.48 (dd, *J* = 14.8, 6.0 Hz, 1H), 4.39 (dd, *J* = 14.8, 5.6 Hz, 1H), 4.32 (s, 2H), 2.93-3.05 (m, 1H), 2.55 (dd, *J* = 14.4, 4.8 Hz, 1H), 2.05 (dd, *J* = 14.4, 9.2 Hz, 1H), 1.94-2.08 (m, 2H), 1.48-1.60 (m, 1H), 1.10-1.23 (m, 1H), 0.89 (s, 9H), 0.88 (s, 9H), 0.09 (s, 3H), 0.07 (s, 3H), 0.06 (s, 3H), 0.05 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 171.8, 146.8, 138.3, 128.7, 127.9, 127.5, 125.1, 72.9, 61.2, 43.6, 41.1, 39.3, 34.9, 29.2, 26.0, 25.8, 18.4, 17.9, -3.7, -4.7, -5.0, -5.1; HRMS (CI) *m/z* (M+H)<sup>+</sup> calcd for <sup>+</sup>[C<sub>28</sub>H<sub>49</sub>NO<sub>3</sub>Si<sub>2</sub>+H]<sup>+</sup> 504.3329, found 504.3332. **15b**: IR (neat) 3288, 2955, 1644, 1549, 1254 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.23-7.35 (m, 5H), 5.86 (br t, *J* = 5.6 Hz, 1H), 5.48 (ddt, *J* = 8.0, 4.0, 1.6 Hz, 1H), 4.62-4.67 (m, 1H), 4.43 (d, *J* = 5.6 Hz, 2H), 4.28 (ddd, *J* = 13.2, 8.0, 0.8 Hz, 1H), 4.18 (ddd, *J* = 13.2, 4.0, 1.6 Hz, 1H), 2.87 (quint, *J* = 7.2 Hz, 1H), 2.50 (dd, *J* = 14.4, 6.8 Hz, 1H), 2.30 (dd, *J* = 14.0, 8.0 Hz, 1H),

1.55-1.92 (m, 4H), 0.89 (s, 9H), 0.82 (s, 9H), 0.06 (s, 6H), 0.05 (s, 3H), 0.05 (s, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  171.9, 147.2, 138.4, 128.6, 127.9, 127.4, 125.7, 71.9, 61.1, 43.5, 42.9, 40.4, 35.0, 29.2, 25.9, 25.7, 18.4, 17.8, -4.1, -4.8, -5.1, -5.1; HRMS (CI)  $m/z$  (M+H) $^+$  calcd for  $^+[\text{C}_{28}\text{H}_{49}\text{NO}_3\text{Si}_2+\text{H}]^+$  504.3329, found 504.3322.

**(Z)-N-benzyl-2-(3-hydroxy-2-(2-hydroxyethylidene)cyclopentyl)acetamide (16)**



A solution of **15** (124 mg, 0.25 mmol) in acetic acid (1.5 mL), water (1.5 mL), and THF (1.5 mL) was stirred for 15 h at room temperature and concentrated under reduced pressure. The residue was purified by flash column chromatography of silica gel (elution with dichloromethane/methanol = 20/1) to give **16** (52.3 mg, 70%) as yellow amorphous (2 : 1 diastereomeric mixture). **16a**: IR (neat) 3297, 2929, 1650, 1556, 1455, 1008  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.04-7.08 (m, 5H), 6.19 (br t,  $J$  = 5.6 Hz, 1H), 5.63 (tt,  $J$  = 6.4, 1.6 Hz, 1H), 4.73 (d,  $J$  = 4.8 Hz, 1H), 4.41 (d,  $J$  = 5.6 Hz, 2H), 4.28 (ddd,  $J$  = 12.8, 7.2, 1.6 Hz, 1H), 4.08 (dd,  $J$  = 12.0, 6.4 Hz, 1H), 2.77-2.88 (m, 1H), 2.54 (dd,  $J$  = 14.8, 6.0 Hz, 1H), 2.39 (dd,  $J$  = 14.8, 7.2 Hz, 1H), 1.64-1.94 (m, 4H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  172.0, 152.1, 138.2, 128.7, 127.9, 127.6, 124.2, 71.5, 59.9, 43.6, 41.1, 39.9, 34.1, 29.2; HRMS (CI)  $m/z$  (M+H) $^+$  calcd for  $^+[\text{C}_{16}\text{H}_{21}\text{NO}_3+\text{H}]^+$  276.1599, found 276.1615. **16b**: IR (neat) 3289, 2930, 1638, 1549  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.25-7.40 (m, 5H), 5.78 (br t,  $J$  = 5.6 Hz, 1H), 5.64 (tt,  $J$  = 6.4, 2.0 Hz, 1H), 4.79 (t,  $J$  = 6.4 Hz, 1H), 4.44 (d,  $J$  = 5.6 Hz, 2H), 4.29 (dd,  $J$  = 13.2, 6.4 Hz, 1H), 4.19 (dd,  $J$  = 13.2, 6.4 Hz, 1H), 3.05-3.15 (m, 1H), 2.81 (br s, 1H), 2.55 (br s, 1H), 2.41 (dd,  $J$  = 14.4, 6.0 Hz, 1H), 2.11 (dd,  $J$  = 14.4, 8.4 Hz, 1H), 2.04-2.15 (m, 2H), 1.58-1.70 (m, 1H), 1.25-1.36 (m, 1H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  171.7, 152.4, 138.4, 129.0, 128.2, 127.9, 124.0, 72.4, 60.1, 43.9, 41.6, 40.3, 34.1, 29.7; HRMS (CI)  $m/z$  (M-H) $^-$  calcd for  $[\text{C}_{16}\text{H}_{21}\text{NO}_3-\text{H}]^-$  274.1443., found 274.1440.

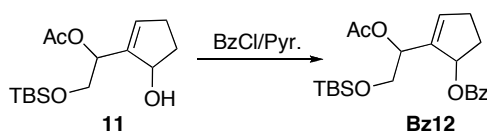
**(Z)-N-benzyl-2-(2-(2-(benzyloxy)ethylidene)-3-hydroxycyclopentyl)acetamide (17)** (2 : 1 diastereomeric mixture)  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.22-7.39 (m, 10Ha+10Hb), 5.85 (br s, 1Hb), 5.72 (br s, 1Ha), 5.42-5.64 (m, 1Ha+1Hb), 4.64-4.72 (m, 1Ha+1Hb), 4.53 (s, 1Ha), 4.52 (s, 1Hb), 4.43 (d,  $J$  = 3.9 Hz, 1Ha), 4.41 (d,  $J$  = 5.4 Hz,

1Hb), 4.18 (dd,  $J = 11.7, 6.0$  Hz, 1Hb), 4.12-4.24 (m, 1Ha), 4.07 (dd,  $J = 11.7, 7.2$  Hz, 1Hb), 4.04-4.12 (m, 1Ha), 3.32 (br s, 1Hb), 3.11 (br s, 1Ha), 2.80-2.96 (m, 1Ha+1Hb), 2.53 (dd,  $J = 14.7, 5.4$  Hz, 1Hb), 2.41 (dd,  $J = 14.7, 5.4$  Hz, 1Ha), 2.35 (dd,  $J = 14.4, 6.9$  Hz, 1Hb), 2.34-2.44 (m, 1Ha), 1.55-2.10 (m, 4Ha+4Hb).

**(Z)-2-(3-hydroxy-2-(2-hydroxyethylidene)cyclopentyl)acetamide (18)**

(2 : 1 diastereomeric mixture)  $^1\text{H}$  NMR (200 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  5.42-5.64 (m, 1Ha+1Hb), 4.56-4.64 (m, 1Ha+1Hb), 4.06-4.20 (m, 2Ha+2Hb), 2.78-2.92 (m, 1Ha), 2.61-2.74 (m, 1Hb), 2.43 (dd,  $J = 14.2, 5.8$  Hz, 1Hb), 2.35 (dd,  $J = 14.2, 5.6$  Hz, 1Ha), 2.14 (dd,  $J = 14.2, 9.0$  Hz, 1Hb), 1.97 (dd,  $J = 14.2, 9.6$  Hz, 1Ha), 1.10-2.05 (m, 4Ha+4Hb).

**(Z)-3-(2-amino-2-oxoethyl)-2-(2-(tert-butyldimethylsilyloxy)ethylidene)cyclopentyl benzoate (19)**

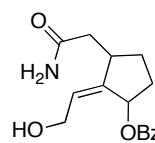


To a solution of **11** (1.62 g, 5.39 mmol) in THF (10 mL) was added pyridine (1.75 mL, 21.6 mmol), DMAP (66 mg, 0.54 mmol), and benzoyl chloride (1.25 mL, 10.8 mmol) at 0 °C. The mixture was stirred for 6 h at room temperature, quenched with sat.  $\text{NH}_4\text{Cl}$ , and extracted with ethyl acetate (x3). The combined organic layers were washed with sat.  $\text{NaHCO}_3$ , dried over anhydrous  $\text{MgSO}_4$ , filtered, concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (elution with hexane/ethyl acetate = 10/1) to give **Bz 12** (2.17 g, quant.) as colorless amorphous (2 : 1 diastereomeric mixture). IR (neat) 2954, 2857, 1790, 1747, 1281  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.09-8.04 (m, 2Ha + 2Hb), 7.46-7.53 (m, 1Ha + 1Hb), 7.34-7.42 (m, 2Ha + 2Hb), 6.11 (br s, 1Ha), 6.02-6.05 (m, 1Hb), 5.97-6.30 (m, 1Ha), 5.89-5.94 (m, 1Hb), 5.57 (t,  $J = 4.8$  Hz, 1Hb), 5.50 (t,  $J = 5.2$  Hz, 1Ha), 3.70-3.85 (m, 2Ha + 2Hb), 2.28-2.58 (m, 3Ha + 3Hb), 2.01 (s, 3Hb), 1.86 (s, 3Ha), 1.82-1.85 (m, 1Ha + 1Hb), 0.81 (s, 9Ha), 0.80 (s, 9Hb), -0.01 (s, 6Ha), -0.04 (s, 6Hb);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  170.1(a), 170.1(b), 166.3(b), 166.1(a), 138.3(b), 138.1(a), 136.3(b), 134.9(a), 134.5(b), 132.8(a), 130.5(b), 130.3(a), 129.5(b), 129.5(a), 128.8(b), 128.2(a), 79.9(a),



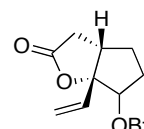


**(Z)-3-(2-amino-2-oxoethyl)-2-(2-hydroxyethylidene)cyclopentyl benzoate (20)**



The mixture of **19** (115 mg, 0.29 mmol) in acetic acid (1.5 mL), H<sub>2</sub>O (0.5 mL), and THF (0.5 mL) was stirred for 4.6 h at room temperature and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (elution with ethyl acetate to ethyl acetate/methanol = 8/1) to give **20** (104 mg, quant.) as colorless amorphous (2 : 1 diastereomeric mixture). IR (neat) 3347, 3198, 2961, 1714, 1452, 1279 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.88-8.01 (m, 2Ha+2Hb), 7.55 (tt, *J* = 7.6, 1.2 Hz, 1Ha+1Hb), 7.42 (td, *J* = 7.6, 1.2 Hz, 2Ha+2Hb), 6.17 (br s, 1Hb), 6.04 (br s, 1Ha), 5.88-5.98 (m, 2Ha+2Hb), 5.77 (tt, *J* = 6.4, 2.0 Hz, 1Ha), 5.69 (tt, *J* = 6.8, 2.0 Hz, 1Hb), 4.14-4.28 (m, 2Ha+2Hb), 3.02-3.18 (m, 1Hb), 2.90-3.0 (m, 1Ha), 2.54 (dd, *J* = 14.4, 6.0 Hz, 1Ha), 2.49 (dd, *J* = 14.4, 6.0 Hz, 1Hb), 2.14-2.40 (m, 1Ha+1Hb), 2.35 (dd, *J* = 14.4, 8.0 Hz, 1Ha), 2.20 (dd, *J* = 14.4, 8.0 Hz, 1Hb), 1.84-2.32 (m, 2Ha+3Hb), 1.72-1.84 (m, 1Hb), 1.60-1.74 (m, 1Ha), 1.39 (dq, *J* = 12.4, 7.6 Hz, 1Ha); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 174.8, 174.7, 166.2, 166.0, 145.0, 144.9, 133.2, 133.2, 130.2, 130.0, 129.6, 129.5, 128.5, 128.4, 128.0, 126.5, 74.6, 74.4, 60.0, 59.6, 41.8, 40.0, 39.8, 39.1, 32.4, 31.4, 29.8, 29.4; HRMS (CI) *m/z* (M+H)<sup>+</sup> calcd for <sup>+</sup> [C<sub>16</sub>H<sub>19</sub>NO<sub>4</sub>+H]<sup>+</sup> 289.1392, found 290.1396.

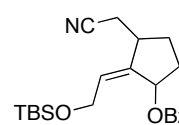
**2-oxo-6a-vinylhexahydro-2H-cyclopenta[b]furan-6-yl benzoate (21)**



To a solution of **19** (14.5 mg, 0.05 mmol) in nitromethane (0.25 mL) was added Hg(OTf)<sub>2</sub> (100 μL, 0.1 M solution in acetonitrile). The mixture was heated to 60 °C for 7 h, quenched with triethylamine (0.2 mL), and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (elution with chloroform) to give **21** (10.9 mg, 80%) as colorless amorphous (1 : 1.5 diastereomeric mixture). IR (neat) 2961, 1787, 1724, 1275, 1117 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ **21a**; 7.96-8.04 (m, 2H), 7.59 (tt, *J* = 7.5, 1.2 Hz, 1H), 7.47 (tt, 8.4, *J* = 1.2 Hz, 2H), 6.07 (dd, *J* = 17.1, 11.1 Hz, 1H), 5.45-5.51 (m, 1H), 5.44 (dd, *J* = 17.1, 0.9 Hz, 1H), 5.25 (dd, *J* = 11.1, 0.9 Hz, 1H), 2.92 (td, *J* = 9.3, 4.5 Hz, 1H), 2.84 (dd, *J* = 17.7, 9.3 Hz, 1H), 2.40 (d, *J* = 17.1 Hz, 1H), 2.20-2.45 (m, 2H), 1.94-2.06 (m, 1H), 1.58-1.72 (m, 1H), **21b**; 8.02-8.10 (m, 2H), 7.57 (tt, *J* = 7.5, 1.8 Hz, 1H), 7.44 (tt, *J* = 7.4, 1.5 Hz, 2H), 5.94 (dd, *J* = 17.4, 10.8 Hz, 1H),

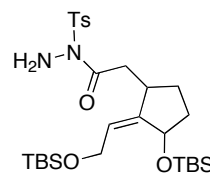
5.43 (dd,  $J = 17.4, 0.6$  Hz, 1H), 5.32 (dd,  $J = 9.3, 5.7$  Hz, 1H), 5.24 (dd,  $J = 10.8, 0.3$  Hz, 1H), 2.90 (dd,  $J = 18.0, 9.3$  Hz, 1H), 2.70-2.80 (m, 1H), 2.38 (dd,  $J = 18.0, 2.1$  Hz, 1H), 1.95-2.28 (m, 3H), 1.61-1.72 (m, 1H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  **21a**; 176.1, 165.0, 133.9, 133.3, 129.8, 129.6, 128.5, 116.2, 95.7, 80.5, 42.2, 35.4, 30.9, 30.8, **21b**; 176.7, 166.0, 135.7, 133.3, 129.9, 129.5, 128.4, 116.5, 93.2, 77.7, 40.7, 36.3, 28.4, 28.3; HRMS (CI)  $m/z$  (M+H) $^+$  calcd for  $^+[\text{C}_{16}\text{H}_{16}\text{O}_4+\text{H}]^+$  273.1127, found 273.1126.

**(Z)-2-(2-(tert-butyl dimethylsilyloxy)ethylidene)-3-(cyanomethyl)cyclopentyl benzoate (1 : 1.5 diastereomeric mixture)**



IR (neat) 2955, 2856, 2247, 1715, 1264, 1109  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.94-8.02 (m, 2Ha+2Hb), 7.49-7.57 (m, 1Ha+1Hb), 7.41 (t,  $J = 7.8$  Hz, 1Ha+1Hb), 5.82-5.95 (m, 1Ha+1Hb), 5.74 (ddt,  $J = 6.9, 4.5, 1.5$  Hz, 1Hb), 5.66 (tt,  $J = 6.0, 2.4$  Hz, 1Ha), 4.40 (ddd,  $J = 13.5, 7.5, 1.5$  Hz, 1Ha), 4.24 (dd,  $J = 6.3, 1.8$  Hz, 2Hb), 4.19 (ddd,  $J = 13.5, 4.5, 2.1$  Hz, 1Ha), 2.92-3.06 (m, 1Ha), 2.76-2.90 (m, 1Hb), 2.64 (dd,  $J = 16.5, 5.4$  Hz, 1Hb), 2.58 (dd,  $J = 16.5, 5.1$  Hz, 1Ha), 2.54 (dd,  $J = 16.5, 7.8$  Hz, 1Hb), 2.39 (dd,  $J = 16.5, 8.1$  Hz, 1Ha), 1.70-2.40 (m, 4Ha+4Hb), 0.82 (s, 9Hb), 0.81 (s, 9Ha), -0.01 (s, 3Ha+3Hb), -0.03 (s, 3Hb), -0.03 (s, 3Ha);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  165.8, 165.7, 133.1, 133.1, 130.6, 130.0, 129.6, 129.6, 129.5, 129.4, 128.4, 128.4, 118.3, 118.3, 74.0, 73.9, 60.8, 60.5, 39.5, 38.9, 32.4, 31.4, 29.5, 29.3, 25.8, 23.0, 21.6, 18.2, -5.3, -5.3; HRMS (FAB)  $m/z$  (M+Na) $^+$  calcd for  $^+[\text{C}_{22}\text{H}_{31}\text{NO}_3\text{Si}+\text{Na}]^+$  408.1971, found 408.1988.

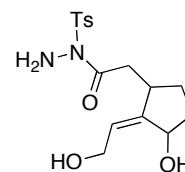
**(Z)-N-(2-(3-(tert-butyl dimethylsilyloxy)-2-(2-(tert-butyl dimethylsilyloxy)ethylidene)cyclopentyl)acetyl)-4-methylbenzenesulfonohydrazide (23)**



To a solution of LHMDS (10.5 mL, 1.6 M solution in THF) in THF (25 mL) was added a solution of TBSCl (3.2 g, 21.3 mmol) in HMPA (14.4 mL, 82.6 mmol) at  $-78$   $^\circ\text{C}$ . The mixture was stirred for 15 min. at  $-78$   $^\circ\text{C}$ . To the mixture was added a solution of **12** (3.8 mg, 9.16 mmol) in THF (5 mL) at  $-78$   $^\circ\text{C}$ . The mixture was stirred for 1 h at  $0$   $^\circ\text{C}$ , quenched with iced water, and extracted with hexane (x2). The combined organic layers were dried over  $\text{MgSO}_4$ , filtered, and concentrated under

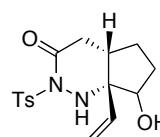
reduced pressure. The solution of residue in toluene (15 mL) was heated to reflux for 50 min, cooled down to room temperature, and concentrated under reduced pressure. To the solution of residue in THF (65 mL) was added H<sub>2</sub>O (10 mL) at room temperature. The mixture was stirred for 3 h, concentrated under reduced pressure to give carboxylic acid **14** which was used without further purification. To a solution of crude **14** in dichloromethane (70 mL) was added *N*-tosylhydrazide (3.3 g, 17.8 mmol), EDCI (4.3 g, 22.2 mmol), and DMAP (362 mg, 3.0 mmol) at room temperature. The mixture was stirred for 1 h, quenched with sat. NH<sub>4</sub>Cl, and extracted with dichloromethane (50 mL x 3). The combined organic layers were washed with brine, dried over anhydrous MgSO<sub>4</sub>, filtered, concentrated under reduce pressure. The residue was purified by flash column chromatography on silica gel (elution with hexane/ethyl acetate = 8/1) to give **23** (3.6 g, 68%) as colorless amorphous (2 : 1 diastereomeric mixture). **23a**: IR (neat) 3363, 2955, 2857, 1703, 1360, 1255, 1167 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.89 (dt, *J* = 8.4, 2.0 Hz, 2H), 7.33 (d, *J* = 8.4 Hz, 2H), 5.29-5.35 (m, 1H), 4.62 (br t, *J* = 6.0 Hz, 1H), 4.42 (s, 2H), 4.33 (ddd, *J* = 13.6, 7.6, 1.6 Hz, 1H), 4.29 (dddd, *J* = 13.6, 4.4, 3.2, 1.2 Hz, 1H), 3.00 (dd, *J* = 15.2, 4.8, 1H), 2.87-2.97 (m, 1H), 2.60 (dd, *J* = 15.6, 9.2 Hz, 1H), 2.44 (s, 3H), 1.91-2.02 (m, 2H), 1.44-1.56 (m, 1H), 1.08-1.16 (m, 1H), 0.90 (s, 9H), 0.86 (s, 9H), 0.09 (s, 3H), 0.07 (s, 3H), 0.06 (s, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 173.8, 146.7, 145.2, 134.9, 129.6, 128.7, 125.0, 72.8, 61.2, 40.0, 38.5, 34.9, 29.3, 26.0, 25.8, 21.7, 18.4, 17.9, -3.8, -4.7, -5.04, -5.07; HRMS (CI) *m/z* (M+H)<sup>+</sup> calcd for <sup>+</sup>[C<sub>28</sub>H<sub>50</sub>N<sub>2</sub>O<sub>5</sub>SSi<sub>2</sub>+H]<sup>+</sup> 583.3057., found 583.3055. **23b**: IR (neat) 3369, 2955, 2857, 1703, 1359, 1169 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 7.89 (dt, *J* = 8.4, 1.6 Hz, 2H), 7.33 (d, *J* = 8.4 Hz, 2H), 5.37 (ddt, *J* = 8.0, 4.0, 1.6 Hz, 1H), 4.61 (t, *J* = 3.2 Hz, 1H), 4.40 (s, 2H), 4.29 (ddd, *J* = 13.2, 8.0, 0.8 Hz, 1H), 4.19 (ddd, *J* = 13.2, 4.0, 1.6 Hz, 1H), 3.03 (d, *J* = 11.2 Hz, 1H), 2.73-2.84 (m, 2H), 2.44 (s, 3H), 1.82 (dtd, *J* = 12.0, 6.8, 4.8 Hz, 1H), 1.64-1.72 (m, 2H), 1.42-1.52 (m, 1H), 0.89 (s, 9H), 0.86 (s, 9H), 0.08 (s, 3H), 0.07 (s, 3H), 0.06 (s, 3H), 0.05 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 174.1, 147.8, 145.4, 135.2, 129.8, 128.9, 125.6, 72.2, 61.5, 42.6, 39.2, 35.3, 29.5, 26.2, 26.1, 21.9, 18.6, 18.1, -3.8, -4.5, -4.8, -4.9; HRMS (CI) *m/z* (M-H)<sup>-</sup> calcd for <sup>+</sup>[C<sub>28</sub>H<sub>50</sub>N<sub>2</sub>O<sub>5</sub>SSi<sub>2</sub>-H]<sup>-</sup> 581.2901, found 581.2900.

***N*-(2-(3-hydroxy-2-(2-hydroxyethylidene)cyclopentyl)acetyl)-4-methylbenzenesulfonohydrazide (**24**)**



A mixture of **23** (3.6 g, 6.25 mmol) in acetic acid (20 mL), H<sub>2</sub>O (20 mL), and THF (20 mL) was stirred for 15 h at room temperature and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (elution with dichloromethane/methanol = 20/1) to give **24** (1.73 g, 78%) as colorless amorphous. **24a**: IR (neat) 3370, 2955, 2871, 1696, 1595, 1354, 1171 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 7.89 (dt, *J* = 8.4, 1.8 Hz, 2H), 7.34 (d, *J* = 8.4 Hz, 2H), 5.53 (tt, *J* = 6.6, 1.8 Hz, 1H), 4.71-4.78 (m, 1H), 4.48 (s, 2H), 4.25 (ddd, *J* = 12.6, 6.9, 1.2 Hz, 1H), 4.08 (dd, *J* = 12.0, 6.6 Hz, 1H), 3.08 (dd, *J* = 16.2, 5.4 Hz, 1H), 2.87 (dd, *J* = 15.9, 7.2 Hz, 1H), 2.74-2.86 (m, 1H), 2.45 (s, 3H), 1.55-1.90 (m, 4H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 173.8, 152.2, 145.4, 134.7, 129.6, 128.8, 124.2, 71.7, 59.8, 40.2, 39.5, 34.1, 29.3, 21.7; HRMS (CI) *m/z* (M-H)<sup>+</sup> calcd for [C<sub>16</sub>H<sub>22</sub>N<sub>2</sub>O<sub>5</sub>S-H]<sup>+</sup> 353.1178., found 353.1180. **24b**: IR (neat) 3362, 2957, 2871, 1695, 1359, 1170 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 7.89 (dt, *J* = 8.4, 1.8 Hz, 2H), 7.34 (d, *J* = 8.4 Hz, 2H), 5.54 (tt, *J* = 6.3, 2.1 Hz, 1H), 4.74-4.83 (m, 1H), 4.43 (s, 2H), 4.26 (ddd, *J* = 13.2, 6.3, 1.5 Hz, 1H), 4.16 (dd, *J* = 13.2, 6.3 Hz, 1H), 2.98-3.10 (m, 1H), 2.93 (dd, *J* = 15.9, 5.7 Hz, 1H), 2.76 (br s, 1H), 2.62 (dd, *J* = 15.9, 8.1 Hz, 1H), 2.45 (s, 3H), 1.95-2.15 (m, 2H), 1.69 (br, s, 1H), 1.53-1.70 (m, 1H), 1.16-1.32 (m, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 173.5, 152.0, 145.4, 134.7, 129.6, 128.7, 123.9, 72.0, 59.8, 40.2, 39.3, 33.8, 29.4, 21.7; HRMS (CI) *m/z* (M-H)<sup>-</sup> calcd for [C<sub>16</sub>H<sub>22</sub>N<sub>2</sub>O<sub>5</sub>S-H]<sup>-</sup> 353.1178., found 353.1189.

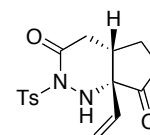
**7-hydroxy-2-tosyl-7a-vinylhexahydro-1*H*-cyclopenta[*c*]pyridazin-3(2*H*)-one (**25**)**



To a solution of **24** (507 mg, 1.43 mmol) in nitromethane (14.3 mL) was added Hg(OTf)<sub>2</sub> (0.29 mL, 0.1 M solution in acetonitrile) at room temperature. The mixture was stirred for 20 min, quenched with triethylamine (10 mL), and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (elution with hexane/ethyl acetate = 1/1) to give **25a** (136 mg, 28%) as colorless crystals and **25b** (270 mg, 56%) as colorless amorphous. **25a**; mp 166 °C (from dichloromethane-diethyl ether); IR (neat) 3492, 3294, 2958, 1715,

1360, 1173  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.97 (d,  $J = 8.4$  Hz, 2H), 7.32 (d,  $J = 8.4$  Hz, 2H), 5.98 (dd,  $J = 17.6, 10.8$  Hz, 1H), 5.28 (d,  $J = 17.6$  Hz, 1H); 5.25 (d,  $J = 10.8$  Hz, 1H), 4.85 (s, 1H), 4.08 (q,  $J = 6.8$  Hz, 1H), 2.81 (d,  $J = 6.8$  Hz, 1H), 2.52-2.61 (m, 2H), 2.49 (dd,  $J = 15.6, 9.2$  Hz, 1H), 2.44 (s, 3H), 1.97-2.08 (m, 1H), 1.93 (dtd,  $J = 13.6, 7.6, 5.6$  Hz, 1H), 1.82 (ddt,  $J = 13.6, 8.0, 5.6$  Hz, 1H), 1.63 (dtd,  $J = 12.4, 7.6, 5.6$  Hz, 1H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  172.0, 145.2, 140.0, 135.2, 129.5, 128.9, 117.0, 78.4, 70.2, 39.3, 32.9, 30.9, 28.3, 21.7; HRMS (CI)  $m/z$  ( $\text{M}+\text{H}$ ) $^+$  calcd for  $[\text{C}_{16}\text{H}_{20}\text{N}_2\text{O}_4\text{S}+\text{H}]^+$  337.1222, found 337.1220. **25 $\beta$** ; IR (neat) 3483, 3282, 2961, 1714, 1359, 1179  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.95 (d,  $J = 8.1$  Hz, 2H), 7.33 (d,  $J = 8.1$  Hz, 2H), 5.97 (dd,  $J = 17.7, 11.1$  Hz, 1H), 5.37 (d,  $J = 11.1$  Hz, 1H); 5.32 (d,  $J = 17.7$  Hz, 1H), 4.48 (s, 1H), 4.05-4.32 (m, 1H), 2.66 (tdd,  $J = 8.7, 8.7, 6.3, 4.5$  Hz, 1H), 2.55 (dd,  $J = 15.3, 6.3$  Hz, 1H), 2.44 (s, 3H), 2.37 (dd,  $J = 15.3, 4.5$  Hz, 1H), 2.32-2.43 (m, 1H), 1.88-2.03 (m, 2H), 1.51 (dddd,  $J = 12.0, 12.0, 10.2, 6.6$  Hz, 1H), 1.19-1.32 (m, 1H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  171.9, 145.2, 137.3, 134.9, 129.4, 128.8, 117.6, 76.9, 70.4, 40.2, 38.0, 30.9, 27.2, 21.7; HRMS (CI)  $m/z$  ( $\text{M}+\text{H}$ ) $^+$  calcd for  $[\text{C}_{16}\text{H}_{20}\text{N}_2\text{O}_4\text{S}+\text{H}]^+$  337.1222, found 337.1213.

