

Supplementary Table S1

Vertebrate Pigmentation Genes

no.	gene	mouse coat color mutant	teleost pigment. mutants	FSGD ¹	LSGD ²	References
a) melanophore development (FSGD duplicate retention ratio: 22/45 = 48.9%)						
1	<i>Adam17</i>			■		1
2	<i>Adamts20</i>	<i>belted</i>				1
3	<i>Apc</i>					1
4	<i>Creb1</i> *			■	◇	2
5	<i>Eda</i>	<i>Tabby</i>				1
6	<i>Edn3</i> *	<i>lethal spotting</i>		■		1
7	<i>Ednrb1</i> *	<i>piebald spotting</i>	<i>rose (ednrb1a)^{Dre}</i>	■		1, 3, 4
8	<i>Ednrb2</i>					5
9	<i>Ece1</i>					1
10	<i>Egfr</i>	<i>dark skin 5</i>		■		1, 6
11	<i>En1</i>			■		1
12	<i>ErbB3</i>		<i>picasso (erbb3b)^{Dre}</i>	■		3, 7
13	<i>Fgfr2</i>					1
14	<i>Foxd3</i> *		<i>mother superior^{Dre}</i>			3, 8
15	<i>Frem2</i>			■		1
16	<i>Fzd4</i>					1
17	<i>Gnaq</i>	<i>dark skin 1, dark skin 10</i>		■	◇	1
18	<i>Gna11</i>	<i>dark skin7</i>		■		1
19	<i>Gpc3</i>					1
20	<i>Gpr161</i>					1
21	<i>Hdac1</i>		<i>colgate (hdac1a)^{Dre}</i>	■		3, 9
22	<i>Ikkkg</i>					1
23	<i>Itgb1</i>			■	◇	1
24	<i>Kit</i>	<i>dominant white-spotting</i>	<i>sparse (kita)^{Dre}</i>	■		1, 3, 10
25	<i>Kitl</i>	<i>steel</i>		■		1, 11
26	<i>Lef1</i>					1
27	<i>Lmx1a</i>	<i>dreher</i>				1
28	<i>Mbtps1</i>					1
29	<i>Mcoln3</i>	<i>varitint-waddler</i>		■		1
30	<i>Mitf</i>	<i>microphthalmia</i>	<i>nacre (mitfa)^{Dre}</i>	■		1, 3, 12
31	<i>Mreg</i>	<i>dilute supressor</i>				1
32	<i>Myc</i>			■		1
33	<i>Pax3</i> *	<i>splotch</i>		■		1, 13
34	<i>Scarb2</i>		<i>hi1463^{Dre}</i>			3, 14
35	<i>Sfxn1</i>	<i>flexed tail</i>				1
36	<i>Snai2</i>					1
37	<i>Sox9</i> *		<i>b971 (sox9b)^{Dre}</i>	■		3, 15, 16
38	<i>Sox10</i> *	<i>Dominant megacolon</i>	<i>colourless (sox10b)^{Dre}</i>	■		1, 3, 17
39	<i>Sox18</i>	<i>ragged</i>	-			1
40	<i>Tfp2a</i>	-	<i>lockjaw^{Dre}</i>			1, 3, 18
41	<i>Trmp1</i>	-		■		19

