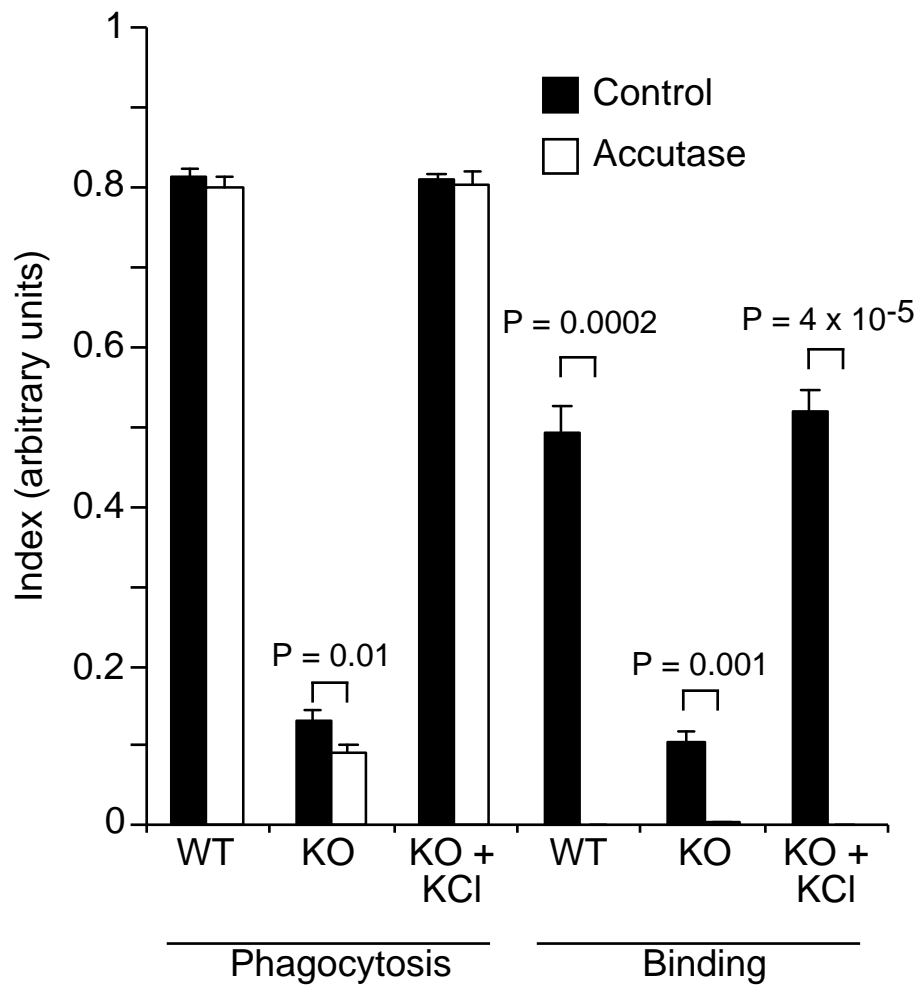
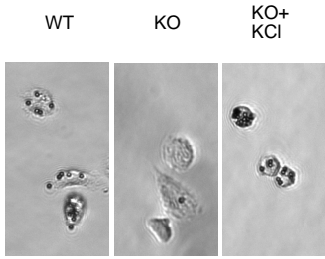