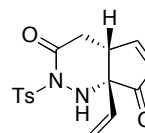
**2-tosyl-7a-vinyltetrahydro-1H-cyclopenta[*c*]pyridazine-3,7(2H,7aH)-dione (26)**



To a solution of the mixture of **25 $\alpha$**  and **25 $\beta$**  (499 mg, 1.48 mmol) in dichloromethane (10 mL) was added DMSO (5 mL, 70.6 mmol), triethylamine (1.24 mL, 8.91 mmol), and a solution of  $\text{SO}_3\cdot\text{Pyr}$  (591 mg, 3.71 mmol) successively at 0  $^\circ\text{C}$ . The mixture was stirred for 1 h at room temperature, quenched with sat.  $\text{NH}_4\text{Cl}$ , extracted with dichloromethane (20 mL x 3). The combined organic layers were washed with brine, dried over anhydrous  $\text{MgSO}_4$ , filtered, concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (elution with hexane/ethyl acetate = 2/1 to 1/1) to give **26** (494 mg, quant.) as colorless amorphous; IR (neat) 3289, 2958, 2254, 1747, 1368, 1175  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.85 (d,  $J = 8.4$  Hz, 2H), 7.28 (d,  $J = 8.4$  Hz, 2H), 5.81 (dd,  $J = 17.4, 10.5$  Hz, 1H), 5.45 (d,  $J = 17.4$  Hz, 1H); 5.32 (d,  $J = 10.5$  Hz, 1H), 5.28 (s, 1H), 2.83 (tt,  $J = 7.8, 5.4$  Hz, 1H), 2.65 (dd,  $J = 15.0, 5.4$  Hz, 1H), 2.48 (dd,  $J = 15.0, 8.1$  Hz, 1H), 2.42 (s, 3H), 2.32-2.45 (m 1H), 2.12-2.34 (m, 2H), 1.45-1.59 (m, 1H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )

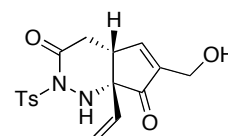
$\delta$  213.6, 171.1, 145.1, 134.7, 134.6, 129.2, 128.9, 118.6, 70.5, 39.0, 37.3, 35.5, 24.0, 21.6; HRMS (CI)  $m/z$  (M)<sup>+</sup> calcd for [C<sub>16</sub>H<sub>18</sub>N<sub>2</sub>O<sub>4</sub>S]<sup>+</sup> 334.0987, found 333.0989.

**2-tosyl-7a-vinyl-4,4a-dihydro-1H-cyclopenta[c]pyridazine-3,7(2H, 7aH)-dione (28)**



A solution of **26** (385 mg, 1.15 mmol) and HMDS (0.48 mL, 2.3 mmol) in dichloromethane (12 mL) was stirred for 30 min at room temperature. To the mixture was added Trimethylsilyliodide (0.25 mL, 1.70 mmol) at 0 °C. The mixture was stirred for 1 h at 0 °C, stirred for 15 min. at room temperature, quenched with sat. NaHCO<sub>3</sub>, extracted with dichloromethane (20 mL x 2). The combined organic layers were dried over anhydrous MgSO<sub>4</sub>, filtered, concentrated under reduced pressure. To a solution of residue in DMSO (12 mL) was added Pd(OAc)<sub>2</sub> (258 mg, 1.15 mmol) was added at room temperature. The mixture was stirred for 4 h, quenched with H<sub>2</sub>O, extracted with ethyl acetate. The combined organic layers were washed with brine, dried over anhydrous MgSO<sub>4</sub>, filtered, concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (elution with hexane/ethyl acetate = 2/3) to give **28** (298 mg, 78% from **26**) as white solid. mp 182 °C (from dichloromethane-diethyl ether); IR (neat) 3243, 2924, 1715, 1354, 1167 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.89 (d,  $J$  = 8.4 Hz, 2H), 7.50 (dd,  $J$  = 6.0, 2.8 Hz, 1H), 7.30 (d,  $J$  = 8.4 Hz, 2H), 6.09 (dd,  $J$  = 6.0, 2.0 Hz, 1H), 5.89 (dd,  $J$  = 17.6, 10.8 Hz, 1H), 5.47 (d,  $J$  = 17.6 Hz, 1H); 5.46 (s, 1H), 5.40 (d,  $J$  = 10.8 Hz, 1H), 3.48 (dddd,  $J$  = 6.8, 4.8, 2.8, 2.0 Hz, 1H), 2.94 (dd,  $J$  = 14.4, 6.8 Hz, 1H), 2.55 (dd,  $J$  = 14.4, 4.8 Hz, 1H), 2.43 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  204.6, 168.3, 163.4, 145.5, 134.6, 134.4, 133.2, 129.4, 129.3, 119.6, 68.1, 46.4, 36.2, 21.8; HRMS (CI)  $m/z$  (M+H)<sup>+</sup> calcd for [C<sub>16</sub>H<sub>16</sub>N<sub>2</sub>O<sub>4</sub>S+H]<sup>+</sup> 333.0909, found 333.0904.

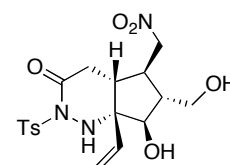
**6-(hydroxymethyl)-2-tosyl-7a-vinyl-4,4a-dihydro-1H-cyclopenta[c]pyridazine-3,7(2H,7aH)-dione (29)**



To a solution of **28** (289 mg, 0.87 mmol) in THF (9 mL) was added formaldehyde (0.35 mL, 4.34 mmol, 37% water solution) and *n*-tributylphosphine (87  $\mu$ L, 0.35 mmol) at room temperature. The mixture was stirred

for 2 h, quenched with H<sub>2</sub>O, extracted with ethyl acetate (x 3). The combined organic layers were dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (elution with hexane/ethyl acetate = 1/4) to give **29** (289 mg, 92%) as colorless amorphous; IR (neat) 3460, 3208, 2979, 2595, 1709, 1357 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CD<sub>3</sub>OD) δ 7.71 (d, *J* = 8.4 Hz, 2H), 7.36 (dt, *J* = 2.7, 1.8 Hz, 1H), 7.27 (d, *J* = 8.4 Hz, 2H), 5.78 (dd, *J* = 17.4, 10.8 Hz, 1H), 5.37 (d, *J* = 17.4 Hz, 1H), 5.22 (d, *J* = 10.8 Hz, 1H), 4.00 (dt, *J* = 15.3, 1.8 Hz, 1H), 3.83 (br d, *J* = 15.3 Hz, 1H), 3.40-3.47 (m, 1H), 2.89 (dd, *J* = 15.3, 7.2 Hz, 1H), 2.45 (dd, *J* = 15.3, 3.6 Hz, 1H), 2.33 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 204.9, 172.7, 159.6, 148.1, 147.0, 137.1, 135.5, 130.7, 130.4, 119.4, 71.2, 57.5, 46.1, 37.1, 21.9; HRMS (CI) *m/z* (M+H)<sup>+</sup> calcd for [C<sub>17</sub>H<sub>18</sub>N<sub>2</sub>O<sub>5</sub>S+H]<sup>+</sup> 363.1014, found 363.1008

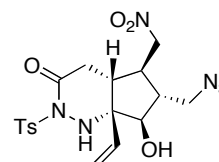
**7-hydroxy-6-(hydroxymethyl)-5-(nitromethyl)-2-tosyl-7a-vinyl hexahydro-1*H*-cyclopenta[*c*]pyridazin-3(2*H*)-one (**31**)**



To a solution of **29** (229 mg, 0.63 mmol) in nitromethane (12.6 mL) was added tetramethylguanidine (33 μL, 0.27 mmol) at 0 °C. The mixture was stirred for 3.5 h, quenched with sat. 1 M HCl, and extracted with ethyl acetate (30 mL x 3). The combined organic layers were washed with sat. NaHCO<sub>3</sub> and brine, dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure to give crude **30**. The solution of crude **30** in methanol (5 mL) was added a solution of NaBH<sub>4</sub> (19.0 mg, 0.5 mmol) in methanol (1.3 mL) at -78 °C. The mixture was stirred for 30 min, quenched with sat. NH<sub>4</sub>Cl, and extracted with ethyl acetate (20 mL x 3). The combined organic layers were dried over anhydrous MgSO<sub>4</sub>, filtered, concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (elution with dichloromethane/methanol = 20/1) to give **31** (132 mg, 49% from **29**) as colorless amorphous. IR (neat) 3487, 3289, 2922, 2253, 1715, 1550, 1361, 1169 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.89 (d, *J* = 8.4 Hz, 2H), 7.31 (d, *J* = 8.4 Hz, 2H), 5.97 (dd, *J* = 17.6, 10.8 Hz, 1H), 5.33 (d, *J* = 10.8 Hz, 1H), 5.25 (d, *J* = 17.6 Hz, 1H), 5.03 (s, 1H), 4.62 (dd, *J* = 12.4, 4.0 Hz, 1H), 4.43 (dd, *J* = 12.4, 7.2 Hz, 1H), 4.08 (d, *J* = 10.8 Hz, 1H), 3.96 (br s, 1H), 3.82 (br d, *J* = 9.2 Hz, 1H), 3.76 (br d, *J* = 9.2 Hz, 1H), 3.30 (br s, 1H), 2.52-2.65 (m, 2H), 2.49 (dd, *J* = 16.8, 8.0 Hz, 1H), 2.42 (s, 3H), 2.18-2.32 (m,

1H), 1.65-1.76 (m, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 171.3, 145.8, 136.6, 134.4, 129.6, 129.0, 118.7, 76.6, 75.4, 69.4, 59.5, 46.6, 42.2, 39.3, 36.8, 21.7; HRMS (CI) m/z (M+H)<sup>+</sup> calcd for [C<sub>18</sub>H<sub>23</sub>N<sub>3</sub>O<sub>7</sub>S+H]<sup>+</sup> 426.1335, found 426.1338.

**6-(azidomethyl)-7-hydroxy-5-(nitromethyl)-2-tosyl-7a-vinylhexahydro-1H-cyclopenta[c]pyridazin-3(2H)-one (32)**



To a solution of **31** (22.4 mg, 0.053 mmol) in pyridine (0.27 mL) was added methanesulfonylchloride (9.0 mg, 0.079 mmol) at 0 °C.

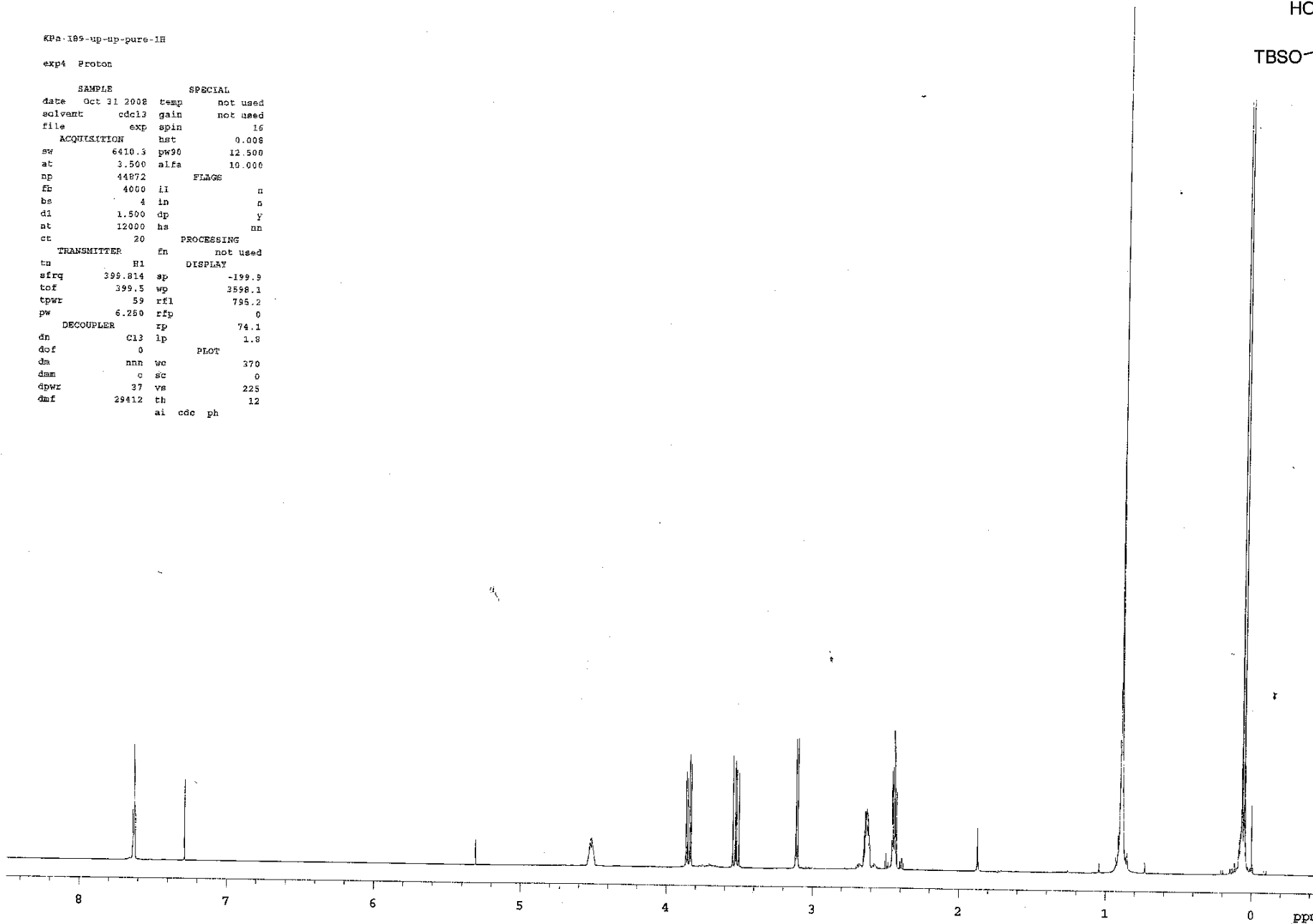
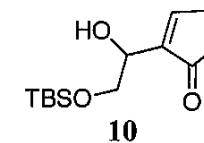
The mixture was stirred for 45 min at room temperature, quenched with sat. NaHCO<sub>3</sub>, and extracted with ethyl acetate (20 mL x 2). The combined organic layers were dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure. The residue was filtered through silica gel pad (elution with dichloromethane/methanol = 9/1) and concentrated under reduced pressure. The solution of residue in DMF (0.57 mL) was added sodium azide (7.2 mg, 0.11 mmol) at room temperature. The mixture was heated to 60 °C for 3 h, diluted with ethyl acetate (5 mL), filtered, and washed with sat. NaHCO<sub>3</sub>. The aqueous layer was extracted with ethyl acetate (20 mL x 3). The combined organic layers were dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure. The residue was purified by flash column chromatography of silica gel (elution with hexane/ethyl acetate = 1/1) to give **32** (16 mg, 67% from **31**) as colorless crystal. mp 151 °C (from dichloromethane-diethyl ether); IR (neat) 3489, 3289, 2924, 2104, 1715, 1555, 1366, 1276, 1174cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 7.94 (d, J = 8.1 Hz, 2H), 7.35 (d, J = 8.1 Hz, 2H), 5.96 (dd, J = 17.4, 11.1 Hz, 1H), 5.43 (d, J = 11.1 Hz, 1H), 5.33 (d, J = 17.4 Hz, 1H), 4.66 (s, 1H), 4.58 (dd, J = 12.3, 4.2 Hz, 1H), 4.41 (dd, J = 12.3, 6.6 Hz, 1H), 3.85 (d, J = 10.8 Hz, 1H), 3.63 (dd, J = 12.9, 3.3 Hz, 1H), 3.54 (dd, J = 12.9, 5.1 Hz, 1H), 2.57-2.69 (m, 2H), 2.45 (s, 3H), 2.42-2.48 (m, 1H), 1.95-2.10 (m, 1H), 1.74 (tdd, J = 11.4, 5.1, 3.6 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 170.5, 145.8, 135.8, 134.4, 129.6, 129.0, 118.8, 75.9, 75.1, 68.7, 49.5, 43.7, 42.1, 40.2, 36.3, 21.8; HRMS (CI) m/z (M+H)<sup>+</sup> calcd for [C<sub>18</sub>H<sub>22</sub>N<sub>6</sub>O<sub>6</sub>S+H]<sup>+</sup> 451.1400, found 451.1403.



KPa-189-up-up-pure-1H

exp4 Proton

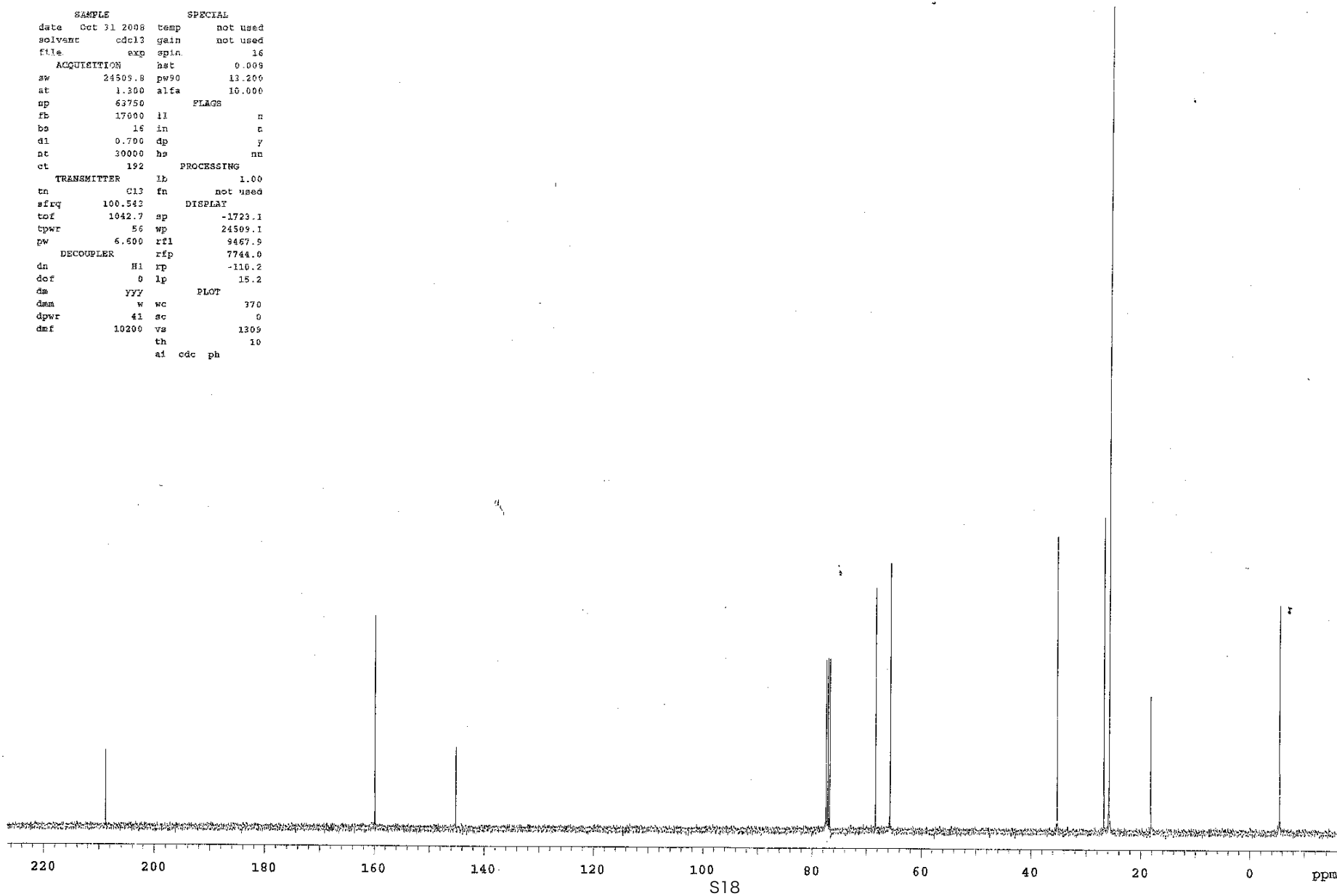
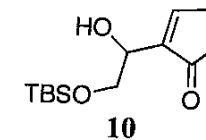
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date	Oct 31 2008	Temp	not used
solvent	cdcl3	gain	not used
file	exp	spin	16
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at	3.500	alfa	10.000
np	44872	FLAGE	
fb	4000	li	n
bs	4	in	n
dl	1.500	dp	y
nt	12000	hs	nn
ct	20	PROCESSING	
TRANSMITTER		fn	not used
tn	H1	DISPLAY	
sfrq	399.814	sp	-199.9
tof	399.5	wp	3598.1
tpwr	59	rfl	795.2
pw	6.250	rfp	0
DECOUPLER		rp	74.1
dn	C13	lp	1.8
dof	0	PLOT	
dn	nnn	vc	370
dmm	c	sc	0
gpwr	37	vs	225
dmf	29412	th	12
		ai	cdc ph



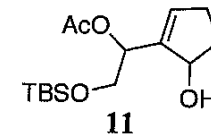
KPa 18% up-up-pure-13C

exp5 Carbon

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solvent	cdcl3	gain	not used
file	exp	spin	16
ACQUISITION		hst	0.009
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at	1.300	alfa	10.000
ap	63750	PLAGS	
fb	17600	il	n
bs	16	in	n
dl	0.700	dp	y
nt	30000	hs	nn
ct	192	PROCESSING	
TRANSMITTER		lb	1.00
tn	013	fn	not used
sfrq	100.542	DISPLAY	
tof	1042.7	sp	-1723.1
tpwr	56	wp	24509.1
pw	6.600	rfl	9467.6
DECOUPLER		rflp	7744.0
dn	H1	rp	-110.2
dof	0	lp	15.2
dm	YYY	ELOT	
dmm	w	wc	770
dpwr	41	sc	0
dmf	10200	va	1309
		th	10
	al	cdc	ph







KPa-373-down-13C

Pulse Sequence: s2pul

Solvent: CDCl3

Ambient temperature

File: KPa-373-down-13C-102008\_23\_24

INOVA-600 "NMR"

Pulse 54.2 degrees

Acq. time 1.815 sec

Width 18761.7 Hz

288 repetitions

OBSERVE C13, 75.4279239 MHz

DECOUPLE H1, 299.9730594 MHz

Power 39 dB

continuously on

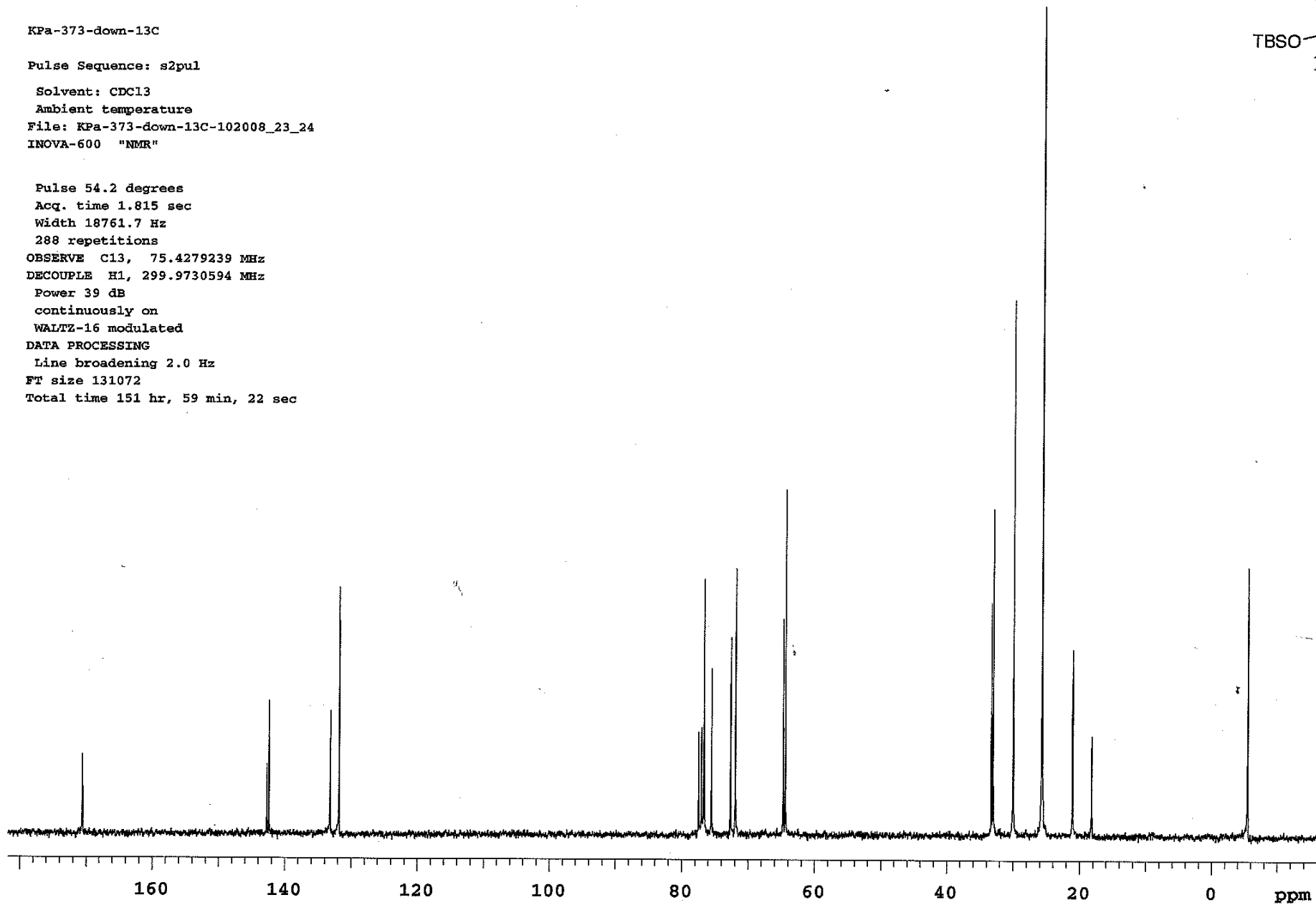
WALTZ-16 modulated

DATA PROCESSING

Line broadening 2.0 Hz

FT size 131072

Total time 151 hr, 59 min, 22 sec

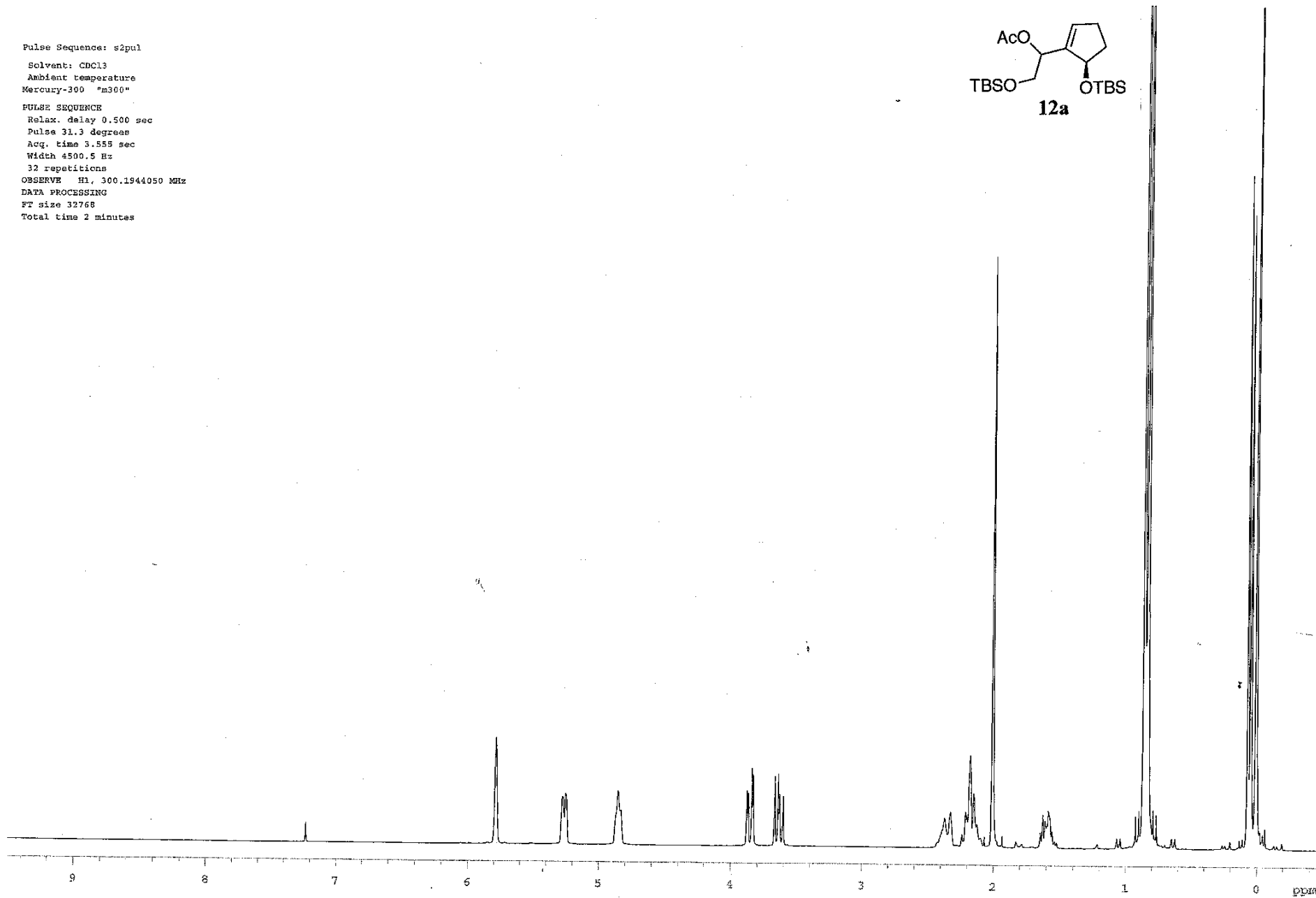
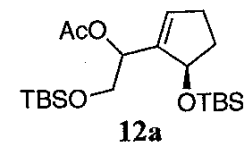