42	<i>Trmp7</i>	-	<i>touchtone</i> ^{Dre}		3, 19
43	<i>Wnt1</i>	-	-		1
44	<i>Wnt3a</i>	-	-		1
45	<i>Zic2</i>	<i>Kumba</i>	-	■	1
b) components of melanosomes (FSGD duplicate retention ratio: 6/10 = 60.0%)					
46	<i>Dct</i>	<i>slaty</i>	-		1
47	<i>Gpnmb</i>	<i>iris pigment dispersion</i>	-		1
48	<i>Rab38</i>	<i>chocolate</i>	-	■	1
49	<i>Rab32</i>	-	-	■	20
50	<i>Silv</i>	<i>silver</i>	<i>fading vision (silva)</i> ^{Dre}	■	1, 3, 21
51	<i>Slc24a4</i>	-	-	■	22
52	<i>Slc24a5</i>	-	<i>golden</i> ^{Dre}		3, 23
53	<i>Slc45a2</i>	<i>underwhite</i>	<i>b</i> ^{Ola}		1, 24
54	<i>Tyr</i>	<i>albino</i>	<i>sandy (tyra)</i> ^{Dre} ; <i>i (tyra)</i> ^{Ola}	■	1, 3, 25, 26
55	<i>Tyrp1</i>	<i>brown</i>	-	■	1
c) melanosome construction (FSGD duplicate retention ratio: 2/22 = 9.1%)					
56	<i>Ap3b1</i>	<i>pearl</i>		◇	1
57	<i>Ap3d1</i>	<i>mocha</i>			1
58	<i>Bloc1s3</i>	<i>reduced pigmentation</i>			1
59	<i>Cno</i>	<i>cappuchino</i>			1
60	<i>Dtnbp1</i>	<i>sandy</i>		■	1
61	<i>Fig4</i>	<i>pale temor</i>			1
62	<i>Gpr143</i>	<i>Oa1</i>			1
63	<i>Hps1</i>	<i>pale ear</i>			1
64	<i>Hps3</i>	<i>cocoa</i>			1
65	<i>Hps4</i>	<i>light ear</i>			1
66	<i>Hps5</i>	<i>Ruby-eye 2</i>			1
67	<i>Hps6</i>	<i>Ruby-eye</i>			1
68	<i>Lyst</i>	<i>beige</i>		◇	1
69	<i>Nsf</i>		<i>hi2869 (nsfb)</i> ^{Dre}	■	3
70	<i>Oca2</i>	<i>pinkeyed-dilution</i>	<i>i-3</i> ^{Ola}		1, 27
71	<i>Pldn</i>	<i>pallid</i>			1
72	<i>Rabggta</i>	<i>gunmetal</i>			1
73	<i>Txndc5</i>	<i>muted</i>			1
74	<i>Vps33a</i>	<i>buff</i>			1
75	<i>Vps11</i>		<i>pale grey eyes</i> ^{Ola}		28
76	<i>Vps18*</i>		<i>vps18</i> ^{Dre}		3, 29
77	<i>Vps39*</i>		<i>lbk/vam6</i> ^{Dre}		3, 30
d) melanosome transport (retention ratio : 3/4 = 75.0%)					
78	<i>Mlph</i>	<i>leaden</i>	<i>j120</i> ^{Dre}	■	1, 3, 31
79	<i>Myo5a</i>	<i>dilute</i>		■	1
80	<i>Myo7a</i>	<i>shaker-1</i>		■	1
81	<i>Rab27a</i>	<i>ashen</i>			1
e) regulation of melanogenesis (FSGD duplicate retention ratio: 3/7 = 42.9%)					
82	<i>Asip</i>	<i>nonagouti</i>			1
83	<i>Atrn</i>	<i>mahogany</i>			1
4	<i>Creb1*</i>			■ ◇	2
84	<i>Drd2</i>				1
85	<i>Mc1r</i>	<i>extension</i>			1
86	<i>Mgrn1</i>	<i>Mahoganoid</i>		■	1
87	<i>Pomc</i>			■	1
f) systemic effects (FSGD duplicate retention ratio: 2/13 = 15.4%)					
88	<i>Atox1</i>				1
89	<i>Atp7a</i>	<i>mottled</i>	<i>calamity</i> ^{Dre}		1, 3
90	<i>Atp7b</i>	<i>toxic milk</i>			1

91	<i>Atp6ap1</i>		<i>bleached blond</i> ^{Dre}		◇	3, 14
92	<i>Atp6ap2</i>		<i>hi3681</i> ^{Dre}			3
93	<i>Atp6v0c</i>		<i>hi1207 (atp6v0c a)</i> ^{Dre}	■		3, 14
94	<i>Atp6v0d1</i>		<i>hi2188b</i> ^{Dre}			3
95	<i>Atp6v1e1*</i>		<i>hi577a (atp6v1e1a)</i> ^{Dre}	■		3, 14
96	<i>Atp6v1f</i>		<i>hi1988</i> ^{Dre}		◇	3
97	<i>Atp6v1h*</i>		<i>hi923</i>			3, 14
98	<i>Rpl24</i>	<i>belly spot and tail</i>				1
99	<i>Rps19</i>	<i>dark skin 3</i>				1
100	<i>Rps20</i>	<i>dark skin 4</i>				1

g) xanthophore development (retention rate: 9/11 = 81.8%)