Link et al. Supplementary Figure 1

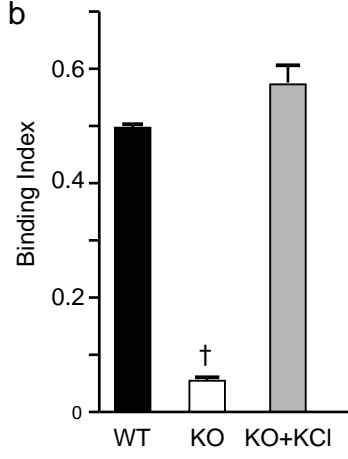


Link et al. Supplementary Fig. 2

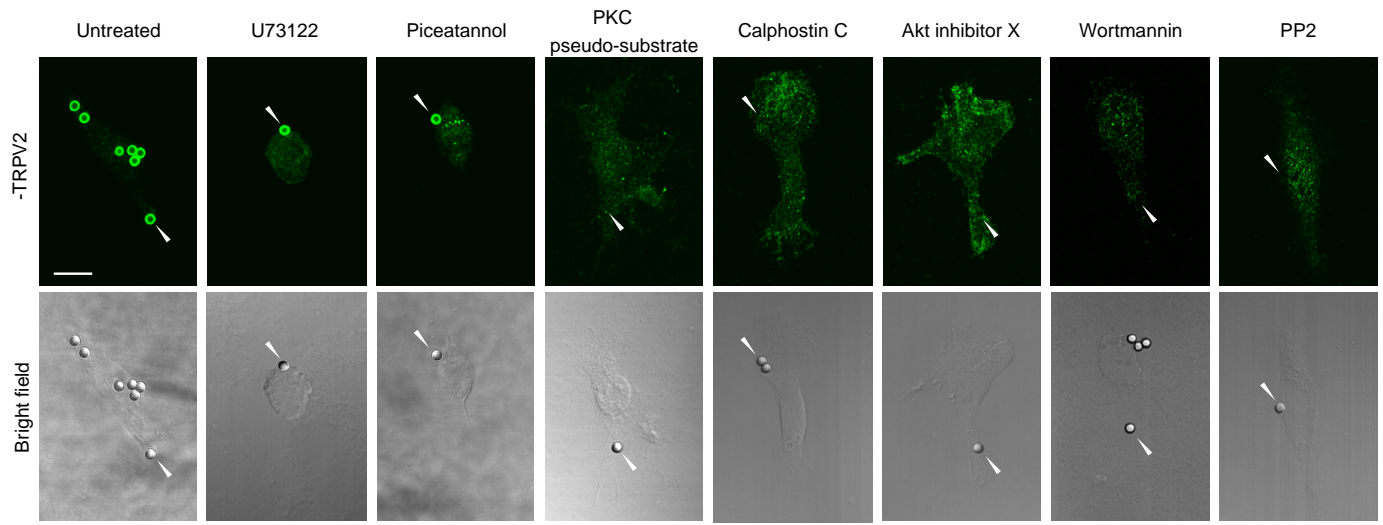
a



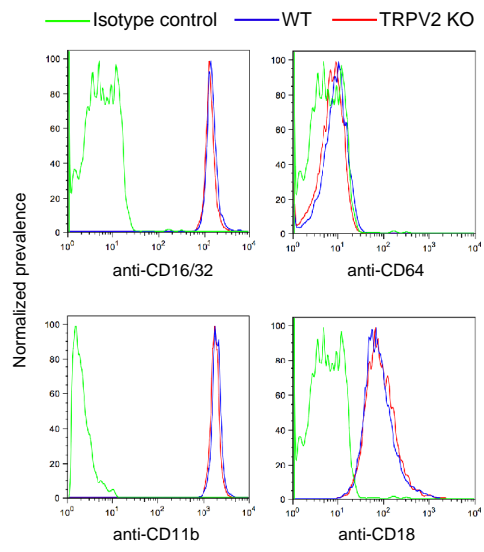
b



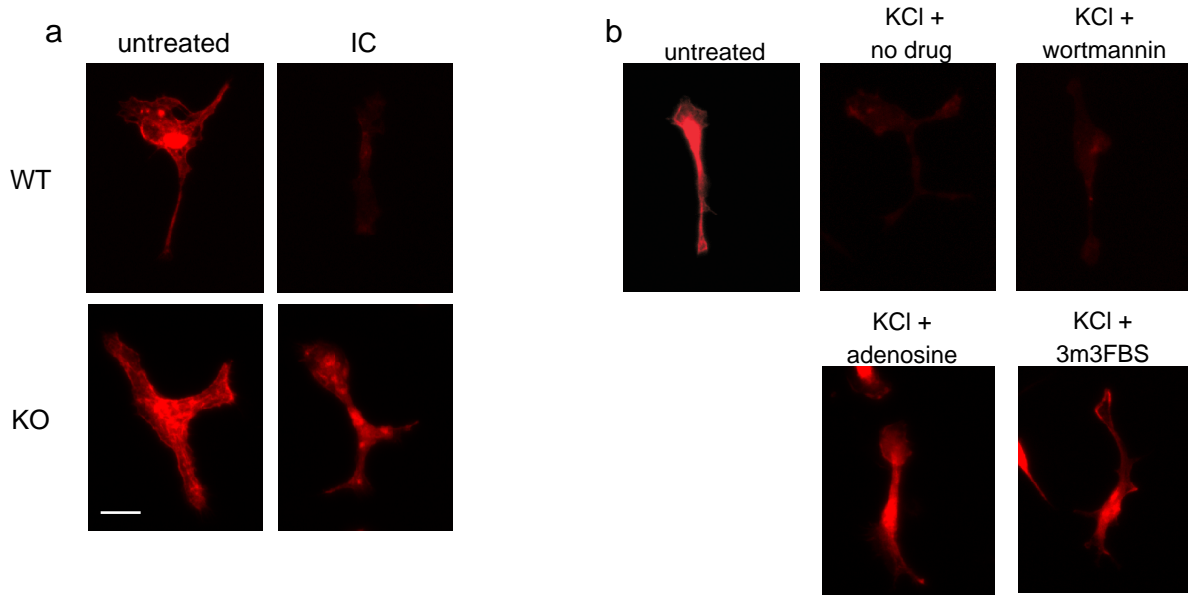
Link et al. Supplementary Figure 3



Link et al. Supplementary Fig. 4



Link et al. Supplementary Figure 5



Link et al. Supplementary Figure 6

## SUPPLEMENTARY FIGURE LEGENDS

### **Supplementary Figure 1:** Flow cytometric analysis of peritoneal macrophages. (a)

Scatter plots from flow cytometric examination of resident peritoneal cells from a representative wild-type and TRPV2KO mouse pair. Percent macrophages are indicated in gated regions. b. Quantification of cell types in peritoneal lavage samples from each genotype. Mean  $\pm$  SEM; wild-type,  $n = 10$ ; TRPV2KO,  $n = 9$ . Macrophages were defined as F4/80 and CD11b high/Gr1, CD3, and B220 negative. Lymphocytes were defined as CD3 or B220 positive. Neutrophils were defined as Gr1 high/ CD11b medium. c. Actin staining of wild-type and TRPV2KO peritoneal macrophages with rhodamine-phalloidin. Scale bar, 20  $\mu$ m.