Pulse Sequence: s2pul

Solvent: CDCl3  
Ambient temperature  
Mercury-300 "m300"

PULSE SEQUENCE  
Relax. delay 0.500 sec  
Pulse 31.3 degrees  
Acq. time 3.555 sec  
Width 4500.5 Hz  
32 repetitions

OBSERVE H1, 300.1944050 MHz  
DATA PROCESSING  
FT size 32768  
Total time 2 minutes



13C OBSERVE

Pulse Sequence: n2pul

Solvent: CDCl3  
Ambient temperature  
Mercury-300 "m300"

PULSER SEQUENCE

Relax. delay 0.294 sec  
Pulse 40.0 degrees  
Acq. time 1.705 sec  
Width 18863.2 Hz  
464 repetitions

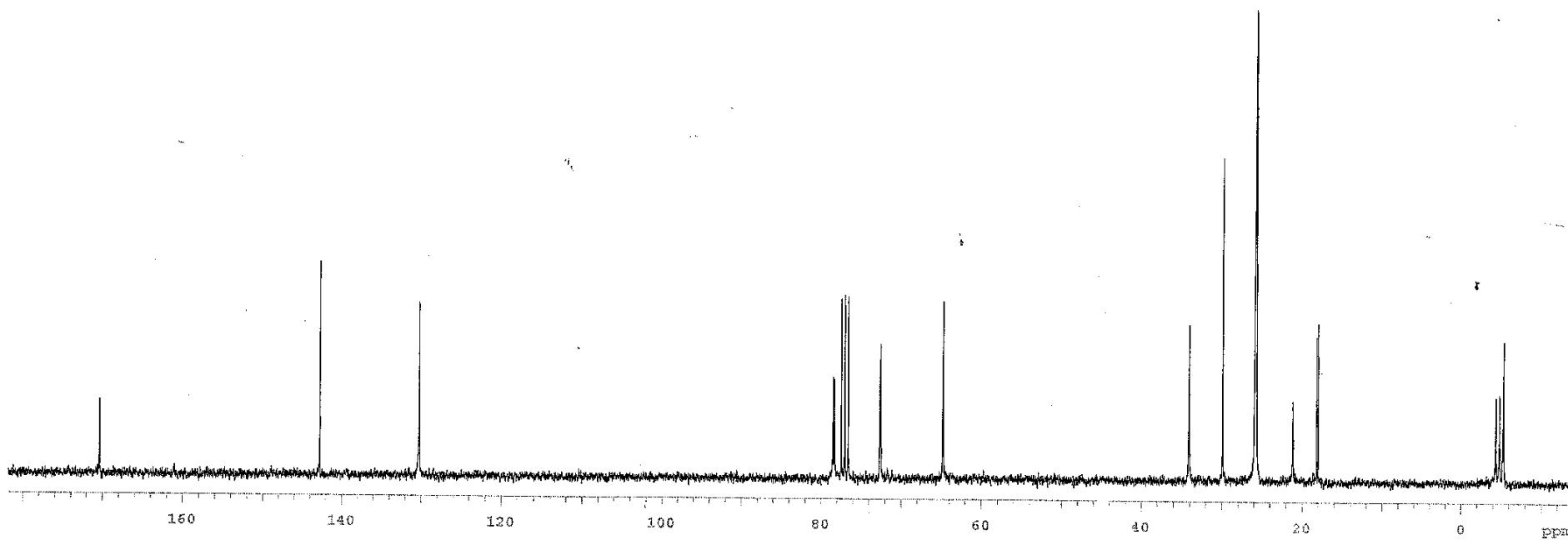
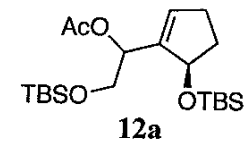
OBSERVE C13, 75.4839403 MHz  
DECOUPLE H1, 300.1958432 MHz

Power 38 dB  
continuously on  
WALTZ-16 modulated

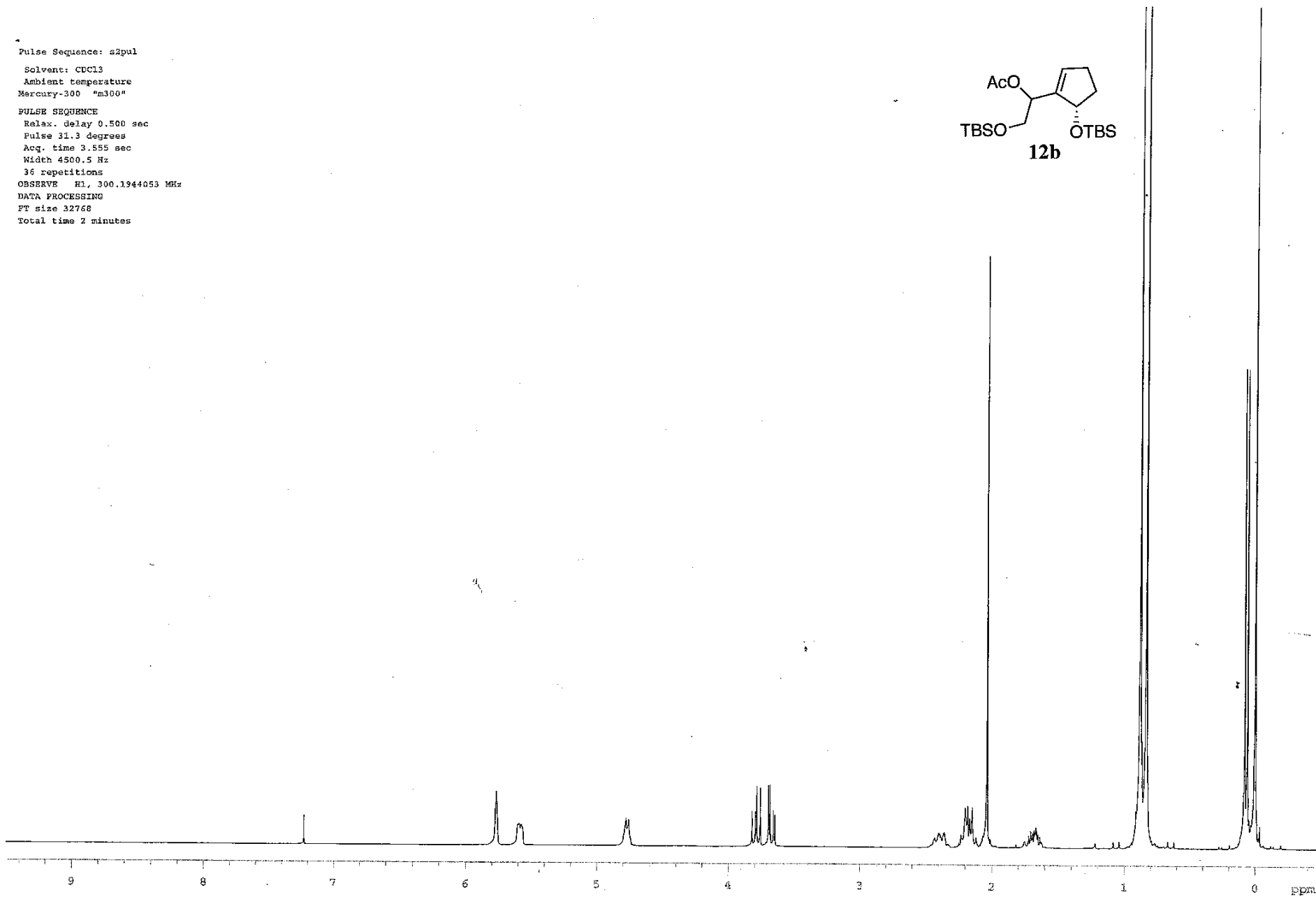
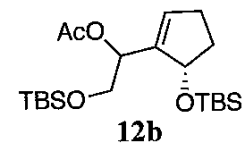
DATA PROCESSING

Line broadening 1.0 Hz  
FT size 65536

Total time 15 minutes



Pulse Sequence: s2pul  
Solvent: CDCl3  
Ambient temperature  
Mercury-300 "m300"  
PULSE SEQUENCE  
Relax. delay 0.500 sec  
Pulse 31.3 degrees  
Acq. time 3.555 sec  
Width 4500.5 Hz  
36 repetitions  
OBSERVE H1, 300.1944053 MHz  
DATA PROCESSING  
FT size 32768  
Total time 2 minutes



13C OBSERVE

Pulse Sequence: s2pul

Solvent: CDCl3  
Ambient temperature  
Mercury-300 "m300"

PULSE SEQUENCE

Relax. delay 0.294 sec

Pulse 46.0 degrees

Acq. time 1.705 sec

Width 18863.2 Hz

280 repetitions

OBSERVE C13, 75.4839403 MHz

DECOUPLE H1, 300.1958432 MHz

Power 38 dB

continuously on

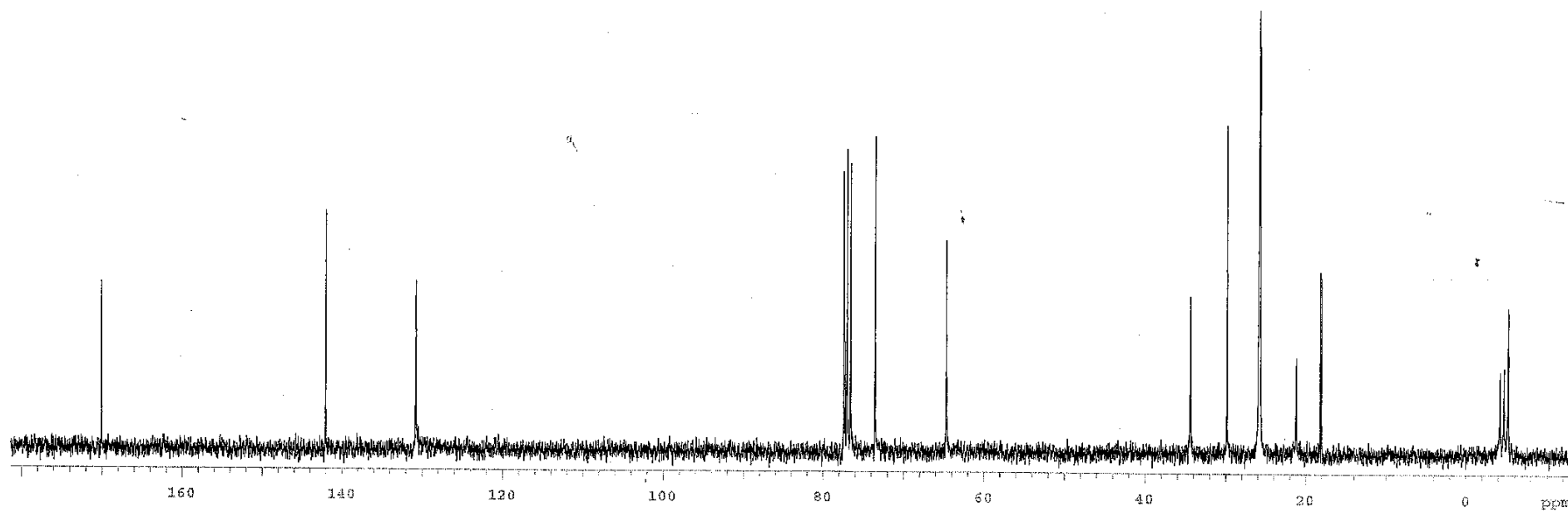
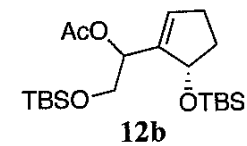
WALTZ-16 modulated

DATA PROCESSING

Line broadening 1.0 Hz

FT size 65536

Total time 9 minutes

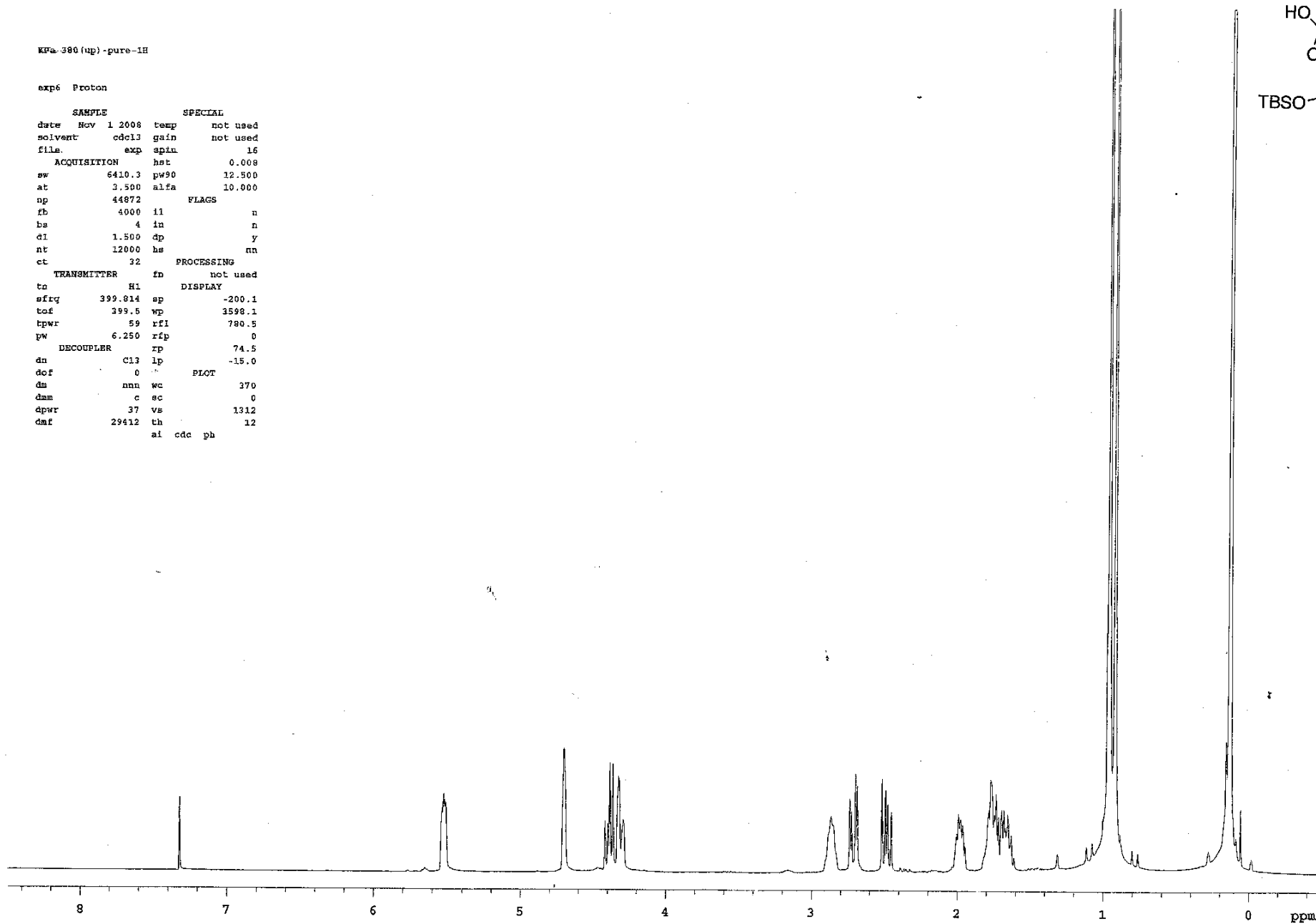
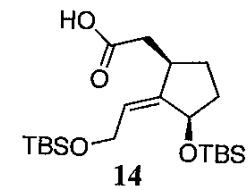




KPa-380 (up) -pure-1H

exp6 Proton

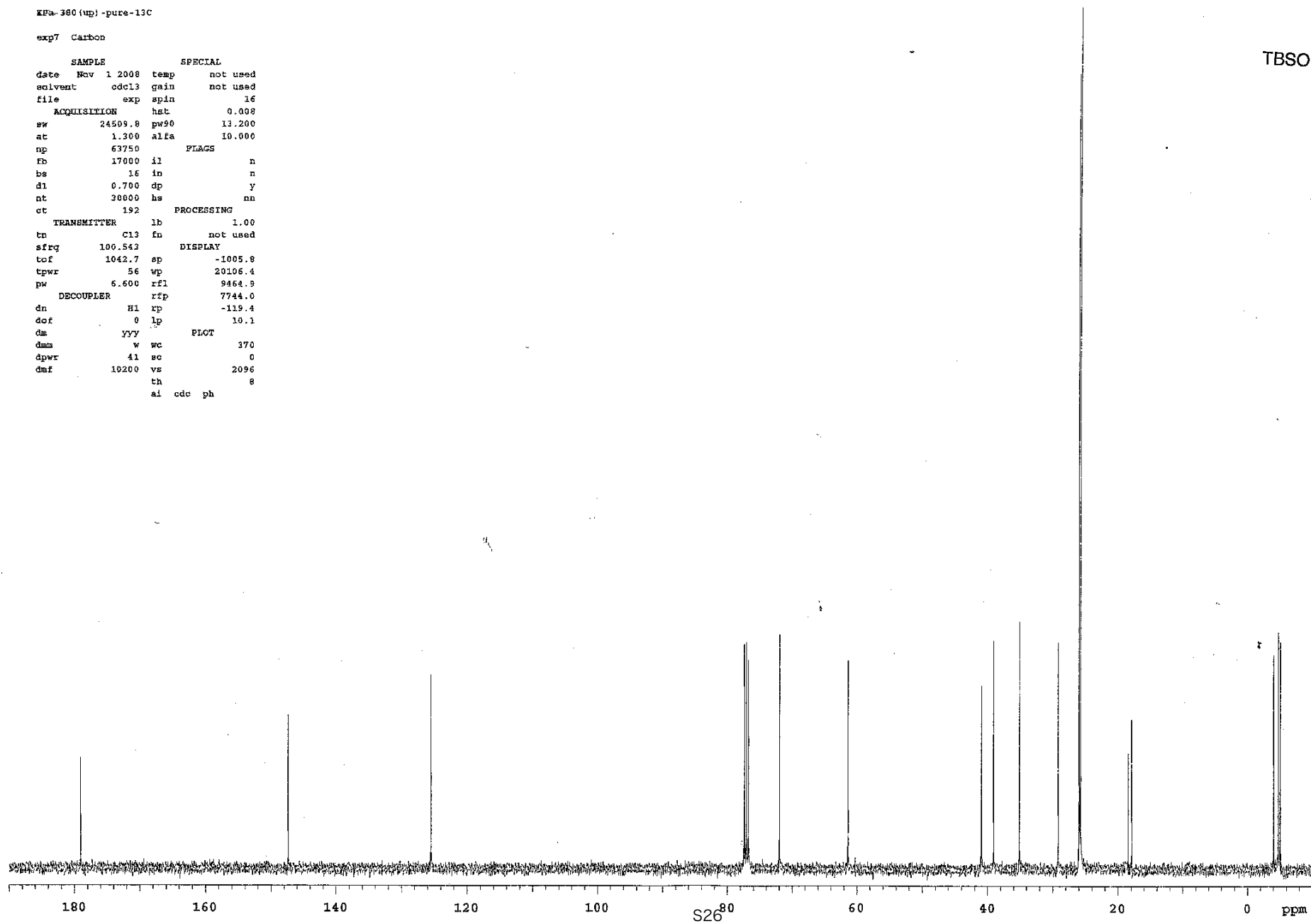
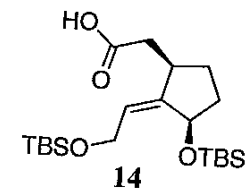
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at	3.500	alfa	10.000
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zb	4000	l1	n
ba	4	in	n
cl	1.500	dp	y
nt	12000	hs	nn
ct	32	PROCESSING	
TRANSMITTER		fn	not used
ta	H1	DISPLAY	
sfrq	399.814	ap	-200.1
tof	399.5	wp	1598.1
tpwr	59	rfl	780.5
pw	6.250	rfp	0
DECOUPLER		zp	74.5
dn	C13	lp	-15.0
dof	0	PLOT	
dm	nnn	wc	370
dmm	c	ec	0
dpwr	37	vs	131.2
dmf	29412	th	12
		ai	cdc ph



KPa-380 (up) -pure-13C

exp7 Carbon

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ACQUISITION		hst	0.008
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op	63750	FLAGS	
fb	17000	il	n
bs	16	in	n
dl	0.700	dp	y
nt	30000	hs	nn
ct	192	PROCESSING	
TRANSMITTER		lb	1.00
tn	C13	fn	not used
sfrq	100.542	DISPLAY	
tof	1042.7	ep	-1005.9
tpwr	56	vp	20106.4
pw	6.600	rfl	9464.9
DECOUPLER		rfp	7744.0
dn	H1	rp	-119.4
dof	0	lp	10.1
dm	yyy	PLOT	
dms	w	wc	370
dpwr	41	sc	0
dmi	10200	vs	2096
		th	8
		ai	cdc ph



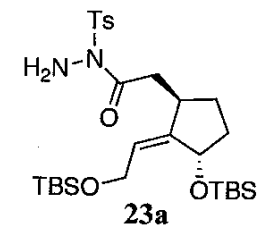
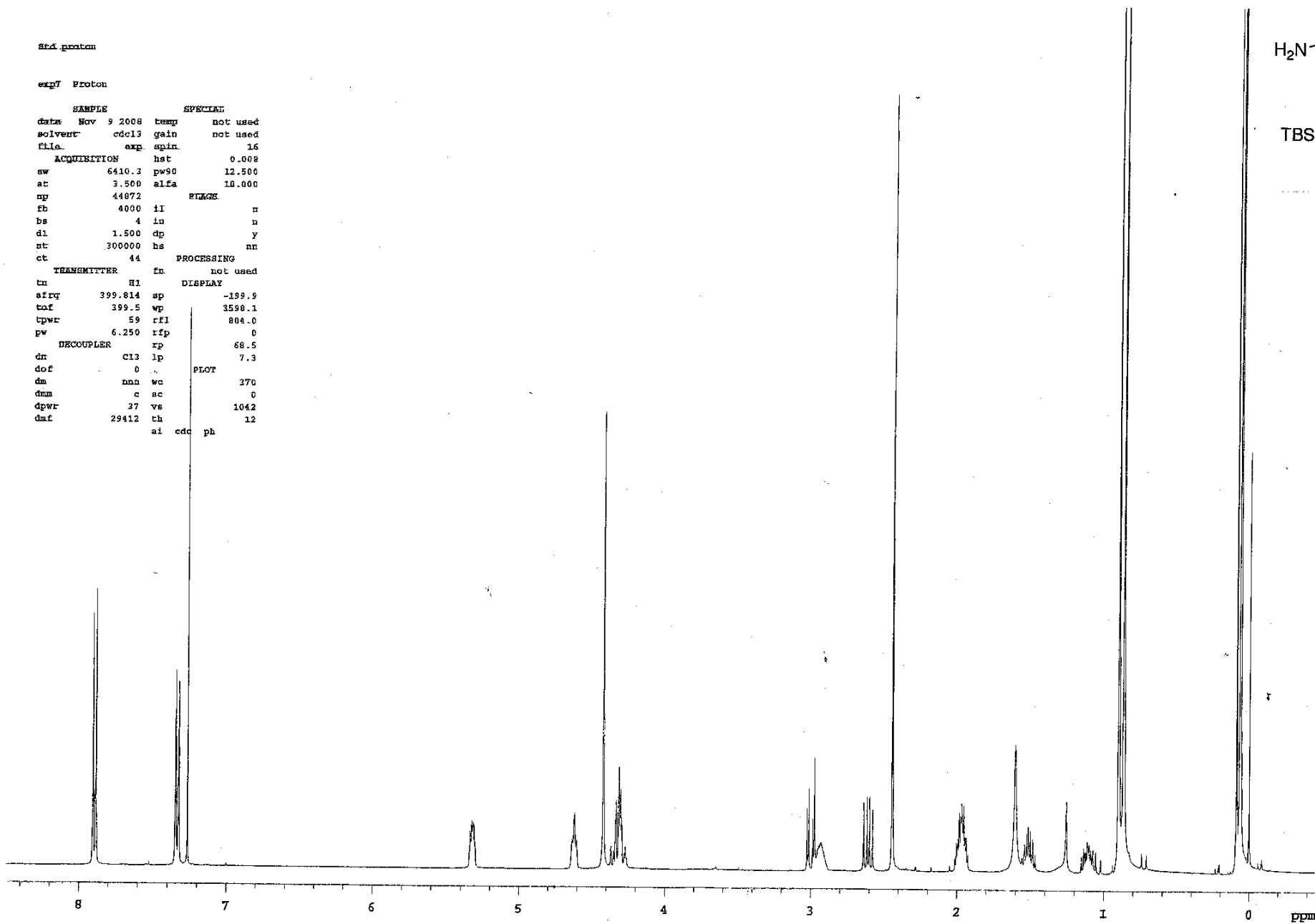