95	<i>Atp6v1e1*</i>		<i>hi577a (atp6v1e1a)</i> ^{Dre}	■		3, 14
97	<i>Atp6v1h*</i>		<i>hi923</i>			3, 14
101	<i>Csf1r</i>		<i>panther (csf1ra)</i> ^{Dre}	■		3, 32
6	<i>Edn3*</i>	<i>lethal spotting</i>		■		1
7	<i>Ednrb1*</i>	<i>piebald spotting</i>	<i>rose (ednrb1a)</i> ^{Dre}	■		1, 3, 444
14	<i>Foxd3*</i>		<i>mother superior</i> ^{Dre}			3, 8
102	<i>Ghr</i>			■		33
33	<i>Pax3*</i>	<i>splotch</i>		■		1, 13
103	<i>Pax7</i>			■		1, 13
104	<i>Smtl</i>		<i>color interfere</i> ^{Ola}	■		34
38	<i>Sox10*</i>	<i>Dominant megacolon</i>	<i>colourless (sox10b)</i> ^{Dre}	■		1, 3, 17

h) pteridine synthesis (FSGD duplicate retention rate: 2/11 = 18.2%)

105	<i>Gch1a</i>					35
106	<i>Gch1b</i>					3, 35, 36
107	<i>Gchfr</i>					35
108	<i>Mycbp2</i>		<i>esrom</i> ^{Dre}			3, 37
109	<i>Paics</i>		<i>hi2688</i> ^{Dre}			3
110	<i>Pcbd1</i>					35
111	<i>Pcbd2</i>					35
112	<i>Pts</i>					35
113	<i>Qdpr</i>			■	◇	35
114	<i>Spr</i>			■		35
115	<i>Xdh</i>					3, 35

i) iridophore development (FSGD duplicate retention rate: 4/11 = 36.4%)

97	<i>Atp6v1h*</i>		<i>hi923</i>			3, 14
116	<i>Dac</i>		<i>hagoromo</i> ^{Dre}			38
6	<i>Edn3*</i>	<i>lethal spotting</i>		■		1
7	<i>Ednrb1*</i>	<i>piebald spotting</i>	<i>rose (ednrb1a)</i> ^{Dre}	■		1, 3, 4
14	<i>Foxd3*</i>		<i>mother superior</i> ^{Dre}			3, 8
117	<i>Ltk</i>		<i>shady</i> ^{Dre}			39
37	<i>Sox9*</i>		<i>b971 (sox9b)</i> ^{Dre}	■		3, 15, 16
38	<i>Sox10*</i>	<i>Dominant megacolon</i>	<i>colourless (sox10b)</i> ^{Dre}	■		1, 3, 17
118	<i>Trim33</i>		<i>moonshine</i> ^{Dre}			3
76	<i>Vps18*</i>		<i>vps18</i> ^{Dre}			3, 29
77	<i>Vps39*</i>		<i>lbk/vam6</i> ^{Dre}			3, 30

j) uncategorized function (FSGD duplicate retention rate: 3/10 = 30.0%)

119	<i>Abhd11</i>		<i>hi3305</i> ^{Dre}			3
120	<i>Atoh7</i>		<i>lakritz</i> ^{Dre}			3, 40
121	<i>Ebna1bp2</i>		<i>hi3625a</i> ^{Dre}			3
122	<i>Gfpt1</i>		<i>eartha</i> ^{Dre}			3
123	<i>Gja5</i>		<i>leopard</i> ^{Dre}	■		3, 41
124	<i>Irf4</i>			■	◇	42
125	<i>Kcnj13</i>		<i>jaguar</i> ^{Dre}			3, 43
126	<i>Pabpc1</i>		<i>hi3202b (pabpc1a)</i> ^{Dre}	■		3
127	<i>Skiv2l2</i>		<i>julie</i> ^{Dre}			3
128	<i>Tpcn2</i>					44

* gene belongs to more than one category

^{Dre} *Danio rerio* (zebrafish); ^{Ola} *Oryzias latipes* (medaka)

¹ ■ indicates that a FSGD duplicate has been retained in teleosts

² ◇ indicates that a lineage-specific gene duplication (LSGD) has occurred in one or more teleosts