Ref. proton

exp7 Proton

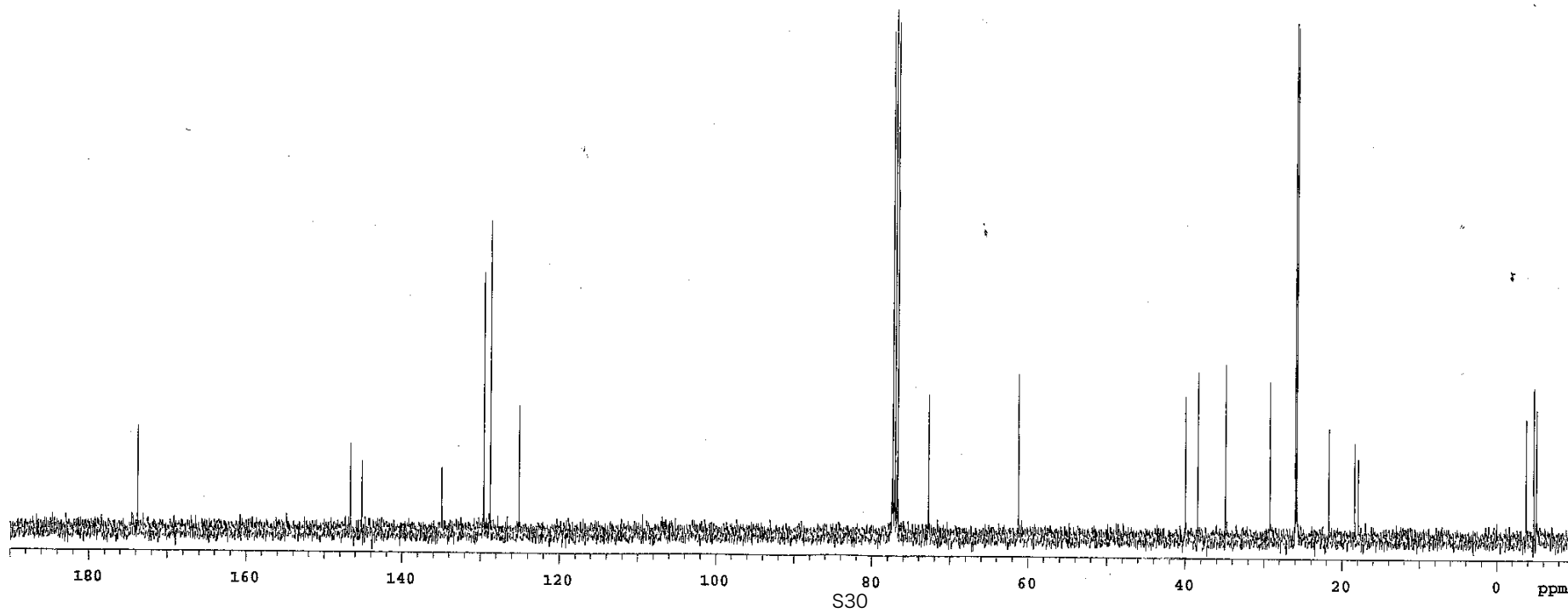
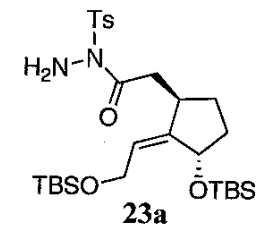
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date	Nov 9 2008	temp	not used
solvent	cdcl3	gain	not used
file	exp	spin	16
ACQUISITION		ht	0.008
sw	6410.3	pw90	12.500
at	3.500	alfa	10.000
ny	44872	PULSES	
fb	4000	ii	n
bs	4	in	n
dl	1.500	dp	y
nt	300000	hs	nn
ct	44	PROCESSING	
TRANSMITTER		fn	not used
tn	q1	DISPLAY	
afrq	399.814	sp	-199.9
taf	399.5	wp	3558.1
tpwr	59	rfl	804.0
pw	6.250	rfp	0
DECOUPLER		rp	68.5
dr	c13	lp	7.3
dof	0	PLOT	
dm	nan	wc	270
dmm	c	ac	0
dpwr	37	vs	1042
daf	29412	th	12
	ai	cdc	ph



RF-386-down-down

exp5 Carbon

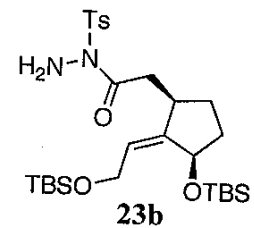
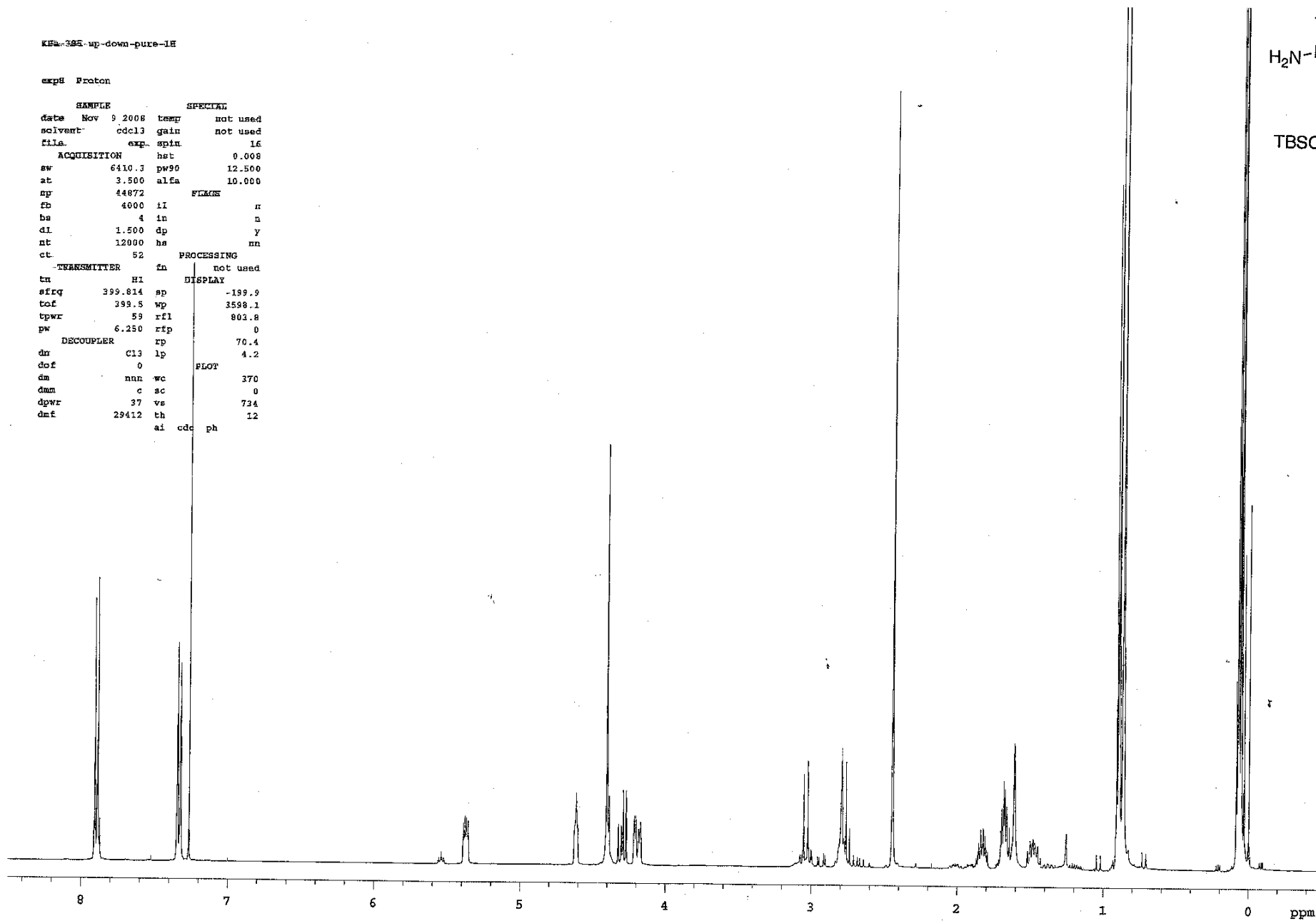
SAMPLE		SPECIAL	
date	Nov 7 2008	temp	not used
solvent	cdcl3	gain	not used
file	exp	spin	16
ACQUISITION		hst	0.008
sw	24509.8	pw90	13.200
at	1.300	alfa	10.000
ap	63750	FLAGE	
fb	17600	ii	r
ba	16	in	n
dl	0.700	dp	y
nt	30000	hs	nn
ct	272	PROCESSING	
TRANSMITTER		lb	1.00
tn	C13	fn	not used
sfrq	100.543	DISPLAY	
tof	1042.7	sp	-1005.8
tpwl	56	wp	20106.4
pw	6.600	rfl	9465.7
DECOUPLER		rEp	7744.0
dx	H1	rp	-135.2
dot	0	ip	62.5
dm	yyy	FLGT	
dum	w	wc	370
dpwr	41	sc	0
dmf	10200	va	3682
		th	11
	ai	cdc	ph



KEA-325-up-down-pure-1H

expH Proton

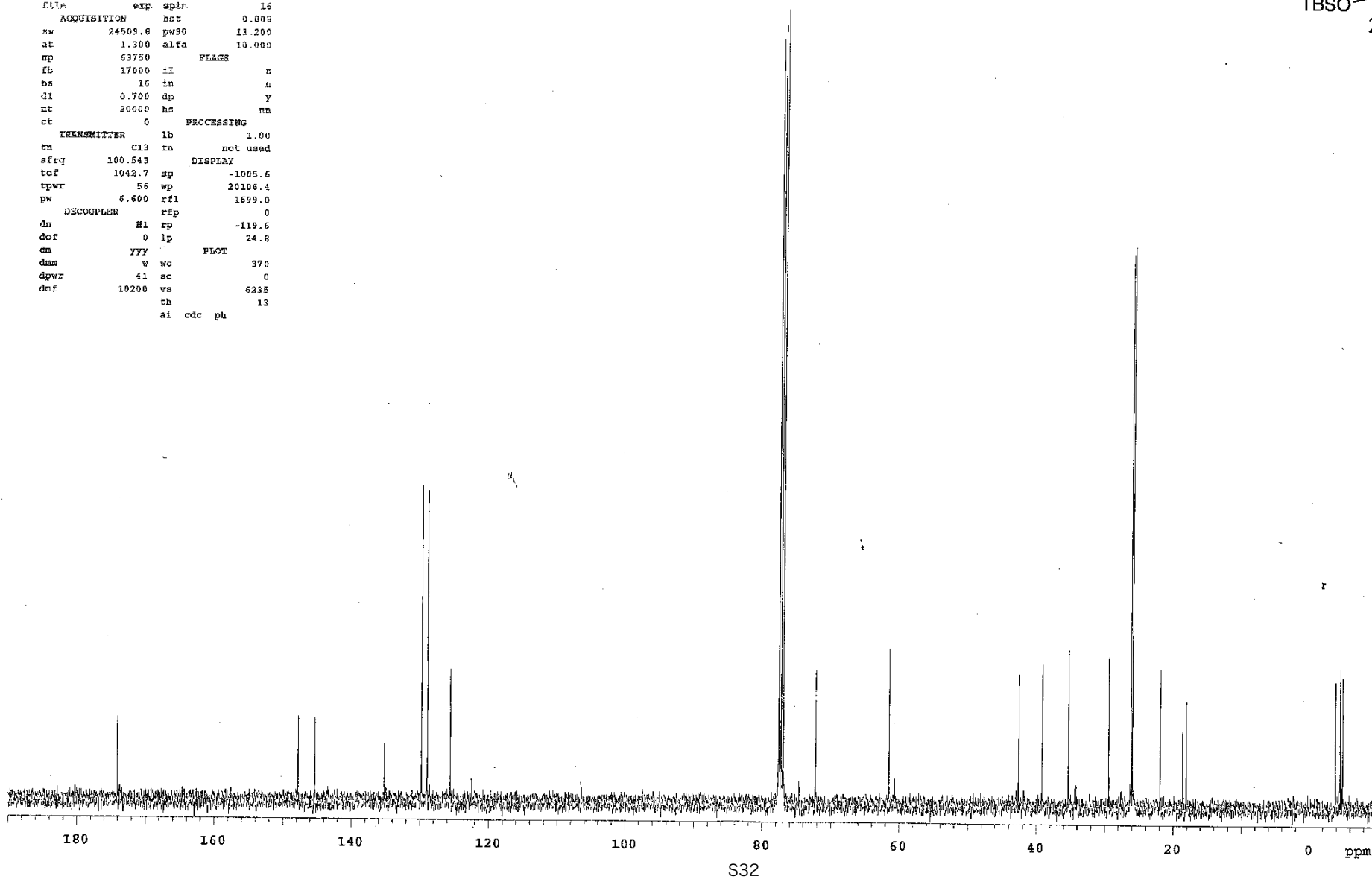
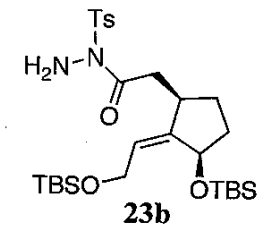
SAMPLE		SPECIAL	
date	Nov 9 2008	temp	not used
solvent	cdcl3	gain	not used
file	exp	spin	1f
ACQUISITION		hst	0.008
sw	6410.3	pw90	12.500
at	3.500	alfa	10.000
ny	44872	FNAME	
fb	4000	ii	n
ba	4	in	n
dl	1.500	dp	y
nt	12000	hs	nn
ct	52	PROCESSING	
TRANSMITTER		fn	not used
tn	H1	DISPLAY	
afiq	399.814	sp	-199.9
tof	399.5	wp	3598.1
tpwr	59	rfl	803.8
pw	6.250	rtp	0
DECOUPLER		rp	70.4
dn	C13	lp	4.2
dof	0	FLOT	
dm	nan	wc	370
dmm	c	ac	0
dpwr	37	vs	734
dmf	29412	th	12
	ai	cdc	ph



KW 185 up-down

exp7 Carbon

SAMPLE		SPECIAL	
date	Nov 7 2008	temp	not used
solvent	cdcl3	gain	not used
file		exp	spin 16
ACQUISITION		hst	0.008
sw	24509.8	pw90	11.200
at	1.300	alfa	10.000
np	53750	FLAGS	
fb	17500	ti	n
bs	16	in	n
dl	0.700	dp	y
nt	30000	hs	nn
ct	0	PROCESSING	
TRANSMITTER		lb	1.00
tn	Cl3	fn	not used
sfrq	100.543	DISPLAY	
tof	1042.7	sp	-1005.6
tpwr	56	wp	20106.4
pw	6.600	rfl	1699.0
DECOUPLER		rfp	0
dn	H1	rp	-119.6
dof	0	lp	24.8
dm	YYY	PLOT	
dum	w	wc	370
dpwr	41	sc	0
dmf	10200	vs	6235
		th	13
		ai	cdc ph

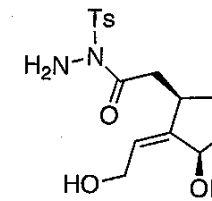




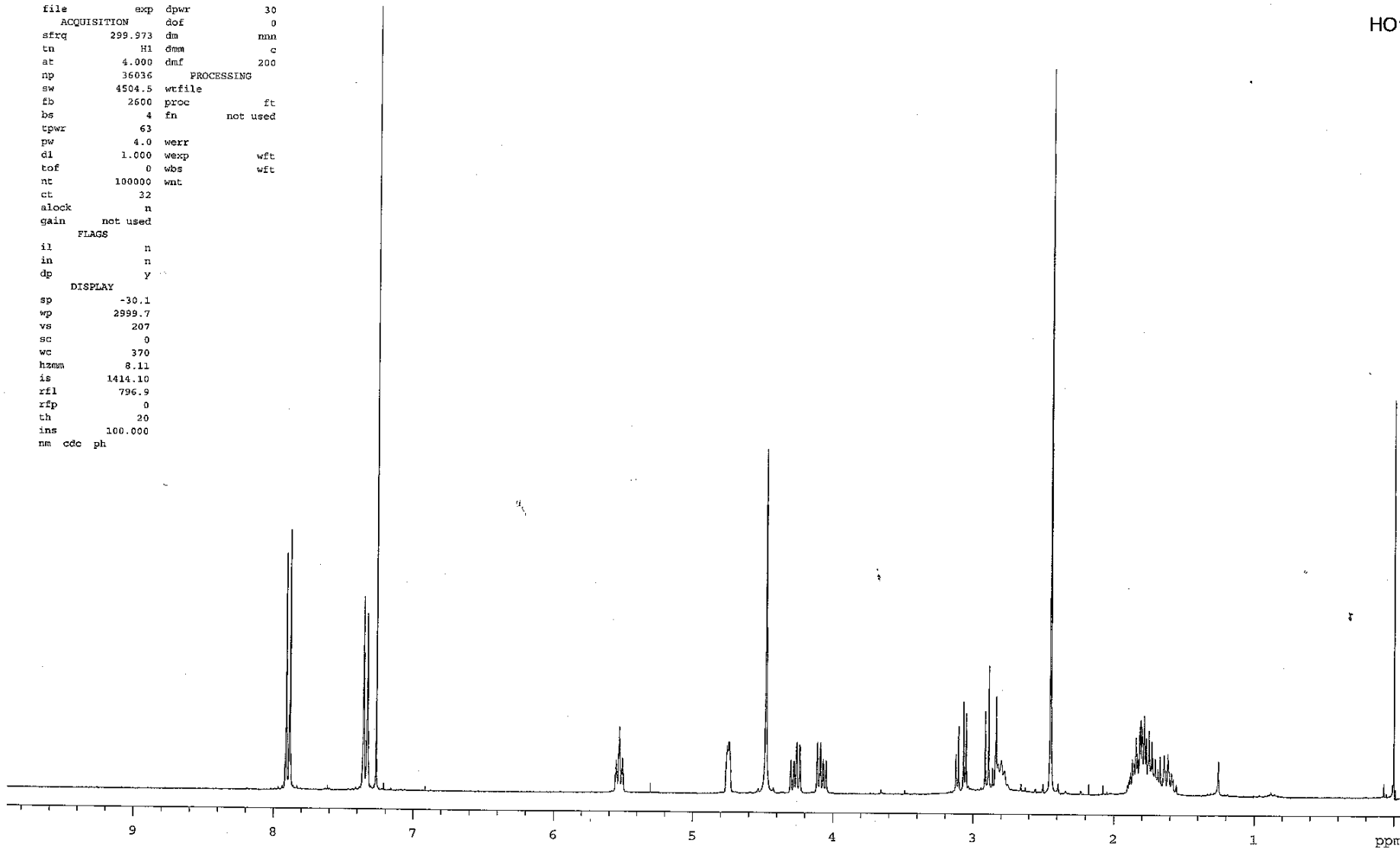
KPa-388-up-major-H

exp7 std1h

SAMPLE		DEC. & VT	
date	Nov 11 2008	dfrq	299.973
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		doF	
sfrq	299.973	dm	mm
tn	H1	dmm	c
at	4.000	dmf	200
np	36036	PROCESSING	
sw	4504.5	wtfile	
fb	2600	proc	ft
bs	4	fn	not used
tpwr	63		
pw	4.0	werr	
d1	1.000	wexp	wft
tof	0	wbs	wft
nt	100000	wat	
ct	32		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-30.1		
wp	2999.7		
vs	207		
sc	0		
wc	370		
hzmm	8.11		
is	1414.10		
rfl	796.9		
rfp	0		
th	20		
ins	100.000		
nm	cdc ph		



24a



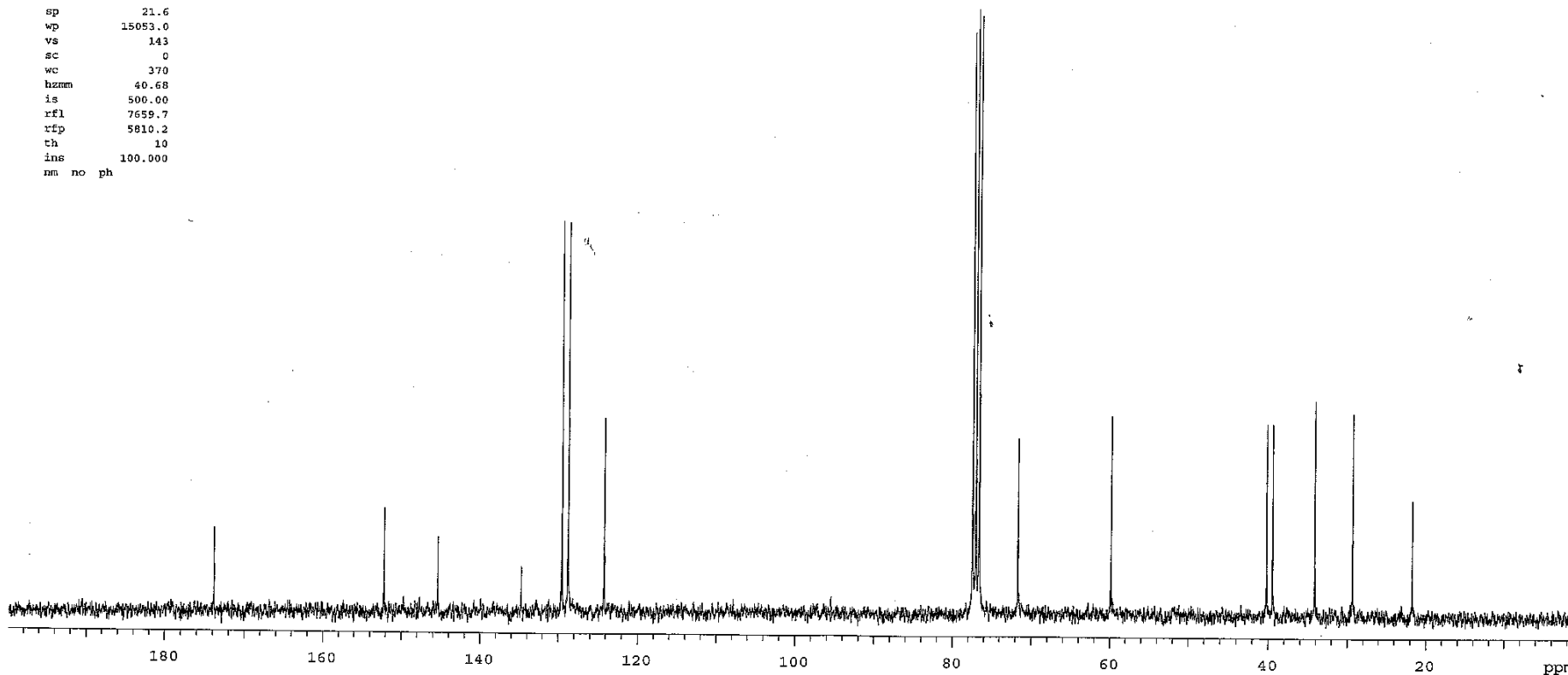
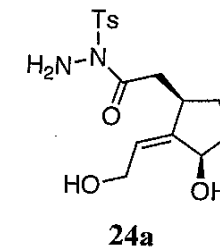
KPa-up-major-C

exp7 std13c

```
SAMPLE          DEC. & VT
date  Nov 11 2008  dfrq      299.973
solvent  CDCl3      dn        H1
file     exp       dpwr      39
ACQUISITION      dof        0
sfrq     75.435    dm         YYY
tn       C13      dmm        W
at       1.815    dmf        11000
np       68106    PROCESSING
sw       18761.7  lb         2.00
fb       10400    wfile
bs       16      proc
lpwr     57      fn         not used
pw       8.7
dl       0      warr
tof      0      wexp      wft
nt       100000  wbs       wft
ct       2208   wat
alock    n
gain     not used

FLAGS
il       n
in       n
dp       y

DISPLAY
sp       21.6
wp       15053.0
vs       143
sc       0
wc       370
hzmm     40.68
ls       500.00
rfl      7659.7
rfp      5810.2
th       10
ins      100.000
nm no ph
```



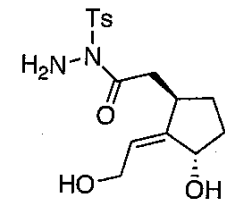
KPa-389-down-H

exp7 std1h

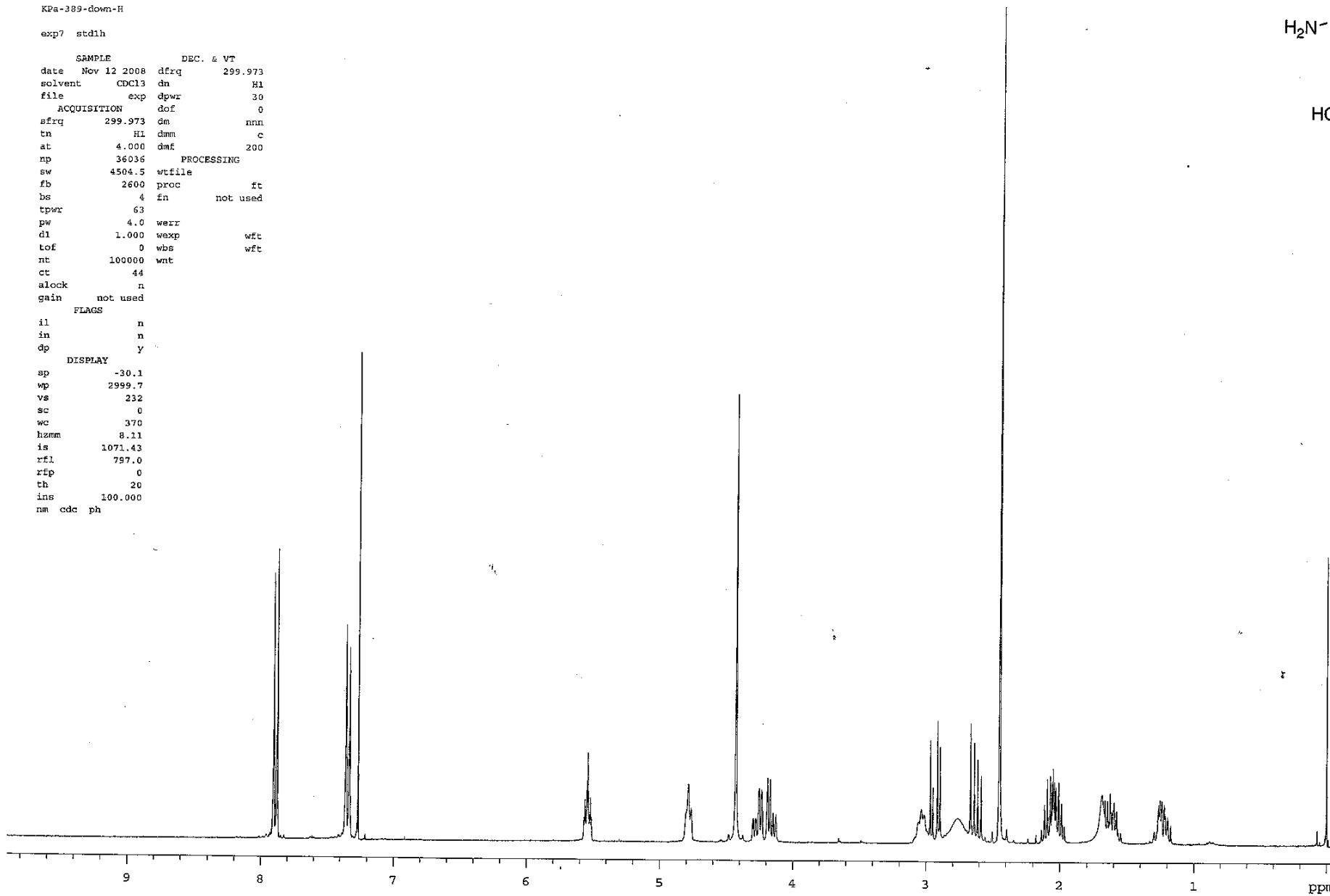
```
SAMPLE          DEC. & VT
date Nov 12 2008 dfrq      299.973
solvent CDCl3  dn         H1
file      exp  dpwr       30
ACQUISITION    dof       0
sfrq      299.973 dm      nnn
tn         H1  dnm       c
at         4.000 dmf      200
np         36036          PROCESSING
sw         4504.5 wfile
fb         2600  proc     ft
bs         4  fn      not used
tpwr       63
pw         4.0  werr
d1         1.000 wexp     wft
tof        0  wbs     wft
nt        100000 wnt
ct         44
alock      n
gain      not used

FLAGS
il         n
in         n
dp         y

DISPLAY
sp         -30.1
wp         2999.7
vs         232
sc         0
wc         370
hnm       8.11
is        1071.43
rfl       797.0
rEp       0
th        20
ins       100.000
nm cdc ph
```



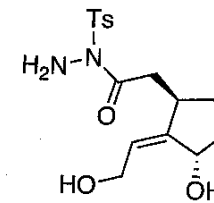
24b



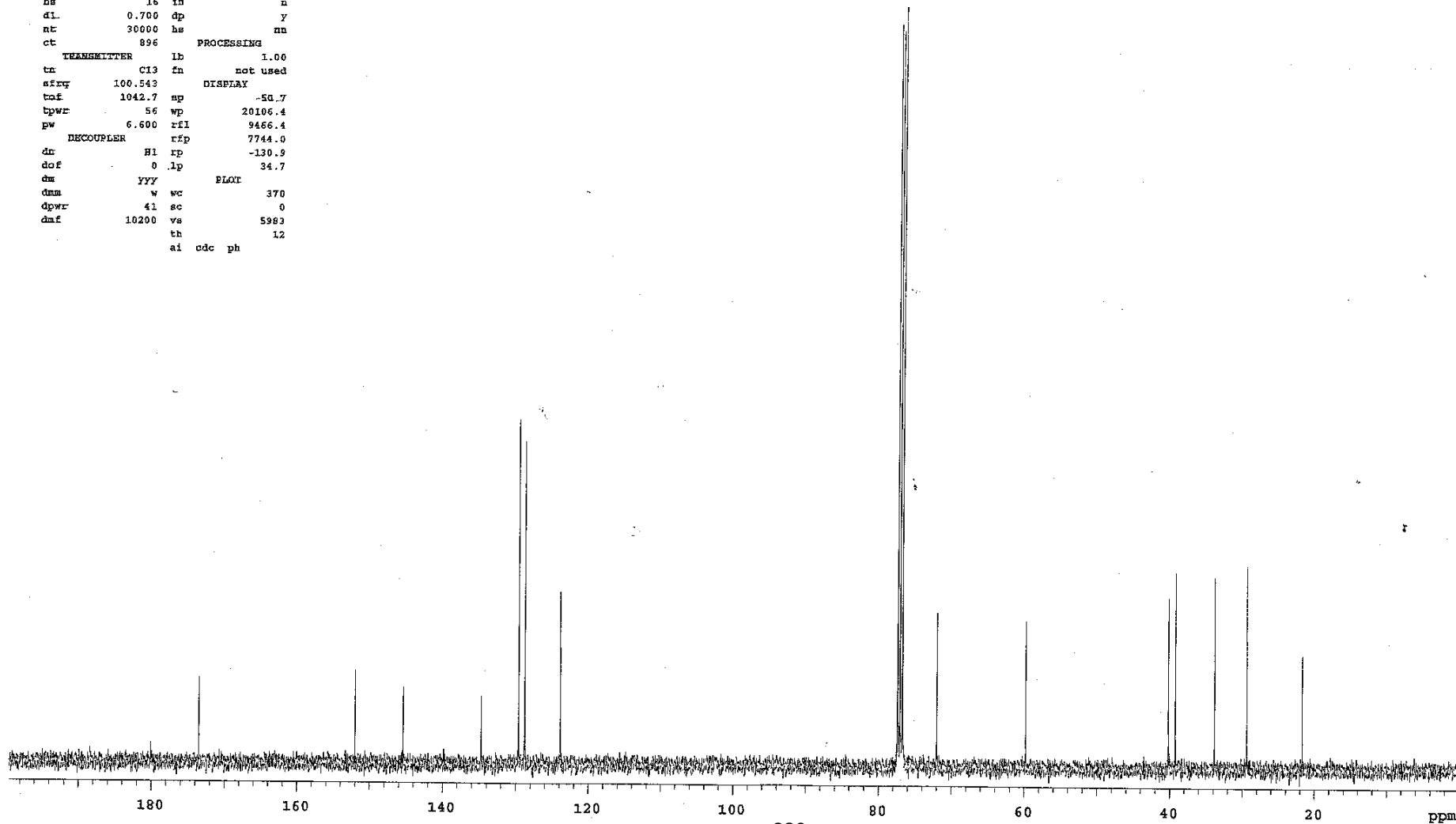
KEA-MS-13C

expl. Carbon

SAMPLE		SPECIALS	
date	Nov 15 2008	temp	not used
solvent	cdc13	gain	not used
file	exp	spin	16
ACQUISITION		ht	0.008
sw	24509.8	pw90	13.200
at	1.300	alfa	10.000
np	63750	PLANS	
fn	17000	ii	rr
hs	16	in	n
dl	0.700	dp	Y
nt	30000	hs	nn
ct	896	PROCESSING	
TRANSMITTER		lb	1.00
tn	CI3	fn	not used
sfrq	100.543	DISPLAY	
taf	1042.7	sp	-50.7
tpwr	56	wp	20106.4
pw	6.600	rfl	9466.4
DECOUPLER		rfp	7744.0
dr	H1	rp	-130.9
dof	0	lp	34.7
dm	YYY	ELOT	
dmm	w	wc	370
dpwr	41	sc	0
dmf	10200	vs	5983
		th	12
		ai	cdc ph



24b

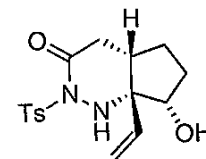


S36

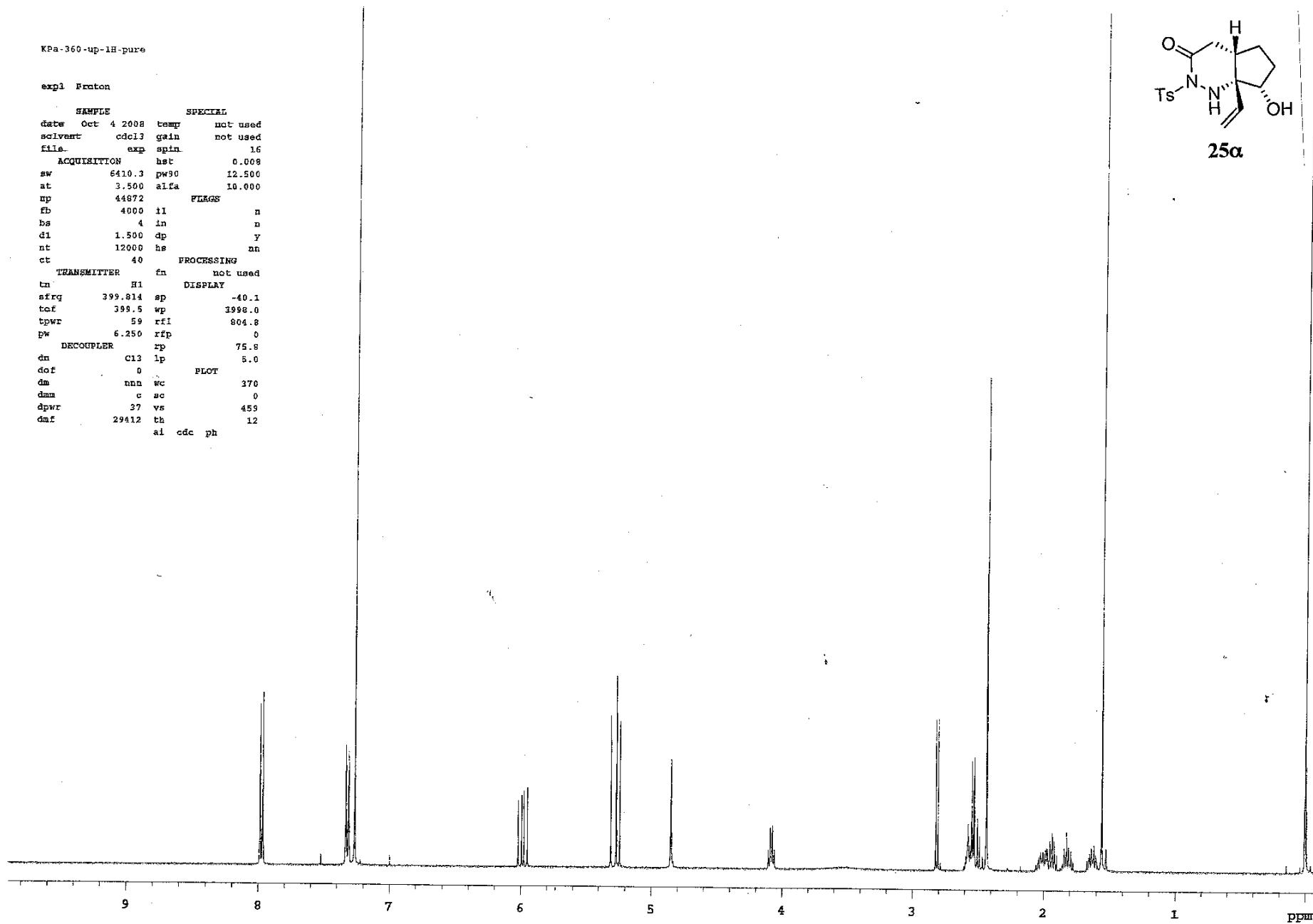
KPa-360-up-1H-pure

exp1 Fracton

SAMPLE		SPECIAL	
date	Oct 4 2008	temp	not used
solvent	cdcl3	gain	not used
file	exp	spin	16
ACQUISITION		FLAGS	
sw	6410.3	pw90	12.500
at	3.500	alfa	10.000
up	44872		
fb	4000	il	n
bs	4	in	n
dl	1.500	dp	y
nt	12000	hs	an
ct	40	PROCESSING	
tn	H1	fn	not used
TRANSMITTER		DISPLAY	
sfreq	399.814	sp	-40.1
toaf	399.5	wp	1998.0
tpwr	59	rfl	804.8
pw	6.250	rfp	0
DECOUPLER		PLOT	
dn	C13	lp	8.0
dof	0		
dm	nnn	wc	170
dmm	c	ac	0
dpwr	37	vs	459
dmf	29412	th	12
	al	cdc	ph



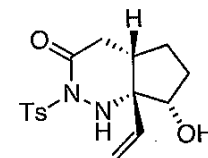
25a



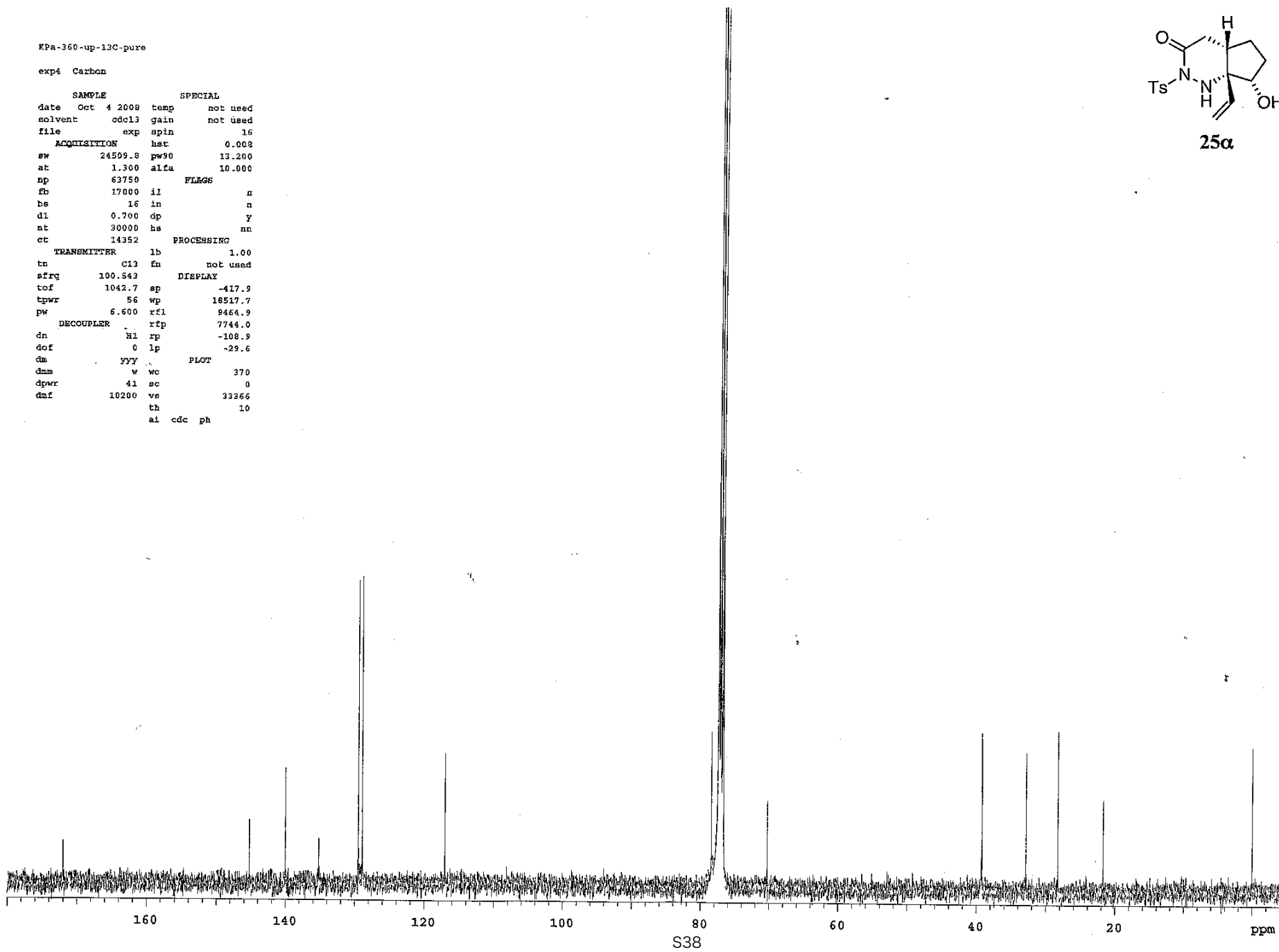
Kpa-360-up-13C-pure

exp4 Carbon

SAMPLE		SPECIAL	
date	Oct 4 2008	temp	not used
solvent	cdcl3	gain	not used
file	exp	spin	16
ACQUISITION		FLAGS	
sw	24509.8	pw90	13.200
at	1.300	alfa	10.000
np	63750		
fb	17000	ij	n
bs	16	in	n
d1	0.700	dp	Y
nt	30000	hs	nn
ct	14352	PROCESSING	
TRANSMITTER		lb	1.00
tn	C13	fn	not used
afreq	100.543	DISPLAY	
tof	1042.7	sp	-417.9
tpwr	56	wp	18517.7
pw	6.600	rfl	9464.9
DECOUPLER		rfd	7744.0
dn	H1	rp	-108.9
dof	0	lp	-29.6
dm	yyy	PLOT	
dmm	w	wc	370
dpwr	41	sc	0
dnr	10200	vs	33366
		th	10
		ai	cdc ph



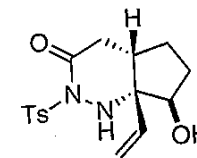
25a



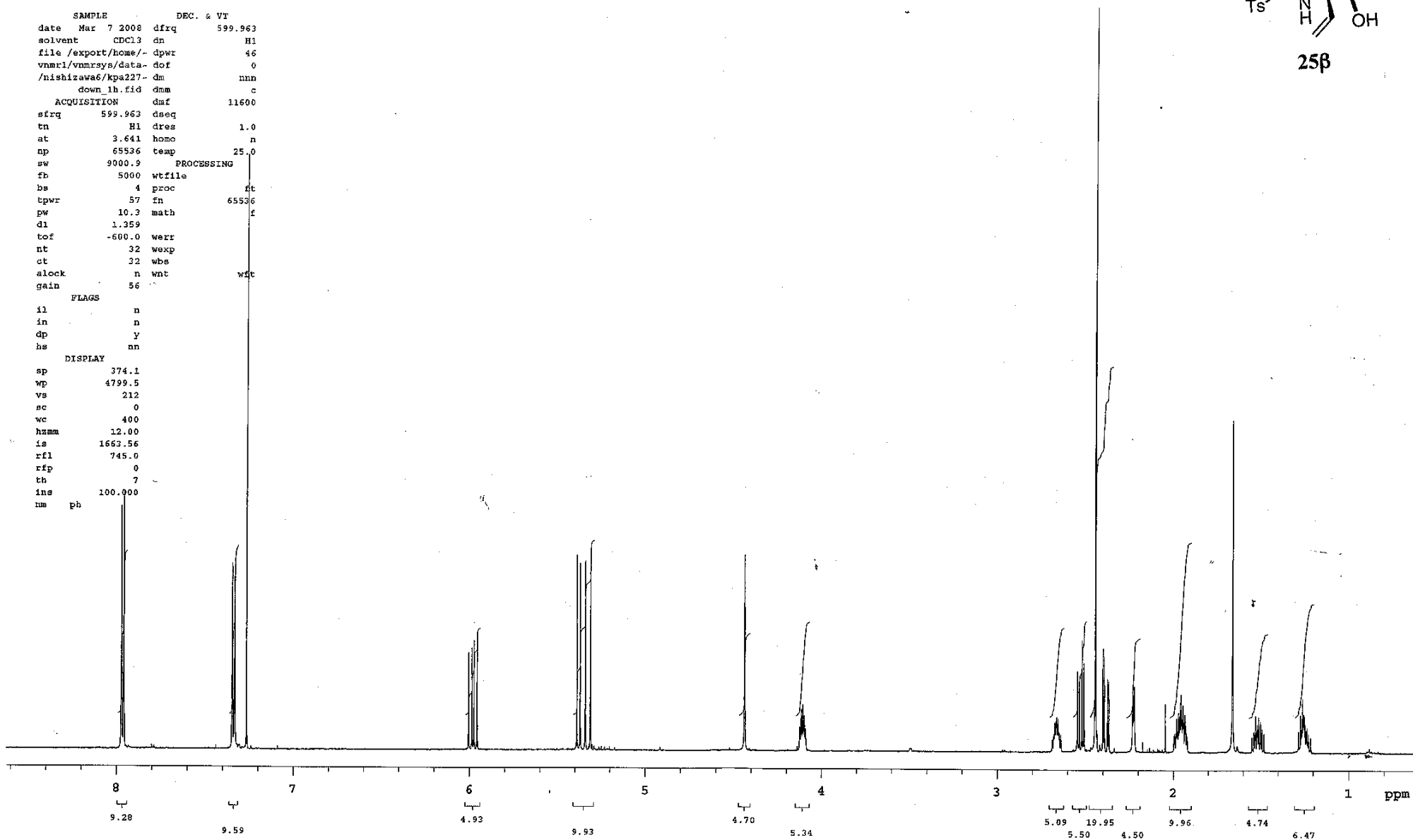
KPa-227-down

expl s2pul

```
SAMPLE          DEC. & VT
date Mar 7 2008  dirq 599.963
solvent CDCl3    dn      H1
file /export/home/- dpwr 46
vnmr1/vnmrsys/data- dof  0
/nishizawa6/kpa227- dm   nnn
down_1h.fid     dnm     c
ACQUISITION     dmf     11600
sfrq 599.963    dseq
tn    H1        drez    1.0
at    3.641     homo    n
np    65536     temp    25.0
sw    9000.9    PROCESsing
fb    5000     wtfile
bs    4        proc
tpwr  57       fn      65536
pw    10.3     math    f
dl    1.359
tof   -600.0   werr
nt    32       wexp
ct    32       wbs
alock n        wnt
gain  56
FLAGS
il    n
in    n
dp    y
hs    nn
DISPLAY
sp    374.1
wp    4799.5
vs    212
sc    0
wc    400
hzmm  12.00
ls    1663.56
rfl   745.0
rfp   0
th    7
ins   100.000
nm    ph
```



25β



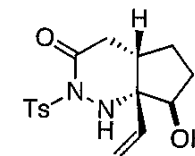
13C OBSERVE

Pulse Sequence: s2pul

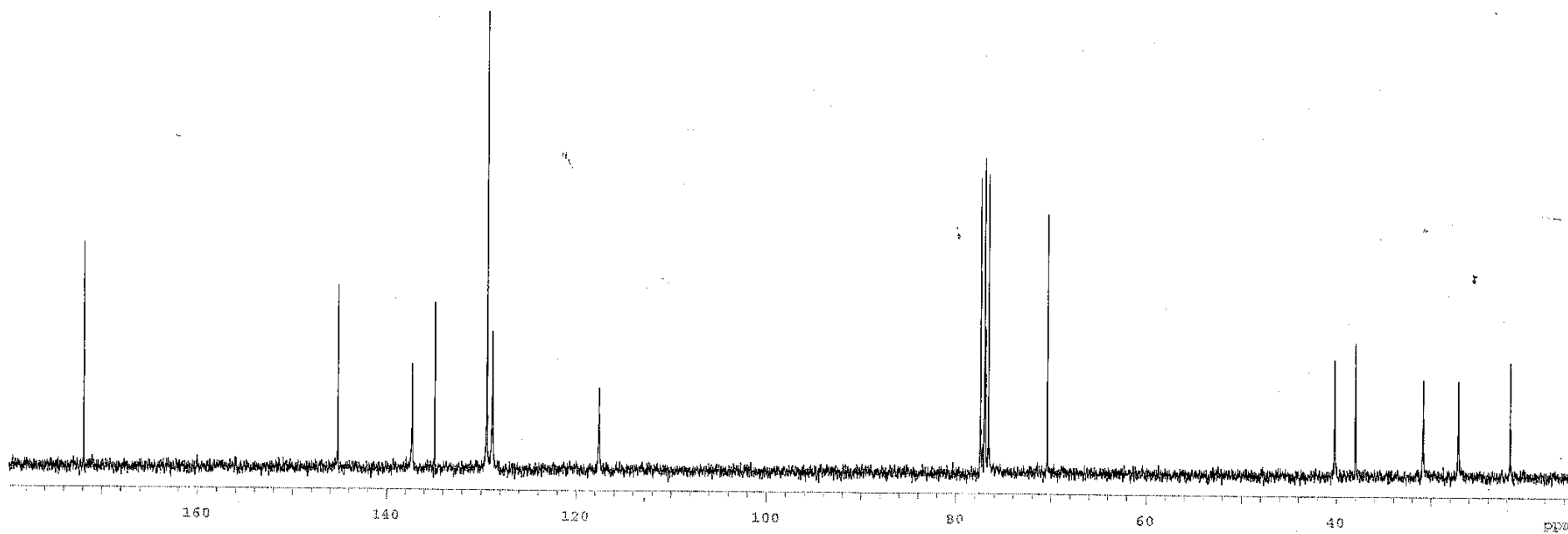
Solvent: CDCl3  
Ambient temperature  
Mercury-300 "m300"

PULSE SEQUENCE

Relax. delay 0.294 sec  
Pulse 40.0 degrees  
Acq. time 1.705 sec  
Width 18863.2 Hz  
608 repetitions  
OBSERVE C13, 75.4839462 MHz  
DECOUPLE H1, 300.1958432 MHz  
Power 38 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
FT size 65536  
Total time 20 minutes



25β

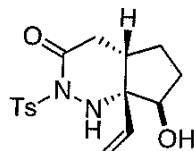




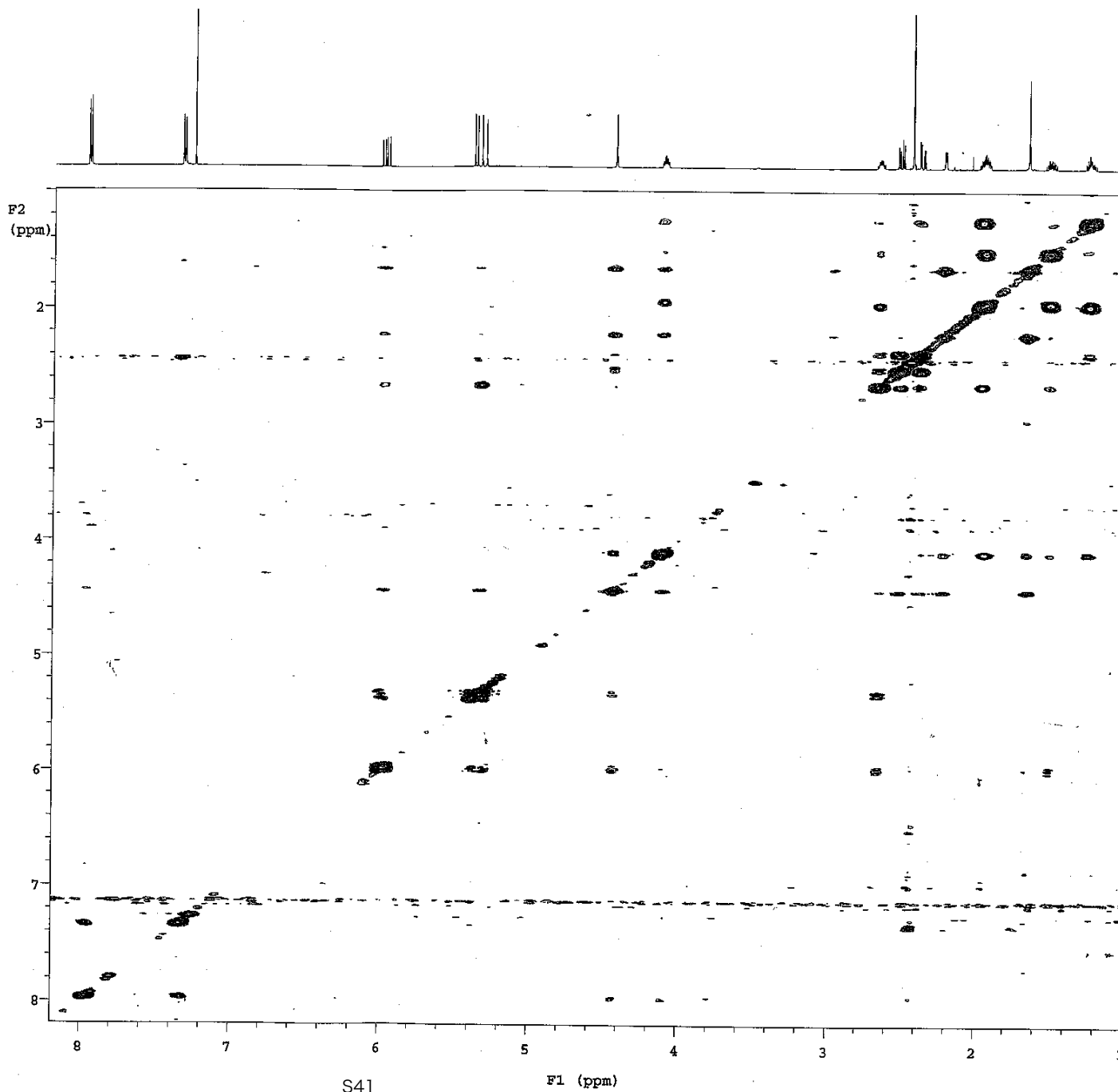
KPa-227-down

exp4 noesy

```
SAMPLE          DEC. & VT      ACQUISITION ARRAYS
date   Mar 7 2008   dfrq      599.963   array      phase
solvent   CDCl3   dn         H1       arraydim    256
file /export/home/- dpwr         45
vnmr1/vnmr3a/data- dof         0
/ntshizsws6/Kpa-227- dn         n         phase
down noesy.fid   dm         c         1
                                2         2
ACQUISITION
sfzq      599.961   cseq      12500
tn         H1   dres      1.0
at         0.170   homo     n
ap         2048   temp      25.0
sw         6024.1   PROCESSING
fb         3400   gf         0.079
bs         4       gfa     not used
ss         8       wtfile
tpwr      57     proc      ft
pw         14.0   fn         2048
dl         1.500   math     f
presat    0
mix       1.200   wvzz
tof       -2077.0 wvxp
nt        16     wbs
ct         8     wnt
alock     n     2D PROCESSING
gain      10    gfl     0.020
          FLAGS   gfl     not used
il        .y    wtfile1
in        n     procl   lp
dp        y     fnl     2048
hs        yn
sspul     y
2D ACQUISITION
sw1       6024.1
n1        128
phase     arrayed
DISPLAY
ap         597.0
wp        4316.4
vs        14500
sc         9
wc         270
hzma     16.00
is        33.57
xf1       733.8
rfp       0
ch        1
ins       100.000
ai        ph
2D DISPLAY
ap1       597.0
wp1       4316.4
sc2       0
wc2       210
xf11     733.8
rfp1     0
```