Literature Cited

- 1) Mouse Coat Color Genes (<http://www.espcr.org/micemut/>).
- 2) Lin, J.Y. and D.E. Fisher. 2007. Melanocyte biology and skin pigmentation. *Nature* **445**: 843-850.
- 3) Zebrafish Model Organism Database (ZFIN; <http://zfin.org>).
- 4) Parichy, D.M., E.M. Mellgren, J.F. Rawls, S.S. Lopes, R.N. Kelsh, and S.L. Johnson. 2000. Mutational analysis of endothelin receptor b1 (rose) during neural crest and pigment pattern development in the zebrafish *Danio rerio*. *Dev Biol* **227**: 294-306.
- 5) Miwa, M., M. Inoue-Murayama, H. Aoki, T. Kunisada, T. Hiragaki, M. Mizutani, and S. Ito. 2007. Endothelin receptor B2 (EDNRB2) is associated with the panda plumage colour mutation in Japanese quail. *Anim Genet* **38**: 103-108.
- 6) Meierjohann, S. and M. Schartl. 2006. From Mendelian to molecular genetics: the *Xiphophorus* melanoma model. *Trends Genet* **22**: 654-661.
- 7) Budi, E.H., L.B. Patterson, and D.M. Parichy. 2008. Embryonic requirements for ErbB signaling in neural crest development and adult pigment pattern formation. *Development* **135**: 2603-2614.
- 8) Montero-Balaguer, M., M.R. Lang, S.W. Sachdev, C. Knappmeyer, R.A. Stewart, A. De La Guardia, A.K. Hatzopoulos, and E.W. Knapik. 2006. The mother superior mutation ablates *foxd3* activity in neural crest progenitor cells and depletes neural crest derivatives in zebrafish. *Dev Dyn* **235**: 3199-3212.
- 9) Ignatius, M.S., H.E. Moose, H.M. El-Hodiri, and P.D. Henion. 2008. *colgate/hdac1* Repression of *foxd3* expression is required to permit *mitfa*-dependent melanogenesis. *Dev Biol* **313**: 568-583.
- 10) Parichy, D.M., J.F. Rawls, S.J. Pratt, T.T. Whitfield, and S.L. Johnson. 1999. Zebrafish *sparse* corresponds to an orthologue of *c-kit* and is required for the morphogenesis of a subpopulation of melanocytes, but is not essential for hematopoiesis or primordial germ cell development. *Development* **126**: 3425-3436.
- 11) Hultman, K.A., N. Bahary, L.I. Zon, and S.L. Johnson. 2007. Gene Duplication of the zebrafish *kit* ligand and partitioning of melanocyte development functions to *kit* ligand a. *PLoS Genet* **3**: e17.
- 12) Lister, J.A., C.P. Robertson, T. Lepage, S.L. Johnson, and D.W. Raible. 1999. *nacre* encodes a zebrafish microphthalmia-related protein that regulates neural-crest-derived pigment cell fate. *Development* **126**: 3757-3767.
- 13) Minchin, J.E. and S.M. Hughes. 2008. Sequential actions of *Pax3* and *Pax7* drive xanthophore development in zebrafish neural crest. *Dev Biol* **317**: 508-522.
- 14) Pickart, M.A., S. Sivasubbu, A.L. Nielsen, S. Shriram, R.A. King, and S.C. Ekker. 2004. Functional genomics tools for the analysis of zebrafish pigment. *Pigment Cell Res* **17**: 461-470.
- 15) Passeron, T., J.C. Valencia, C. Bertolotto, T. Hoashi, E. Le Pape, K. Takahashi, R. Ballotti, and V.J. Hearing. 2007. *SOX9* is a key player in ultraviolet B-induced melanocyte differentiation and pigmentation. *Proc Natl Acad Sci U S A* **104**: 13984-13989.