25β

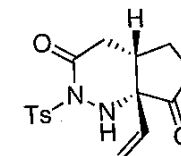


Pulse Sequence: s2pul

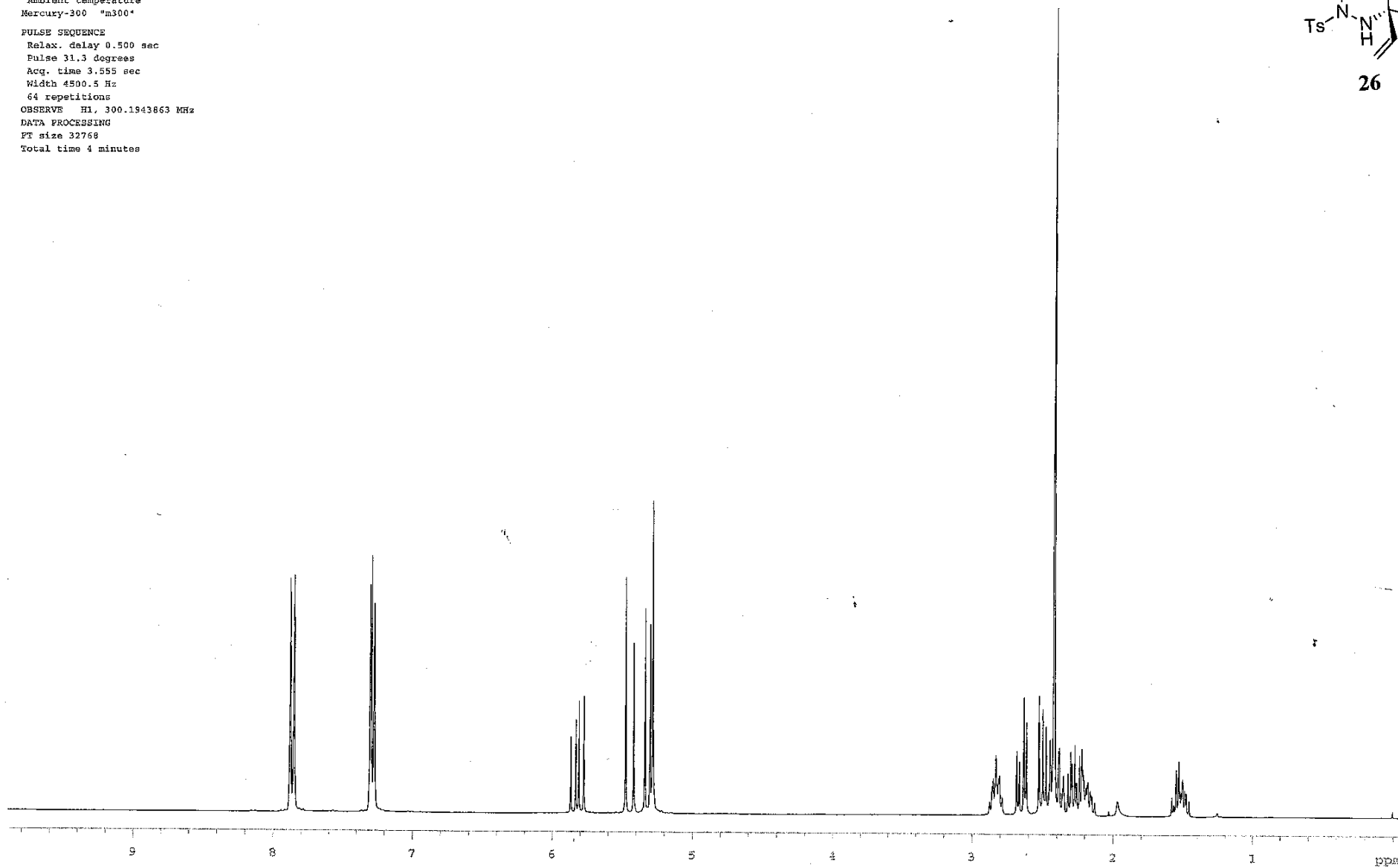
Solvent: CDCl3  
Ambient temperature  
Mercury-300 "m300"

PULSE SEQUENCE

Relax. delay 0.500 sec  
Pulse 31.3 degrees  
Acq. time 3.555 sec  
Width 4500.5 Hz  
64 repetitions  
OBSERVE H1, 300.1943863 MHz  
DATA PROCESSING  
PT size 32768  
Total time 4 minutes



26



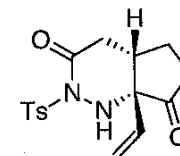
13C OBSERVE

Pulse Sequence: s2pul

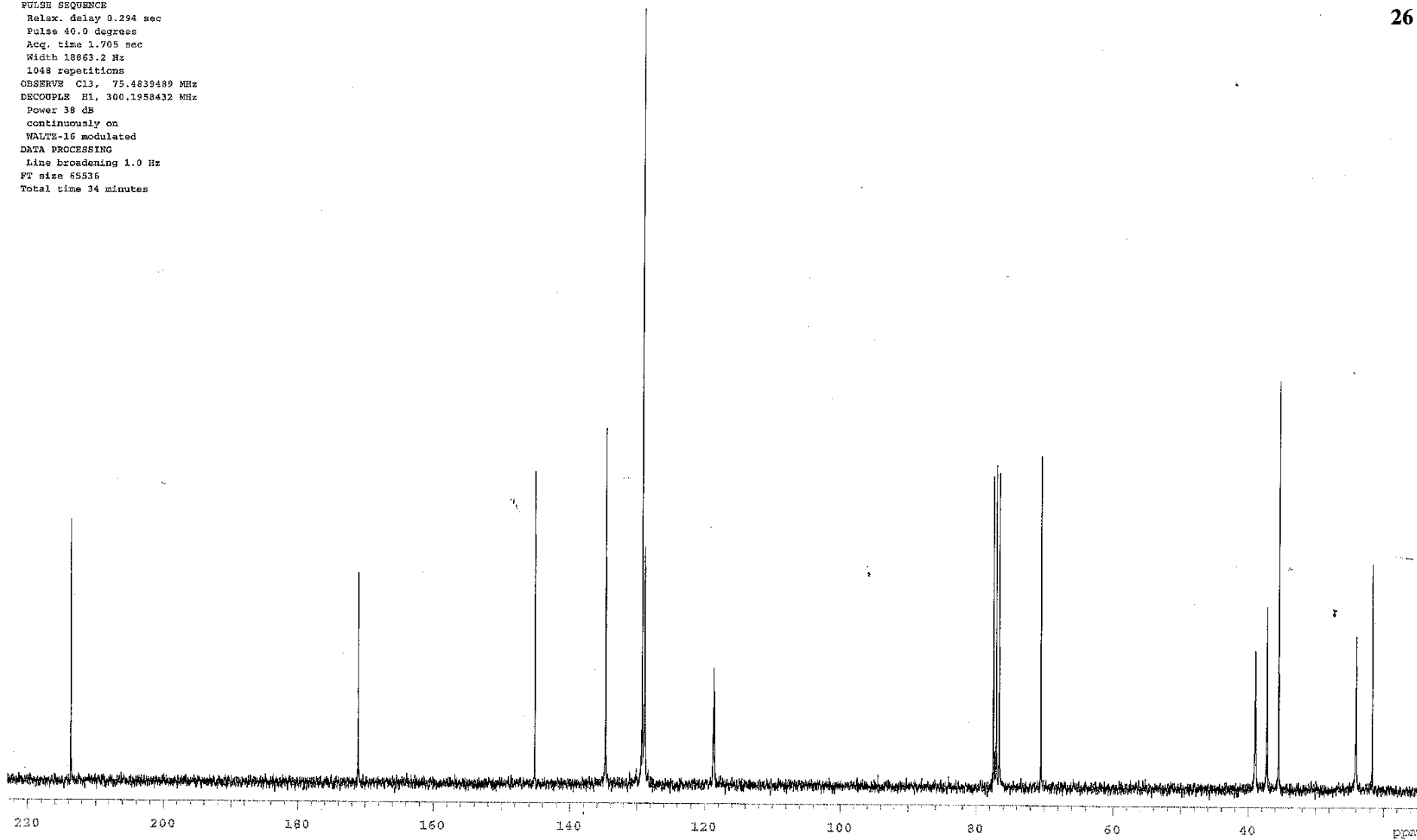
Solvent: CDCl3  
Ambient temperature  
Mercury-300 "m300"

PULSE SEQUENCE

Relax. delay 0.294 sec  
Pulse 40.0 degrees  
Acq. time 1.705 sec  
Width 18663.2 Hz  
1048 repetitions  
OBSERVE C13, 75.4839489 MHz  
DECOUPLE H1, 300.1958432 MHz  
Power 38 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
FT size 65536  
Total time 34 minutes



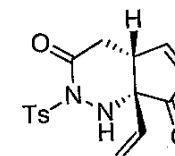
26



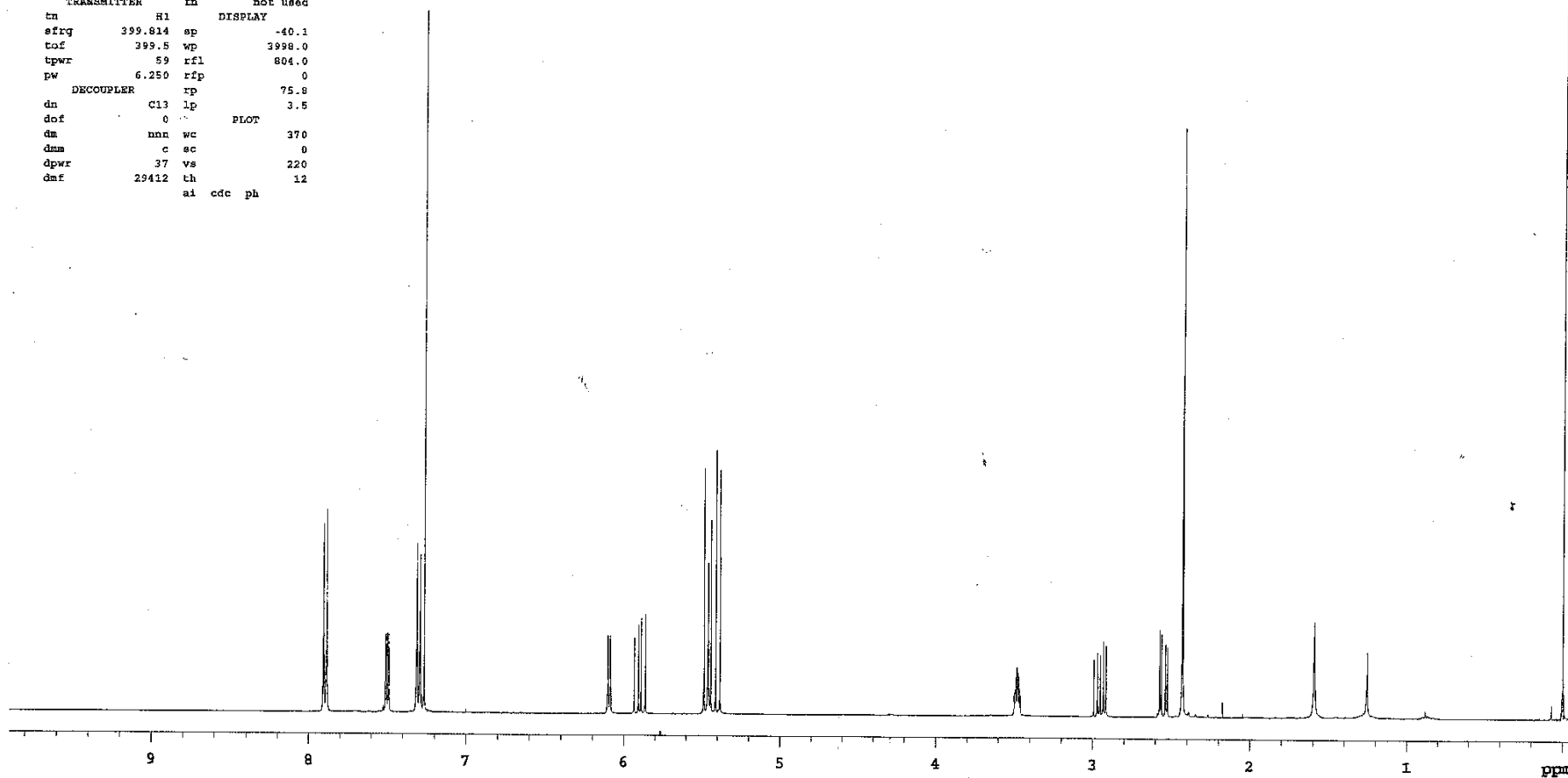
KPa-362-1H-enone

exp2 Proton

SAMPLE		SPECIAL	
date	Oct 3 2008	temp	not used
solvent	cdc13	gain	not used
file	exp	spin	16
ACQUISITION		hst	0.008
sv	6410.3	pw90	12.500
at	3.500	alfa	10.000
np	44972	FLAGS	
fb	4000	il	n
bs	4	in	n
dl	1.500	dp	y
nt	12000	hs	na
ct	28	PROCESSING	
TRANSMITTER		fn	not used
tn	H1	DISPLAY	
sfreq	399.814	sp	-40.1
tof	399.5	wp	3998.0
tpwr	59	rfl	804.0
pw	6.250	rfp	0
DECOUPLER		rp	75.8
dn	C13	lp	3.5
dof	0	PLOT	
dm	mn	wc	370
dmm	c	sc	0
dpwr	37	vs	220
daf	29412	th	12
	ai	cdc	ph



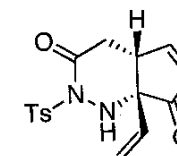
28



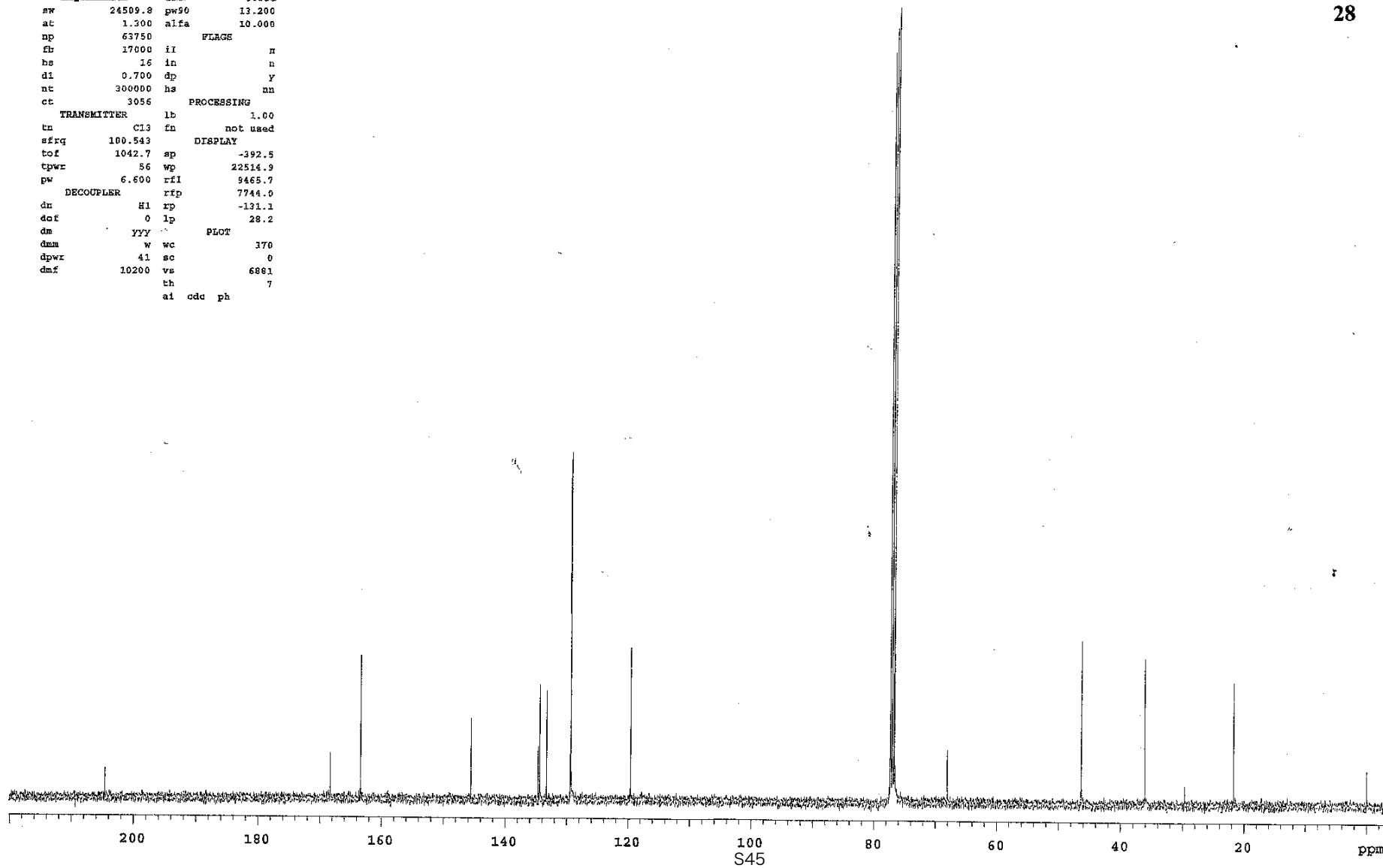
KPa-363-13C-enone

expl Carbon

SAMPLE		SPECIAL	
date	Oct 3 2008	temp	not used
solvent	cdcl3	gain	not used
file	exp	spin	16
ACQUISITION		hst	0.008
sw	24509.8	pw90	13.200
at	1.300	alfa	10.000
np	63750	FLAGE	
fb	17000	ii	n
bs	16	in	n
di	0.700	dp	y
nt	300000	hs	nn
ct	3056	PROCESSING	
TRANSMITTER		lb	1.00
tn	CI3	fn	not used
sfrq	100.543	DISPLAY	
tof	1042.7	sp	-392.5
tpwr	56	wp	22514.9
pw	6.600	rfl	9465.7
DECOUPLER		rtp	7744.0
dn	H1	rp	-131.1
dof	0	lp	28.2
dm	yyy	PLOT	
dmm	w	wc	170
dpwr	41	sc	0
dmf	10200	vs	6881
		th	7
	al	cdc	ph



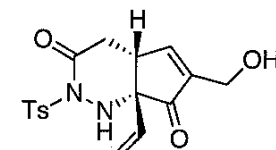
28



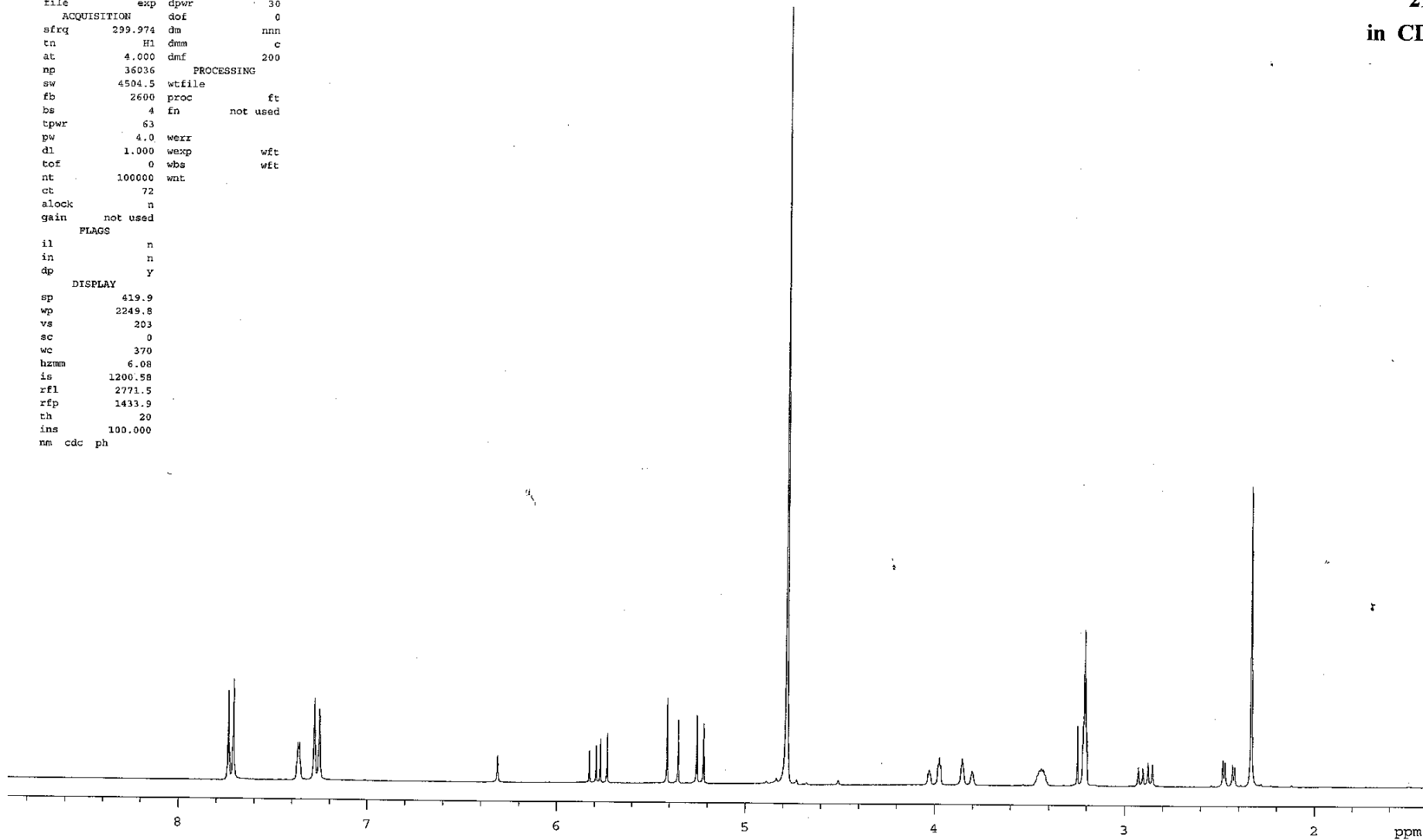
STANDARD 1H OBSERVE

expt stdlh

```
SAMPLE          DEC. & VT
date Sep 21 2008 dfrq 299.974
solvent methanol dn H1
file exp dpwr 30
ACQUISITION dof 0
sfrq 299.974 dm nnn
tn H1 dnm c
at 4.000 dmf 200
np 36036 PROCESSING
sw 4504.5 wtfile
fb 2600 proc ft
bs 4 fn not used
tpwr 63
pw 4.0 wexr
dl 1.000 wexp wft
tof 0 wbs wft
nt 100000 wnt
ct 72
alock n
gain not used
FLAGS
il n
in n
dp y
DISPLAY
sp 419.9
wp 2249.8
vs 203
sc 0
wc 370
hzmm 6.08
ls 1200.58
rfl 2771.5
rfp 1433.9
th 20
ins 100.000
nm cdc ph
```



29  
in CD<sub>3</sub>OD



13C OBSERVE

Pulse Sequence: s2pul

Solvent: methanol

Ambient temperature

File: KPa-354-13C-092208\_02\_43

INOVA-600 "NMR"

Pulse 54.2 degrees

Acq. time 1.815 sec

Width 18761.7 Hz

11237 repetitions

OBSERVE C13, 75.4280880 MHz

DECOUPLE H1, 299.9737314 MHz

Power 39 dB

continuously on

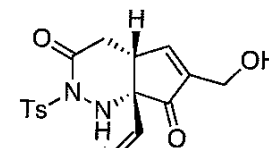
WALTZ-16 modulated

DATA PROCESSING

Line broadening 2.0 Hz

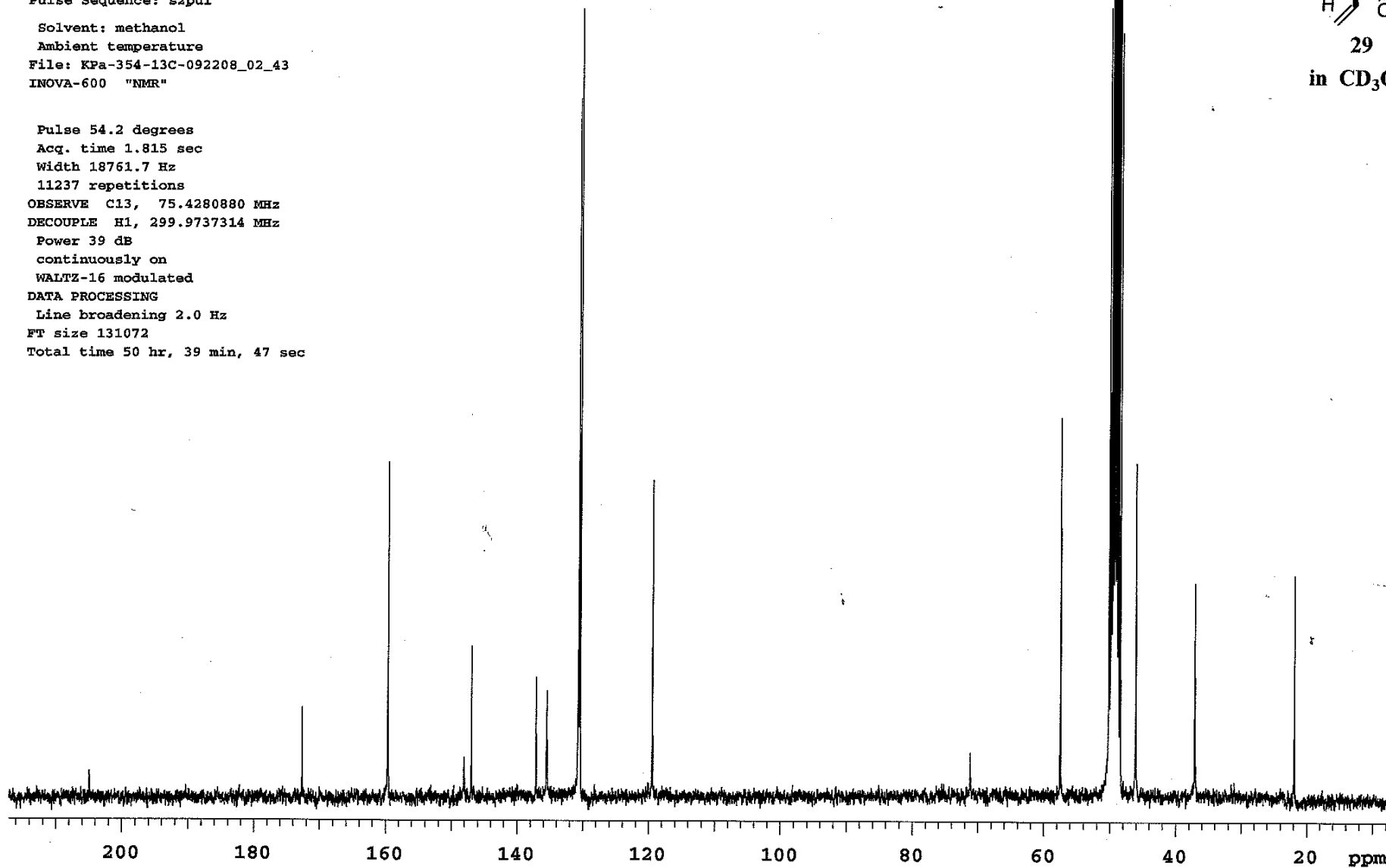
FT size 131072

Total time 50 hr, 39 min, 47 sec



29

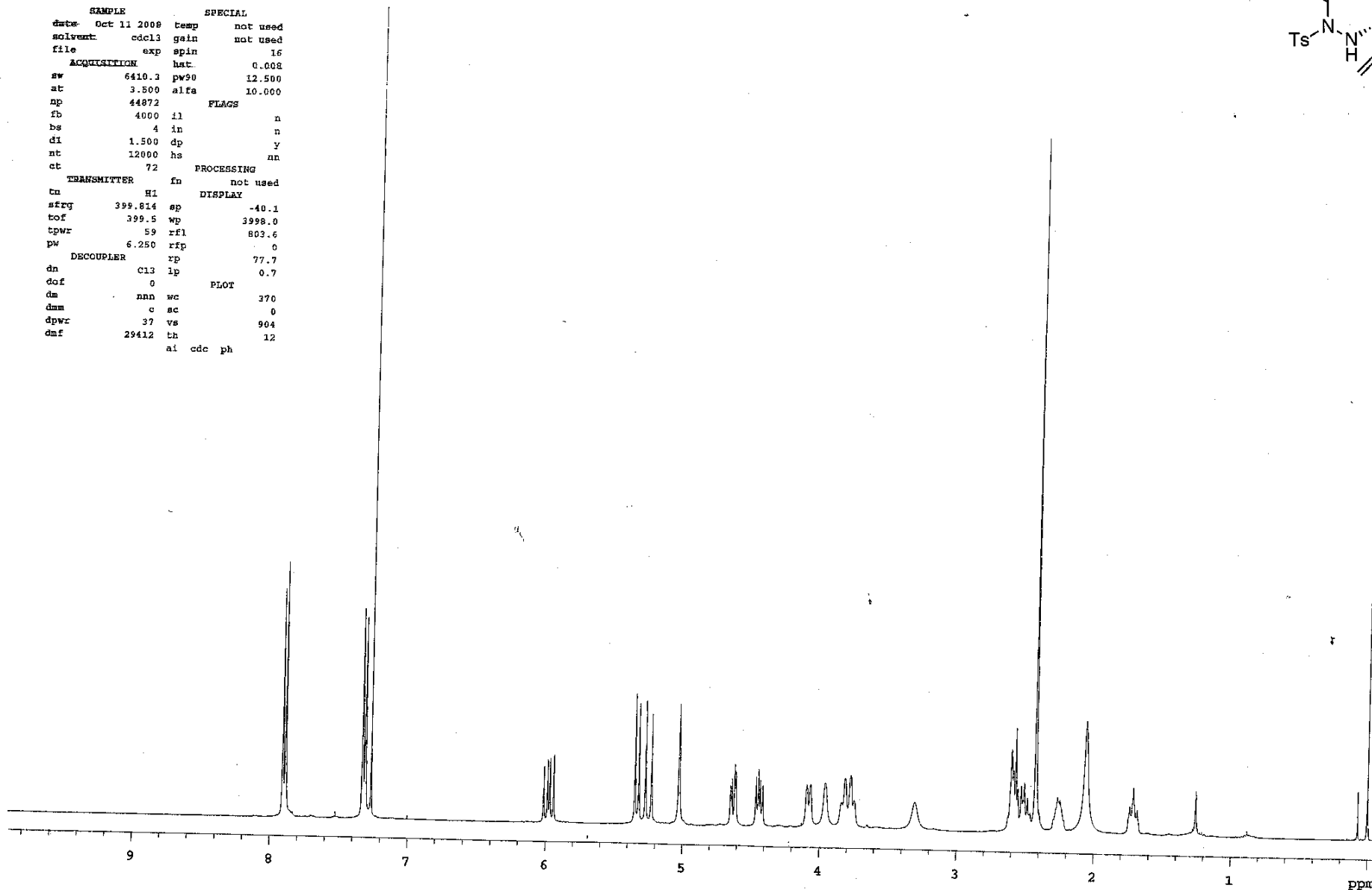
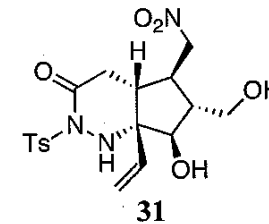
in CD<sub>3</sub>OD



KPa-366-1H-pure

expt Proton

SAMPLE		SPECIAL	
date	Oct 11 2009	temp	not used
solvent	cdcl3	gain	not used
file	exp	spin	16
ACQUISITION		FLAGS	
sw	6410.1	pw90	12.500
at	3.500	alfa	10.000
np	44872		
fb	4000	il	n
bs	4	in	n
dl	1.500	dp	y
nt	12000	hs	an
ct	72		
TRANSMITTER		PROCESSING	
tn	H1	fn	not used
sfry	399.814	sp	-40.1
tof	399.5	wp	3998.0
tpwr	59	rfl	803.6
pw	6.250	rfp	0
DECOUPLER		PLOT	
dn	C13	lp	0.7
dof	0		
dm	nnn	wc	270
dmm	c	ac	0
dpwr	37	vs	904
dmf	29412	tn	12
	ai	cdc	ph

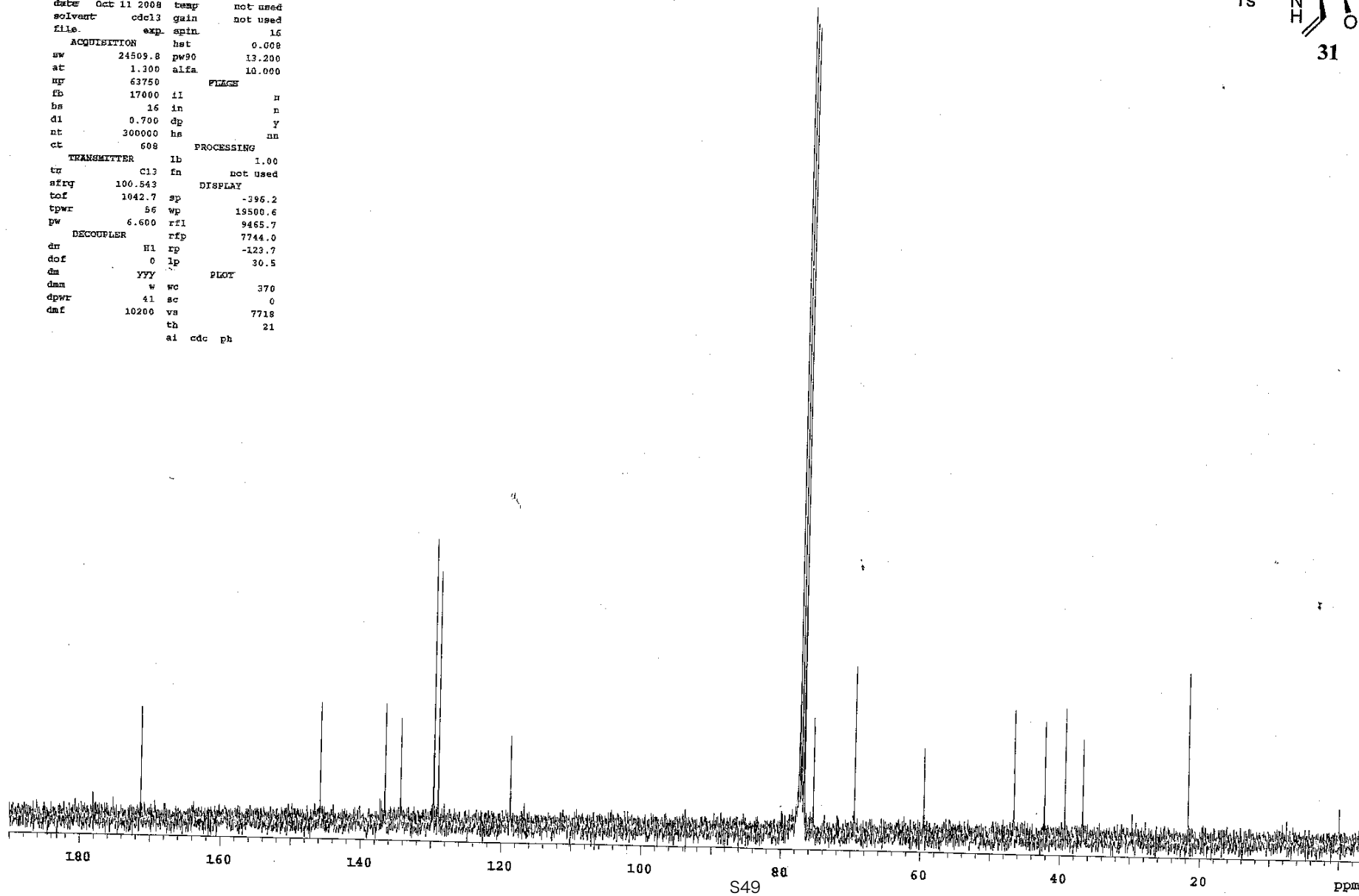
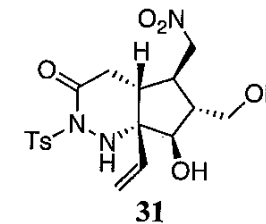


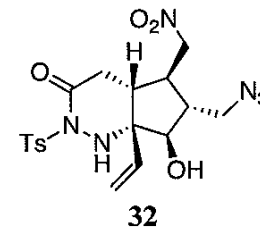


KEa-166-11C-pure

exp4 Carbon

SAMPLE		SPECIAL	
date	Oct 11 2008	temp	not used
solvent	cdcl3	gain	not used
file	exp	spin	16
ACQUISITION		bst	0.008
sw	24509.8	pw90	13.200
at	1.300	alfa	10.000
np	63750	PLACS	
fb	17000	ii	n
bs	16	in	n
d1	0.700	dp	Y
nt	300000	hs	un
ct	608	PROCESSING	
TRANSMITTER		lb	1.00
tu	C13	fn	not used
sfrq	100.643	DISPLAY	
tof	1042.7	sp	-396.2
tpwr	56	wp	19500.6
pw	6.600	rfl	9465.7
DECOUPLER		r1p	7744.0
dn	H1	rp	-123.7
dof	0	lp	30.5
dm	YYY	PLOT	
dmm	w	wc	370
dpwr	41	sc	0
dmf	10200	vs	7718
		th	21
		al	cdc ph





Pulse Sequence: s2pul

Solvent: CDCl3

Ambient temperature

File: KPa-403-pure-1H-112608\_23\_51

INOVA-600 "NMR"

Relax. delay 0.500 sec

Pulse 31.3 degrees

Acq. time 3.555 sec

Width 4500.5 Hz

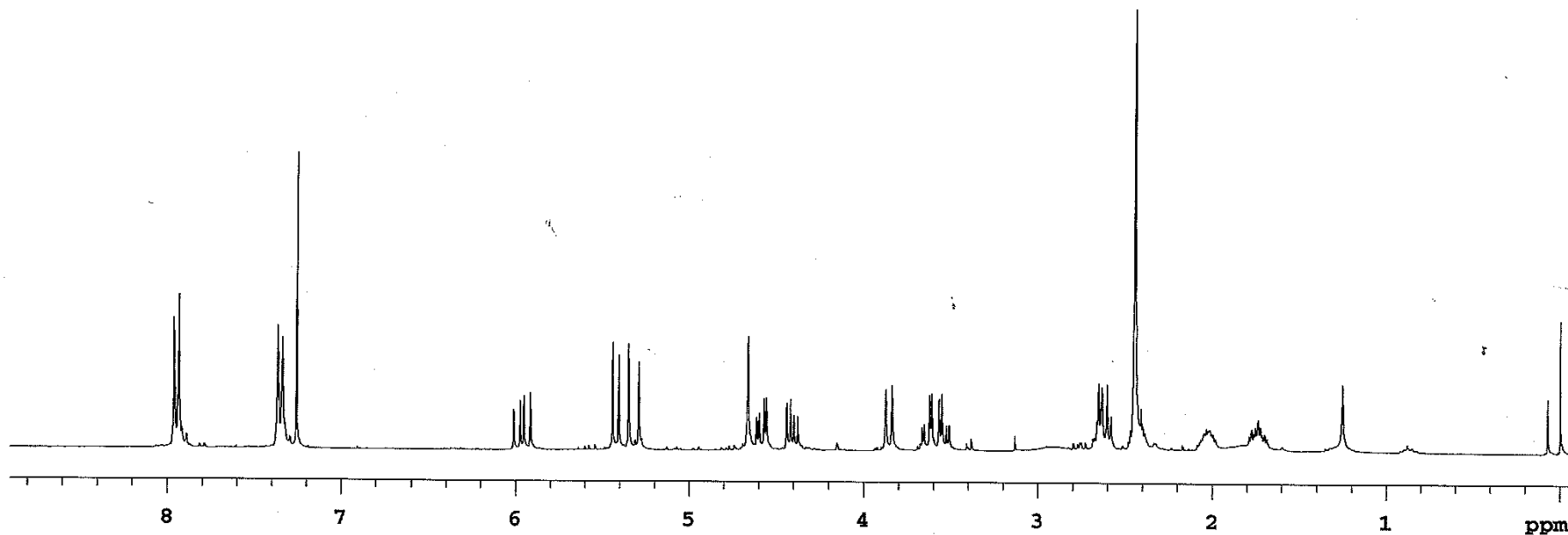
268 repetitions

OBSERVE H1, 300.1943971 MHz

DATA PROCESSING

FT size 32768

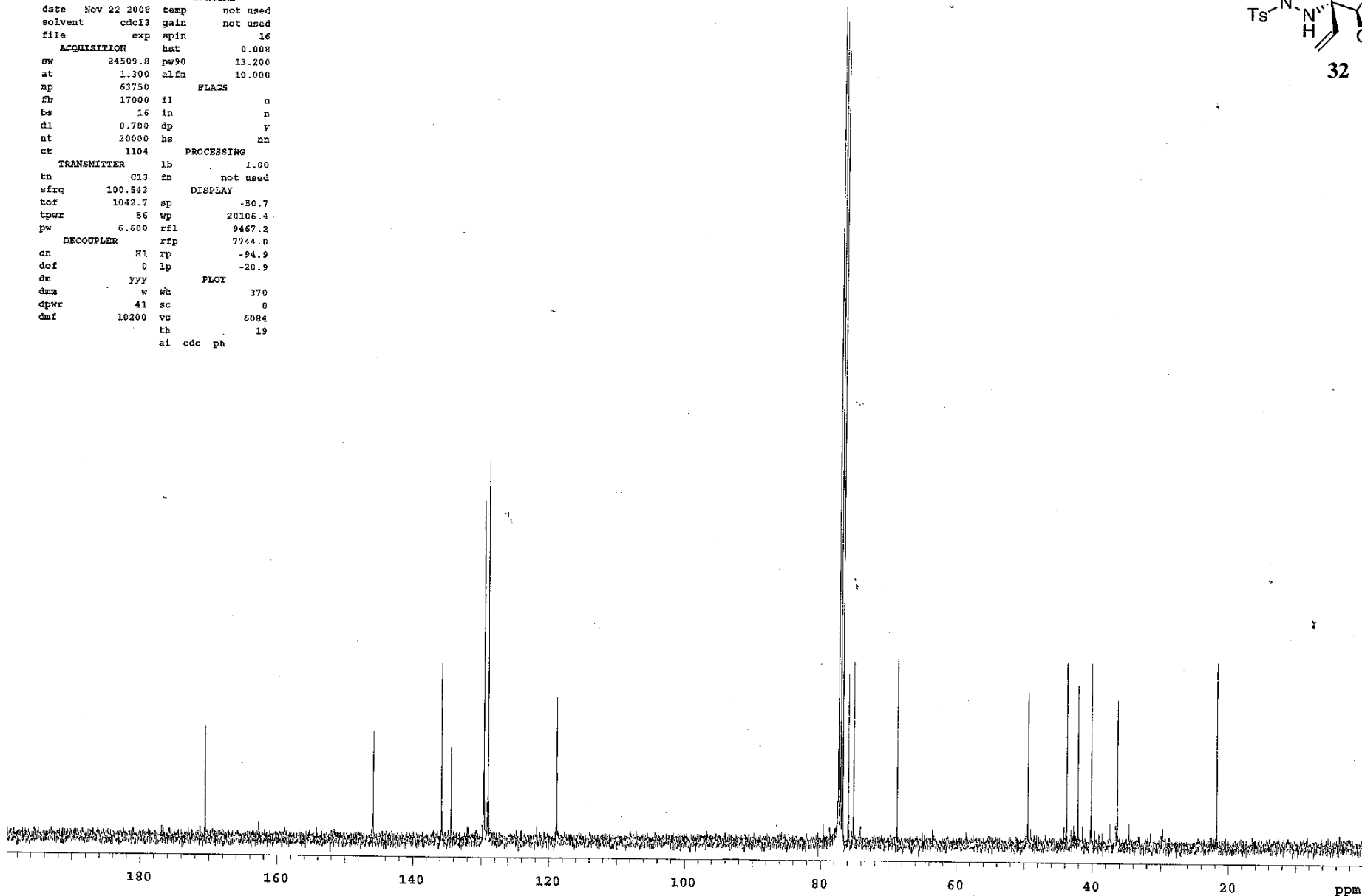
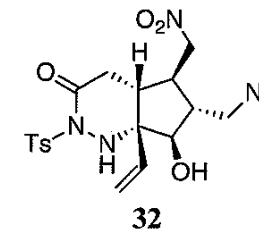
Total time 11 hr, 17 min, 20 sec



KPa-403-L3C

exp4 Carbon

SAMPLE		SPECIAL	
date	Nov 22 2008	temp	not used
solvent	cdc13	gain	not used
file	exp	spin	16
ACQUISITION		hat	0.008
sw	24509.8	pw90	13.200
at	1.300	alfa	10.000
ap	63750	FLAGS	
fb	17000	ii	n
bs	16	in	n
dl	0.700	dp	Y
nt	30000	hs	nn
ct	1104	PROCESSING	
TRANSMITTER		lb	1.00
tn	C13	fb	not used
sfrq	100.543	DISPLAY	
tof	1042.7	sp	-50.7
tpwr	56	wp	20106.4
pw	6.600	rfl	9467.2
DECOUPLER		rfp	7744.0
dn	H1	rp	-94.9
dof	0	lp	-20.9
dm	YYY	PLOT	
dmm	w	wc	370
dpwr	41	sc	0
dmf	10200	vs	6084
		th	19
		al	cdc ph

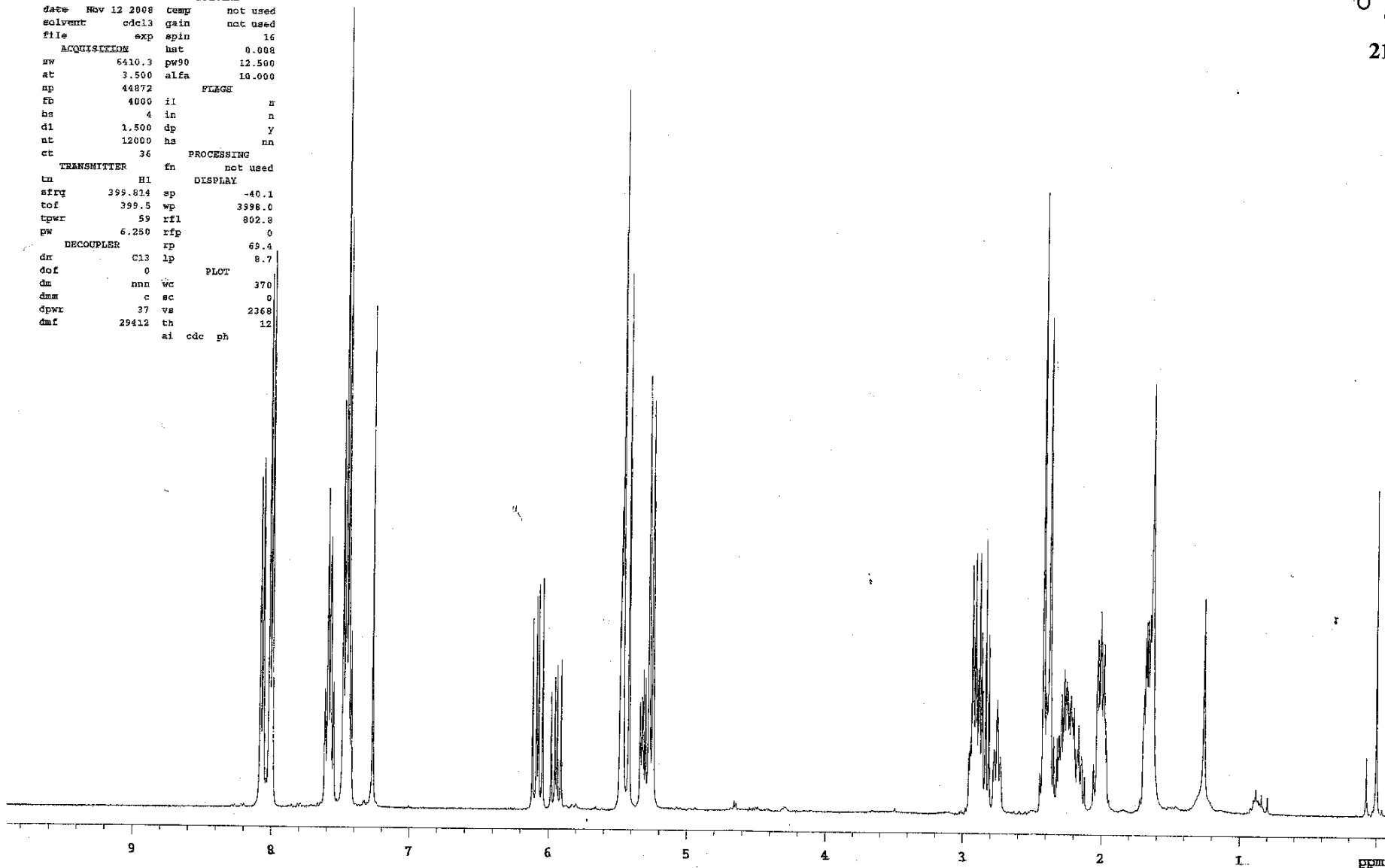
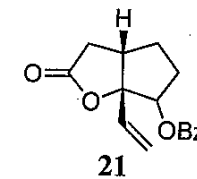


S51

KPa-390(up-middle)-mix-1H

expl. Proton

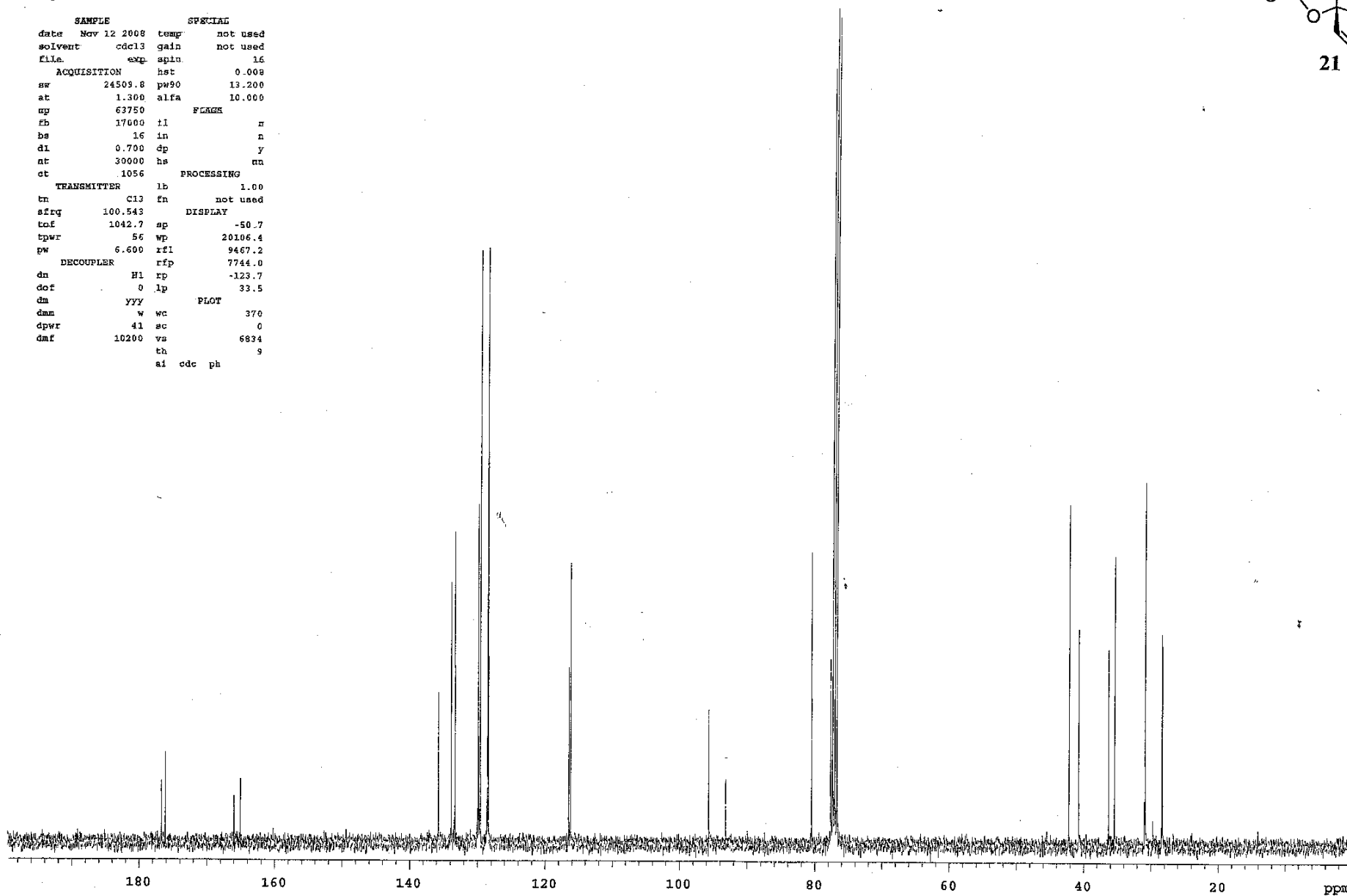
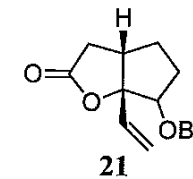
SAMPLE		SPECIAL	
date	Nov 12 2008	temp	not used
solvent	cdcl3	gain	not used
file		exp	spin 16
ACQUISITION		nat	0.008
sw	6410.3	pw90	12.500
at	3.500	alfa	10.000
np	44872	FLAGS	
fb	4000	il	r
hs	4	in	n
dl	1.500	dp	y
nt	12000	hs	nn
ct	36	PROCESSING	
TRANSMITTER		fn	not used
tu	H1	DISPLAY	
sfreq	399.614	sp	-40.1
tof	399.5	wp	3998.0
tpwr	59	rfl	802.8
pw	6.250	rfp	0
DECOUPLER		rp	69.4
dr	Cl3	lp	8.7
dof	0	PLOT	
dm	nnn	wc	370
dmm	c	sc	0
dpwr	37	vs	2368
dmi	29412	th	12
	al	cdc	ph



KEA-190(up-middle)-mix-13C

exp3 Carbon

SAMPLE		SPECIAL	
date	Nov 12 2008	temp	not used
solvent	cdc13	gain	not used
file	exp	spin	16
ACQUISITION		PARAMETERS	
sw	24509.8	ps90	13.200
at	1.300	alfa	10.000
sp	63750	FLAGS	
fb	17000	tl	n
bs	16	in	n
dl	0.700	dp	y
nt	30000	hs	un
ct	1056	PROCESSING	
TRANSMITTER		lb	1.00
tr	C13	fn	not used
sfrq	100.543	DISPLAY	
tof	1042.7	sp	-50.7
tpwr	56	wp	20106.4
pw	6.600	rfl	9467.2
DECOUPLER		rfp	7744.0
dn	H1	rp	-123.7
dof	0	lp	33.5
dm	yyy	PLOT	
dms	w	wc	370
dpwr	41	sc	0
dms	10200	vs	6834
		th	9
		ai	cdc ph



S53

Table 1. Crystal data and structure refinement for p2.