- 16) Yan, Y.L., J. Willoughby, D. Liu, J.G. Crump, C. Wilson, C.T. Miller, A. Singer, C. Kimmel, M. Westerfield, and J.H. Postlethwait. 2005. A pair of Sox: distinct and overlapping functions of zebrafish sox9 co-orthologs in craniofacial and pectoral fin development. *Development* **132**: 1069-1083.
- 17) Dutton, K.A., A. Pauliny, S.S. Lopes, S. Elworthy, T.J. Carney, J. Rauch, R. Geisler, P. Haffter, and R.N. Kelsh. 2001. Zebrafish colourless encodes sox10 and specifies non-ectomesenchymal neural crest fates. *Development* **128**: 4113-4125.
- 18) Knight, R.D., S. Nair, S.S. Nelson, A. Afshar, Y. Javidan, R. Geisler, G.J. Rauch, and T.F. Schilling. 2003. lockjaw encodes a zebrafish tfap2a required for early neural crest development. *Development* **130**: 5755-5768.
- 19) McNeill, M.S., J. Paulsen, G. Bonde, E. Burnight, M.Y. Hsu, and R.A. Cornell. 2007. Cell death of melanophores in zebrafish trpm7 mutant embryos depends on melanin synthesis. *J Invest Dermatol* **127**: 2020-2030.
- 20) Wasmeier, C., M. Romao, L. Plowright, D.C. Bennett, G. Raposo, and M.C. Seabra. 2006. Rab38 and Rab32 control post-Golgi trafficking of melanogenic enzymes. *J Cell Biol* **175**: 271-281.
- 21) Schonhaler, H.B., J.M. Lampert, J. von Lintig, H. Schwarz, R. Geisler, and S.C. Neuhaus. 2005. A mutation in the silver gene leads to defects in melanosome biogenesis and alterations in the visual system in the zebrafish mutant fading vision. *Dev Biol* **284**: 421-436.
- 22) Sulem, P., D.F. Gudbjartsson, S.N. Stacey, A. Helgason, T. Rafnar, K.P. Magnusson, A. Manolescu, A. Karason, A. Palsson, G. Thorleifsson, M. Jakobsdottir, S. Steinberg, S. Palsson, F. Jonasson, B. Sigurgeirsson, K. Thorisdottir, R. Ragnarsson, K.R. Benediksdottir, K.K. Aben, L.A. Kiemenev, J.H. Olafsson, J. Gulcher, A. Kong, U. Thorsteinsdottir, and K. Stefansson. 2007. Genetic determinants of hair, eye and skin pigmentation in Europeans. *Nat Genet* **39**: 1443-1452.
- 23) Lamason, R.L., M.A. Mohideen, J.R. Mest, A.C. Wong, H.L. Norton, M.C. Aros, M.J. Juryneec, X. Mao, V.R. Humphreville, J.E. Humbert, S. Sinha, J.L. Moore, P. Jagadeeswaran, W. Zhao, G. Ning, I. Makalowska, P.M. McKeigue, D. O'Donnell, R. Kittles, E.J. Parra, N.J. Mangini, D.J. Grunwald, M.D. Shriver, V.A. Canfield, and K.C. Cheng. 2005. SLC24A5, a putative cation exchanger, affects pigmentation in zebrafish and humans. *Science* **310**: 1782-1786.
- 24) Fukamachi, S., A. Shimada, and A. Shima. 2001. Mutations in the gene encoding B, a novel transporter protein, reduce melanin content in medaka. *Nat Genet* **28**: 381-385.
- 25) Koga, A., H. Inagaki, Y. Bessho, and H. Hori. 1995. Insertion of a novel transposable element in the tyrosinase gene is responsible for an albino mutation in the medaka fish, *Oryzias latipes*. *Mol Gen Genet* **249**: 400-405.
- 26) Page-McCaw, P.S., S.C. Chung, A. Muto, T. Roeser, W. Staub, K.C. Finger-Baier, J.I. Korenbrot, and H. Baier. 2004. Retinal network adaptation to bright light requires tyrosinase. *Nat Neurosci* **7**: 1329-1336.
- 27) Fukamachi, S., S. Asakawa, Y. Wakamatsu, N. Shimizu, H. Mitani, and A. Shima. 2004. Conserved function of medaka pink-eyed dilution in melanin synthesis and its divergent transcriptional regulation in gonads among vertebrates. *Genetics* **168**: 1519-1527.
- 28) Yu, J.F., S. Fukamachi, H. Mitani, H. Hori, and A. Kanamori. 2006. Reduced expression of vps11 causes less pigmentation in medaka, *Oryzias latipes*. *Pigment Cell Res* **19**: 628-634.
- 29) Maldonado, E., F. Hernandez, C. Lozano, M.E. Castro, and R.E. Navarro. 2006. The zebrafish mutant vps18 as a model for vesicle-traffic related hypopigmentation diseases. *Pigment Cell Res* **19**: 315-326.