Identification code	p2	
Empirical formula	C <sub>16</sub> H <sub>20</sub> N <sub>2</sub> O <sub>4</sub> S	
Formula weight	336.40	
Temperature	296 K	
Wavelength	0.71073 Å	
Crystal system	Monoclinic	
Space group	P2(1)/c	
Unit cell dimensions	a = 15.677(5) Å	$\alpha = 90^\circ$ .
	b = 8.062(3) Å	$\beta = 99.968(4)^\circ$ .
	c = 12.985(4) Å	$\gamma = 90^\circ$ .
Volume	1616.3(9) Å <sup>3</sup>	
Z	4	
Density (calculated)	1.382 Mg/m <sup>3</sup>	
Absorption coefficient	0.222 mm <sup>-1</sup>	
F(000)	712	
Crystal size	0.43 x 0.25 x 0.23 mm <sup>3</sup>	
Theta range for data collection	1.32 to 27.55°.	
Index ranges	-20 ≤ h ≤ 9, -10 ≤ k ≤ 10, -16 ≤ l ≤ 16	
Reflections collected	7922	
Independent reflections	3654 [R(int) = 0.0632]	
Completeness to theta = 27.55°	97.6 %	
Absorption correction	None	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	3654 / 0 / 211	
Goodness-of-fit on F <sup>2</sup>	1.053	
Final R indices [I > 2σ(I)]	R1 = 0.0875, wR2 = 0.2392	
R indices (all data)	R1 = 0.0897, wR2 = 0.2436	
Extinction coefficient	0.002(4)	
Largest diff. peak and hole	0.932 and -0.872 e.Å <sup>-3</sup>	

Table 2. Atomic coordinates (x 10<sup>4</sup>) and equivalent isotropic displacement parameters (Å<sup>2</sup> x 10<sup>3</sup>) for p2. U(eq) is defined as one third of the trace of the orthogonalized U<sup>ij</sup> tensor.

	x	y	z	U(eq)
S(1)	2442(1)	10919(1)	3056(1)	17(1)
C(2)	3985(2)	5770(3)	789(2)	24(1)
C(3)	3350(1)	6960(3)	1215(2)	18(1)
C(4)	3041(2)	6321(3)	2199(2)	23(1)
C(5)	2502(2)	7638(3)	2602(2)	20(1)
C(6)	3849(1)	8638(2)	1451(2)	15(1)
C(7)	4808(1)	8168(3)	1459(2)	20(1)
C(8)	4734(2)	6876(3)	587(2)	25(1)
C(9)	3583(2)	9993(3)	656(2)	23(1)
C(10)	1512(1)	11258(3)	2108(2)	17(1)
C(11)	709(2)	10696(3)	2294(2)	21(1)
C(12)	-26(2)	11071(3)	1565(2)	24(1)
C(13)	34(2)	11997(3)	669(2)	23(1)
C(14)	846(2)	12517(3)	498(2)	24(1)
C(15)	1593(2)	12165(3)	1219(2)	21(1)
C(16)	-776(2)	12413(3)	-102(2)	31(1)
O(1)	5128(1)	7430(2)	2451(1)	23(1)
O(2)	1776(1)	7476(2)	2795(2)	27(1)
N(1)	2927(1)	9172(2)	2705(2)	16(1)
N(2)	3781(1)	9313(2)	2495(1)	14(1)
O(3)	2184(1)	10601(3)	4043(1)	27(1)

O(4)	3061(1)	12211(2)	2972(1)	24(1)
C(17)	2982(2)	9926(4)	-184(2)	37(1)

Table 3. Bond lengths [Å] and angles [°] for p2.

---

S(1)-O(3)	1.4316(18)
S(1)-O(4)	1.4403(17)
S(1)-N(1)	1.6995(19)
S(1)-C(10)	1.759(2)
C(2)-C(8)	1.532(4)
C(2)-C(3)	1.552(3)
C(3)-C(4)	1.532(3)
C(3)-C(6)	1.567(3)
C(4)-C(5)	1.505(3)
C(5)-O(2)	1.215(3)
C(5)-N(1)	1.400(3)
C(6)-N(2)	1.481(3)
C(6)-C(9)	1.511(3)
C(6)-C(7)	1.549(3)
C(7)-O(1)	1.429(3)
C(7)-C(8)	1.528(3)
C(9)-C(17)	1.314(4)
C(10)-C(15)	1.391(3)
C(10)-C(11)	1.397(3)
C(11)-C(12)	1.391(3)
C(12)-C(13)	1.400(3)
C(13)-C(14)	1.393(4)
C(13)-C(16)	1.513(3)
C(14)-C(15)	1.397(3)
N(1)-N(2)	1.417(3)
O(3)-S(1)-O(4)	119.72(11)
O(3)-S(1)-N(1)	107.47(10)
O(4)-S(1)-N(1)	103.91(10)
O(3)-S(1)-C(10)	109.03(11)
O(4)-S(1)-C(10)	108.79(11)
N(1)-S(1)-C(10)	107.18(10)
C(8)-C(2)-C(3)	105.02(18)
C(4)-C(3)-C(2)	113.85(18)
C(4)-C(3)-C(6)	110.27(17)
C(2)-C(3)-C(6)	105.90(18)
C(5)-C(4)-C(3)	109.28(17)
O(2)-C(5)-N(1)	121.5(2)
O(2)-C(5)-C(4)	126.7(2)
N(1)-C(5)-C(4)	111.76(19)
N(2)-C(6)-C(9)	107.74(17)
N(2)-C(6)-C(7)	107.93(16)
C(9)-C(6)-C(7)	109.50(17)
N(2)-C(6)-C(3)	112.57(16)
C(9)-C(6)-C(3)	114.87(18)
C(7)-C(6)-C(3)	103.98(16)
O(1)-C(7)-C(8)	110.75(19)
O(1)-C(7)-C(6)	107.22(17)
C(8)-C(7)-C(6)	102.25(17)
C(7)-C(8)-C(2)	103.45(18)
C(17)-C(9)-C(6)	127.7(2)
C(15)-C(10)-C(11)	121.9(2)

C(15)-C(10)-S(1)	118.63(17)
C(11)-C(10)-S(1)	119.41(17)
C(12)-C(11)-C(10)	118.5(2)
C(11)-C(12)-C(13)	121.0(2)
C(14)-C(13)-C(12)	119.2(2)
C(14)-C(13)-C(16)	120.9(2)
C(12)-C(13)-C(16)	119.9(2)
C(13)-C(14)-C(15)	121.0(2)
C(10)-C(15)-C(14)	118.5(2)
C(5)-N(1)-N(2)	120.11(17)
C(5)-N(1)-S(1)	122.06(16)
N(2)-N(1)-S(1)	117.82(14)
N(1)-N(2)-C(6)	111.50(16)

Symmetry transformations used to generate equivalent atoms:

Table 4. Anisotropic displacement parameters ( $\text{\AA}^2 \times 10^3$ ) for p2. The anisotropic displacement factor exponent takes the form:  $-2\pi^2 [ h^2 a^2 U^{11} + \dots + 2 h k a^* b^* U^{12} ]$

	$U^{11}$	$U^{22}$	$U^{33}$	$U^{23}$	$U^{13}$	$U^{12}$
S(1)	16(1)	16(1)	20(1)	-5(1)	3(1)	2(1)
C(2)	22(1)	18(1)	30(1)	-11(1)	1(1)	3(1)
C(3)	19(1)	12(1)	21(1)	-3(1)	-1(1)	0(1)
C(4)	24(1)	9(1)	36(1)	2(1)	7(1)	-1(1)
C(5)	20(1)	13(1)	26(1)	5(1)	5(1)	1(1)
C(6)	17(1)	12(1)	16(1)	-1(1)	4(1)	0(1)
C(7)	17(1)	19(1)	23(1)	-5(1)	5(1)	0(1)
C(8)	24(1)	26(1)	26(1)	-9(1)	6(1)	5(1)
C(9)	30(1)	16(1)	26(1)	6(1)	10(1)	4(1)
C(10)	15(1)	14(1)	20(1)	-3(1)	1(1)	3(1)
C(11)	19(1)	21(1)	22(1)	-1(1)	6(1)	2(1)
C(12)	17(1)	27(1)	27(1)	-2(1)	4(1)	4(1)
C(13)	23(1)	17(1)	27(1)	-4(1)	1(1)	7(1)
C(14)	28(1)	19(1)	25(1)	3(1)	6(1)	5(1)
C(15)	22(1)	14(1)	28(1)	-1(1)	7(1)	1(1)
C(16)	26(1)	33(1)	30(1)	-3(1)	-4(1)	10(1)
O(1)	18(1)	24(1)	26(1)	-4(1)	0(1)	9(1)
O(2)	22(1)	19(1)	42(1)	5(1)	11(1)	-3(1)
N(1)	16(1)	12(1)	22(1)	1(1)	6(1)	0(1)
N(2)	12(1)	15(1)	17(1)	-2(1)	3(1)	0(1)
O(3)	23(1)	39(1)	19(1)	-5(1)	4(1)	8(1)
O(4)	18(1)	16(1)	38(1)	-10(1)	3(1)	0(1)
C(17)	48(2)	39(2)	24(1)	14(1)	4(1)	6(1)

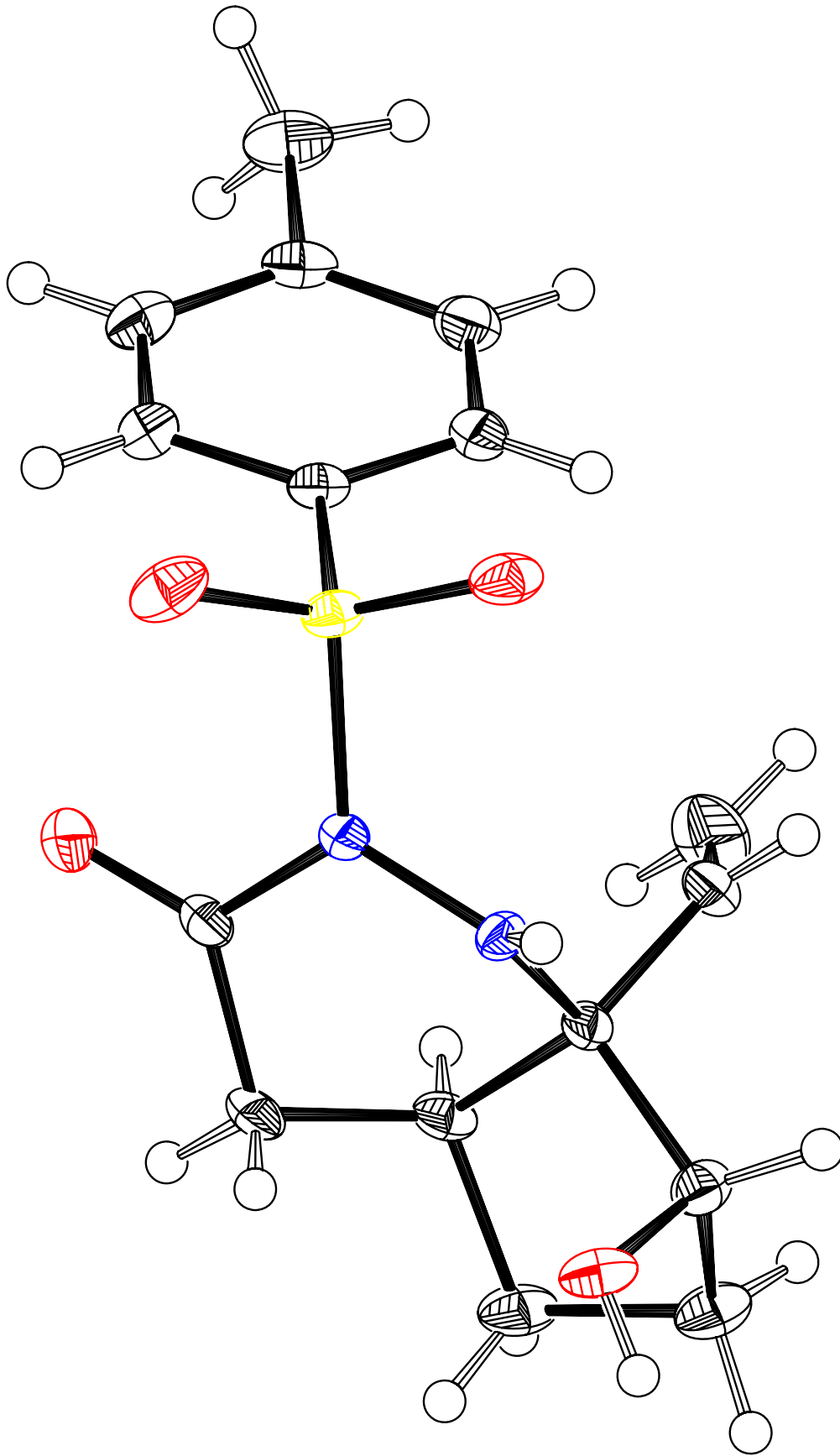
Table 5. Hydrogen coordinates ( $\times 10^4$ ) and isotropic displacement parameters ( $\text{\AA}^2 \times 10^3$ ) for p2.

	x	y	z	U(eq)
H(2)	4191	4917	1298	29
H(2B)	3705	5243	147	29
H(3)	2844	7146	667	21
H(4A)	2698	5323	2035	27
H(4B)	3536	6050	2730	27



H(7)	5156	9127	1319	24
H(8A)	5265	6243	631	30
H(8B)	4602	7396	-96	30
H(9)	3880	10992	776	28
H(11)	667	10085	2890	25
H(12)	-564	10702	1676	28
H(14)	890	13107	-106	29
H(15)	2133	12529	1108	25
H(16A)	-1073	11408	-344	46
H(16B)	-621	13002	-685	46
H(16C)	-1149	13094	234	46
H(1)	5572	6911	2417	35
H(2A)	4207	9737	2920	17
H(17A)	2663	8958	-344	45
H(17B)	2874	10846	-619	45

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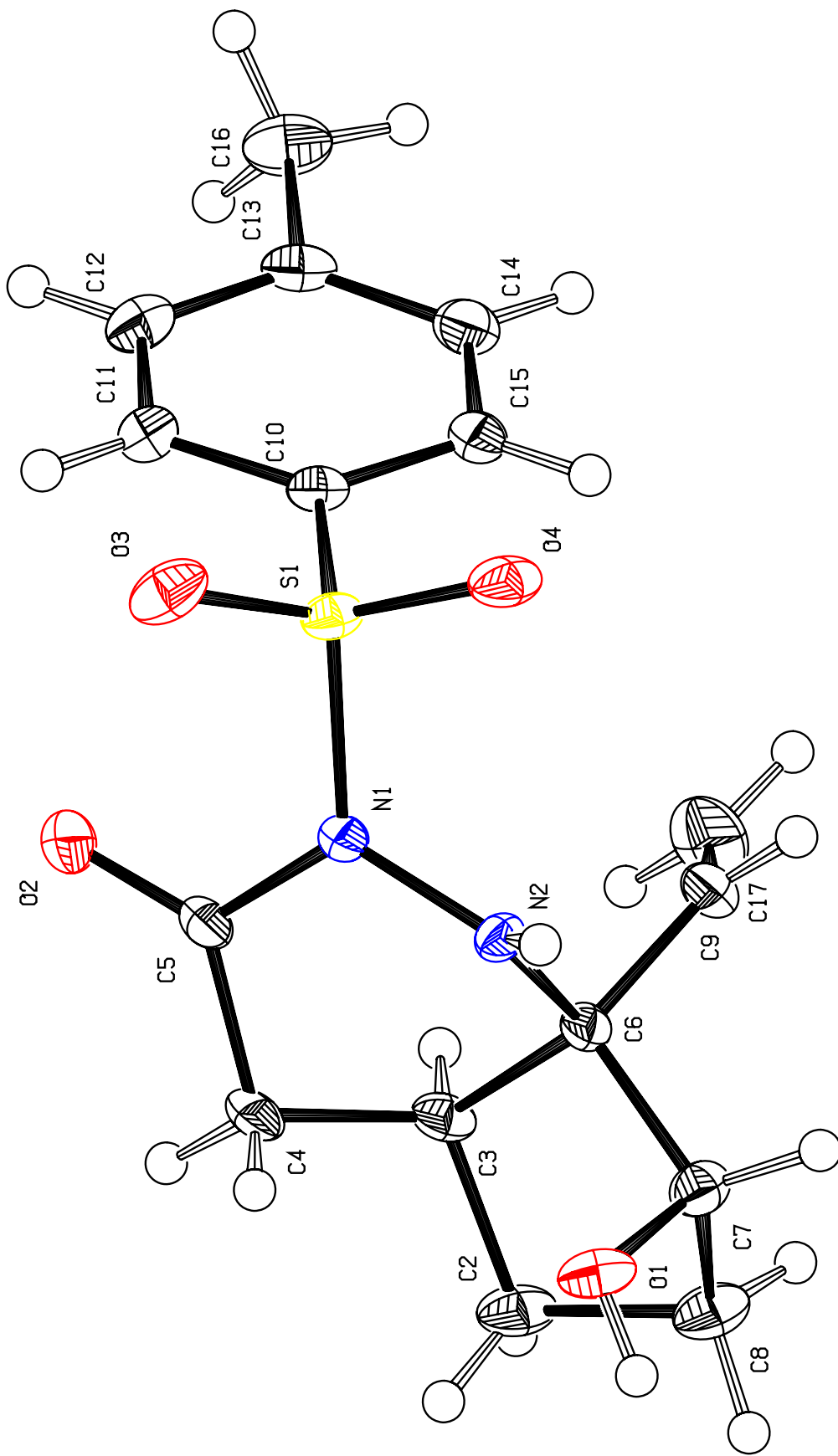


Table 1. Crystal data and structure refinement for **Kpa403**.

Identification code	<b>Kpa403</b>	
Empirical formula	C <sub>18</sub> H <sub>22</sub> N <sub>6</sub> O <sub>6</sub> S	
Formula weight	450.48	
Temperature	296 K	
Wavelength	0.71073	
Crystal system	Monoclinic	
Space group	P2(1)/c	
Unit cell dimensions	a = 13.493(7)	$\alpha$ = 90
	b = 16.902(9)	$\beta$ = 96.042(7)
	c = 9.145(5)	$\gamma$ = 90
Volume	2074.0(18)	
Z	4	
Density (calculated)	1.443 Mg/m <sup>3</sup>	
Absorption coefficient	0.205 mm <sup>-1</sup>	
F(000)	944	
Crystal size	? x ? x ? mm <sup>3</sup>	
Theta range for data collection	1.52 to 27.78	
Index ranges	-17<=h<=12, -21<=k<=21, -11<=l<=11	
Reflections collected	11416	
Independent reflections	4748 [R(int) = 0.0735]	
Completeness to theta = 27.78	96.9 %	
Absorption correction	Empirical	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	4748 / 0 / 287	
Goodness-of-fit on F <sup>2</sup>	1.014	
Final R indices [I>2sigma(I)]	R1 = 0.0660, wR2 = 0.1686	
R indices (all data)	R1 = 0.0894, wR2 = 0.1858	
Extinction coefficient	0.0000(13)	
Largest diff. peak and hole	0.581 and -0.397 e	

Table 2. Atomic coordinates ( $\times 10^4$ ) and equivalent isotropic displacement parameters ( $\text{\AA}^2 \times 10^3$ ) for p21.  $U(\text{eq})$  is defined as one third of the trace of the orthogonalized  $U^{ij}$  tensor.

	x	y	z	$U(\text{eq})$
S(1)	-1077(1)	1757(1)	3146(1)	27(1)
C(5)	1499(2)	532(1)	236(2)	26(1)
C(6)	2617(2)	685(2)	277(2)	30(1)
C(1)	502(2)	2587(1)	2303(2)	26(1)
C(7)	2723(2)	1575(2)	635(2)	29(1)
C(2)	1396(2)	2527(1)	1457(2)	28(1)
C(3)	1938(2)	1735(1)	1717(2)	28(1)
C(4)	1189(2)	1027(1)	1556(2)	26(1)
C(8)	3787(2)	1793(2)	1233(3)	38(1)
C(9)	1168(2)	511(2)	2909(2)	31(1)
C(10)	1761(2)	546(2)	4151(3)	42(1)
C(11)	3033(2)	428(2)	-1133(3)	34(1)
C(12)	-2014(2)	2331(2)	2174(2)	27(1)
C(13)	-2072(2)	3139(2)	2449(2)	31(1)
C(14)	-2766(2)	3586(2)	1594(3)	35(1)
C(15)	-3405(2)	3242(2)	465(3)	36(1)
C(16)	-3347(2)	2430(2)	237(3)	37(1)
C(17)	-2656(2)	1965(2)	1079(2)	31(1)
C(18)	-4104(3)	3748(2)	-532(3)	50(1)
O(1)	-1317(1)	940(1)	2977(2)	34(1)
O(2)	-813(1)	2094(1)	4567(2)	35(1)
O(3)	1301(2)	-285(1)	354(2)	32(1)
O(4)	4279(3)	2863(2)	2739(3)	81(1)
O(5)	3560(2)	3133(1)	614(3)	59(1)
N(1)	-65(2)	1899(1)	2215(2)	24(1)
N(2)	149(2)	1314(1)	1171(2)	24(1)
N(3)	3888(2)	2661(2)	1559(3)	42(1)
N(4)	4129(2)	373(2)	-789(3)	57(1)
N(5)	4632(2)	289(1)	-1805(2)	40(1)
N(6)	5205(2)	216(2)	-2619(3)	56(1)
O(6)	314(2)	3162(1)	3039(2)	32(1)

Table 3. Bond lengths [Å] and angles [°] for p21.

S(1)–O(1)	1.423(2)
S(1)–O(2)	1.4292(17)
S(1)–N(1)	1.700(2)
S(1)–C(12)	1.759(3)
C(5)–O(3)	1.412(3)
C(5)–C(6)	1.528(4)
C(5)–C(4)	1.561(3)
C(6)–C(11)	1.522(3)
C(6)–C(7)	1.543(3)
C(1)–O(6)	1.223(3)
C(1)–N(1)	1.389(3)
C(1)–C(2)	1.504(3)
C(7)–C(8)	1.528(4)
C(7)–C(3)	1.547(3)
C(2)–C(3)	1.533(3)
C(3)–C(4)	1.564(4)
C(4)–N(2)	1.491(3)
C(4)–C(9)	1.516(3)
C(8)–N(3)	1.500(4)
C(9)–C(10)	1.320(4)
C(11)–N(4)	1.482(4)
C(12)–C(13)	1.393(4)
C(12)–C(17)	1.398(3)
C(13)–C(14)	1.380(4)
C(14)–C(15)	1.400(4)
C(15)–C(16)	1.392(4)
C(15)–C(18)	1.507(4)
C(16)–C(17)	1.388(4)
O(4)–N(3)	1.200(3)
O(5)–N(3)	1.225(3)
N(1)–N(2)	1.426(3)
N(4)–N(5)	1.214(3)
N(5)–N(6)	1.135(3)
O(1)–S(1)–O(2)	121.13(10)
O(1)–S(1)–N(1)	105.57(10)

O(2)-S(1)-N(1)	105.22(11)
O(1)-S(1)-C(12)	109.68(12)
O(2)-S(1)-C(12)	109.72(11)
N(1)-S(1)-C(12)	103.99(11)
O(3)-C(5)-C(6)	111.0(2)
O(3)-C(5)-C(4)	113.37(18)
C(6)-C(5)-C(4)	103.57(19)
C(5)-C(6)-C(11)	112.5(2)
C(5)-C(6)-C(7)	103.8(2)
C(11)-C(6)-C(7)	115.0(2)
O(6)-C(1)-N(1)	123.5(2)
O(6)-C(1)-C(2)	124.0(2)
N(1)-C(1)-C(2)	112.4(2)
C(8)-C(7)-C(6)	111.9(2)
C(8)-C(7)-C(3)	114.2(2)
C(6)-C(7)-C(3)	104.63(19)
C(1)-C(2)-C(3)	111.98(19)
C(2)-C(3)-C(7)	113.65(19)
C(2)-C(3)-C(4)	111.2(2)
C(7)-C(3)-C(4)	106.50(19)
N(2)-C(4)-C(9)	106.31(19)
N(2)-C(4)-C(5)	108.25(18)
C(9)-C(4)-C(5)	110.66(19)
N(2)-C(4)-C(3)	110.87(19)
C(9)-C(4)-C(3)	115.50(19)
C(5)-C(4)-C(3)	105.11(19)
N(3)-C(8)-C(7)	111.7(2)
C(10)-C(9)-C(4)	128.0(3)
N(4)-C(11)-C(6)	106.7(2)
C(13)-C(12)-C(17)	121.4(2)
C(13)-C(12)-S(1)	120.07(19)
C(17)-C(12)-S(1)	118.47(19)
C(14)-C(13)-C(12)	118.8(2)
C(13)-C(14)-C(15)	121.3(3)
C(16)-C(15)-C(14)	118.7(3)
C(16)-C(15)-C(18)	120.7(3)
C(14)-C(15)-C(18)	120.5(3)
C(17)-C(16)-C(15)	121.3(3)

C(16)-C(17)-C(12)	118.5(2)
C(1)-N(1)-N(2)	118.08(19)
C(1)-N(1)-S(1)	123.73(16)
N(2)-N(1)-S(1)	117.85(16)
N(1)-N(2)-C(4)	108.61(18)
O(4)-N(3)-O(5)	122.8(3)
O(4)-N(3)-C(8)	118.6(3)
O(5)-N(3)-C(8)	118.6(2)
N(5)-N(4)-C(11)	118.1(3)
N(6)-N(5)-N(4)	171.1(3)

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Symmetry transformations used to generate equivalent atoms:



**Table 4. Anisotropic displacement parameters ( $\text{\AA}^2 \times 10^3$ ) for p21. The anisotropic displacement factor exponent takes the form:  $-2\pi^2 [ h^2 a^{*2} U^{11} + \dots + 2 h k a^* b^* U^{12} ]$**

	U <sup>11</sup>	U <sup>22</sup>	U <sup>33</sup>	U <sup>23</sup>	U <sup>13</sup>	U <sup>12</sup>
S(1)	28(1)	41(1)	14(1)	1(1)	6(1)	0(1)
C(5)	28(1)	34(1)	18(1)	0(1)	4(1)	2(1)
C(6)	28(2)	38(1)	24(1)	2(1)	4(1)	3(1)
C(1)	28(1)	36(1)	14(1)	-1(1)	1(1)	-1(1)
C(7)	26(1)	40(1)	22(1)	2(1)	6(1)	1(1)
C(2)	28(1)	35(1)	20(1)	-3(1)	5(1)	-4(1)
C(3)	28(2)	41(1)	15(1)	-1(1)	1(1)	0(1)
C(4)	26(1)	36(1)	16(1)	2(1)	4(1)	2(1)
C(8)	28(2)	46(2)	41(2)	-1(1)	4(1)	2(1)
C(9)	35(2)	38(1)	20(1)	4(1)	5(1)	2(1)
C(10)	56(2)	48(2)	21(1)	6(1)	-2(1)	-5(1)
C(11)	30(2)	44(2)	29(1)	1(1)	8(1)	4(1)
C(12)	24(1)	42(1)	15(1)	2(1)	6(1)	0(1)
C(13)	28(2)	46(2)	20(1)	-4(1)	7(1)	-2(1)
C(14)	34(2)	42(2)	31(1)	-1(1)	12(1)	5(1)
C(15)	28(2)	56(2)	24(1)	3(1)	8(1)	5(1)
C(16)	28(2)	60(2)	23(1)	-2(1)	2(1)	0(1)
C(17)	27(2)	43(1)	23(1)	-2(1)	7(1)	-2(1)
C(18)	43(2)	73(2)	34(2)	6(1)	1(1)	16(2)
O(1)	35(1)	43(1)	24(1)	4(1)	9(1)	-1(1)
O(2)	32(1)	60(1)	13(1)	-1(1)	7(1)	1(1)
O(3)	37(1)	38(1)	22(1)	0(1)	1(1)	-2(1)
O(4)	112(3)	72(2)	54(2)	-16(1)	-9(2)	-11(2)
O(5)	42(2)	51(1)	80(2)	9(1)	-6(1)	-4(1)
N(1)	23(1)	36(1)	15(1)	-2(1)	7(1)	1(1)
N(2)	27(1)	33(1)	13(1)	-1(1)	4(1)	2(1)
N(3)	29(1)	52(2)	46(1)	-5(1)	9(1)	-4(1)
N(4)	36(2)	103(2)	33(1)	5(1)	11(1)	21(2)
N(5)	37(2)	53(2)	32(1)	-1(1)	6(1)	4(1)
N(6)	39(2)	89(2)	41(1)	-1(1)	14(1)	1(2)
O(6)	36(1)	41(1)	19(1)	-6(1)	7(1)	-2(1)

Table 5. Hydrogen coordinates ( $\times 10^4$ ) and isotropic displacement parameters ( $\text{\AA}^2 \times 10^3$ ) for p21.

	x	y	z	U(eq)
H(5)	1157	737	-685	32
H(6)	2959	385	1098	36
H(7)	2540	1877	-268	35
H(2A)	1853	2955	1750	33
H(2B)	1185	2587	416	33
H(3)	2276	1732	2720	34
H(8A)	3977	1495	2125	46
H(8B)	4238	1649	517	46
H(9)	676	124	2858	37
H(10A)	2266	922	4268	50
H(10B)	1673	194	4909	50
H(11A)	2760	-81	-1456	41
H(11B)	2865	812	-1907	41
H(13)	-1651	3374	3196	37
H(14)	-2811	4126	1771	42
H(16)	-3780	2194	-493	44
H(17)	-2621	1423	918	37
H(18A)	-4021	3630	-1539	75
H(18B)	-3956	4297	-340	75
H(18C)	-4779	3641	-352	75
H(3A)	942	-431	-376	48
H(3)	80(30)	1576(17)	300(40)	46(9)

Table 6. Torsion angles [°] for p21.

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