- 30) Schonthaler, H.B., V.C. Fleisch, O. Biehlmaier, Y. Makhankov, O. Rinner, R. Bahadori, R. Geisler, H. Schwarz, S.C. Neuhauss, and R. Dahm. 2008. The zebrafish mutant *lbk/vam6* resembles human multisystemic disorders caused by aberrant trafficking of endosomal vesicles. *Development* **135**: 387-399.
- 31) Sheets, L., D.G. Ransom, E.M. Mellgren, S.L. Johnson, and B.J. Schnapp. 2007. Zebrafish melanophilin facilitates melanosome dispersion by regulating dynein. *Curr Biol* **17**: 1721-1734.
- 32) Parichy, D.M., D.G. Ransom, B. Paw, L.I. Zon, and S.L. Johnson. 2000. An orthologue of the kit-related gene *fms* is required for development of neural crest-derived xanthophores and a subpopulation of adult melanocytes in the zebrafish, *Danio rerio*. *Development* **127**: 3031-3044.
- 33) Fukamachi, S. and A. Meyer. 2007. Evolution of receptors for growth hormone and somatotactin in fish and land vertebrates: lessons from the lungfish and sturgeon orthologues. *J Mol Evol* **65**: 359-372.
- 34) Fukamachi, S., M. Sugimoto, H. Mitani, and A. Shima. 2004b. Somatotactin selectively regulates proliferation and morphogenesis of neural-crest derived pigment cells in medaka. *Proc Natl Acad Sci U S A* **101**: 10661-10666.
- 35) Braasch, I., M. Scharl, and J.N. Volff. 2007. Evolution of pigment synthesis pathways by gene and genome duplication in fish. *BMC Evol Biol* **7**: 74.
- 36) Pelletier, I., L. Bally-Cuif, and I. Ziegler. 2001. Cloning and developmental expression of zebrafish GTP cyclohydrolase I. *Mech Dev* **109**: 99-103.
- 37) Le Guyader, S., J. Maier, and S. Jesuthasan. 2005. Esrom, an ortholog of PAM (protein associated with c-myc), regulates pteridine synthesis in the zebrafish. *Dev Biol* **277**: 378-386.
- 38) Kawakami, K., A. Amsterdam, N. Shimoda, T. Becker, J. Mugg, A. Shima, and N. Hopkins. 2000. Proviral insertions in the zebrafish *hagoromo* gene, encoding an F-box/WD40-repeat protein, cause stripe pattern anomalies. *Curr Biol* **10**: 463-466.
- 39) Lopes, S.S., X. Yang, J. Muller, T.J. Carney, A.R. McAdow, G.J. Rauch, A.S. Jacoby, L.D. Hurst, M. Delfino-Machin, P. Haffter, R. Geisler, S.L. Johnson, A. Ward, and R.N. Kelsh. 2008. Leukocyte tyrosine kinase functions in pigment cell development. *PLoS Genet* **4**: e1000026.
- 40) Kelsh, R.N., M. Brand, Y.J. Jiang, C.P. Heisenberg, S. Lin, P. Haffter, J. Odenthal, M.C. Mullins, F.J. van Eeden, M. Furutani-Seiki, M. Granato, M. Hammerschmidt, D.A. Kane, R.M. Warga, D. Beuchle, L. Vogelsang, and C. Nusslein-Volhard. 1996. Zebrafish pigmentation mutations and the processes of neural crest development. *Development* **123**: 369-389.
- 41) Watanabe, M., M. Iwashita, M. Ishii, Y. Kurachi, A. Kawakami, S. Kondo, and N. Okada. 2006. Spot pattern of leopard *Danio* is caused by mutation in the zebrafish *connexin41.8* gene. *EMBO Rep* **7**: 893-897.
- 42) Han, J., P. Kraft, H. Nan, Q. Guo, C. Chen, A. Qureshi, S.E. Hankinson, F.B. Hu, D.L. Duffy, Z.Z. Zhao, N.G. Martin, G.W. Montgomery, N.K. Hayward, G. Thomas, R.N. Hoover, S. Chanock, and D.J. Hunter. 2008. A genome-wide association study identifies novel alleles associated with hair color and skin pigmentation. *PLoS Genet* **4**: e1000074.
- 43) Iwashita, M., M. Watanabe, M. Ishii, T. Chen, S.L. Johnson, Y. Kurachi, N. Okada, and S. Kondo. 2006. Pigment pattern in jaguar/obelix zebrafish is caused by a *Kir7.1* mutation: implications for the regulation of melanosome movement. *PLoS Genet* **2**: e197.
- 44) Sulem, P., D.F. Gudbjartsson, S.N. Stacey, A. Helgason, T. Rafnar, M. Jakobsdottir, S. Steinberg, S.A. Gudjonsson, A. Palsson, G. Thorleifsson, S. Palsson, B. Sigurgeirsson, K. Thorisdottir, R. Ragnarsson,

K.R. Benediksdottir, K.K. Aben, S.H. Vermeulen, A.M. Goldstein, M.A. Tucker, L.A. Kiemeny, J.H. Olafsson, J. Gulcher, A. Kong, U. Thorsteinsdottir, and K. Stefansson. 2008. Two newly identified genetic determinants of pigmentation in Europeans. *Nat Genet* **40**: 835-837.