**Supplementary Figure 2:** Discrimination between internalized versus noninternalized particles during phagocytosis and binding. Following phagocytosis (5 min) or binding (5 min) in the presence of 10  $\mu$ M cytochalasin D of IgG-coated latex beads, binding or phagocytosis index was quantified in wild-type, TRPV2KO, or KCl-treated TRPV2KO macrophages before (black bars) and after (open bars) incubation with Accutase (100%, Sigma, 15 min, 37°C) to remove noninternalized particles. Mean  $\pm$  SEM,  $n = 3$  wells per condition per genotype. Note that Accutase treatment removes nearly all particles incubated under binding conditions, but spares most particles incubated under phagocytosis conditions in wild-type mice or in TRPV2KO cells treated with KCl. There is some reduction in particles associated with TRPV2KO cells even under phagocytosis conditions, suggesting that not all phagosomes had closed.

**Supplementary Figure 3:** Defective phagocytosis in TRPV2 deficient BMM and rescue by KCl. a. Representative photomicrographs of wild-type, TRPV2KO, and KCl-treated TRPV2KO BMMs following 5 min phagocytosis of IgG-coated latex beads (2  $\mu$ m, left). Wild-type and TRPV2KO photos show cells exposed to beads under control conditions. KO + KCl, TRPV2KO cells were exposed to beads with KCl (50 mM) added to the medium. b. Corresponding phagocytic indices. Mean  $\pm$  SEM,  $n = 3$  mice per genotype, each assayed in duplicate. †  $P < 10^{-4}$ .

**Supplementary Figure 4:** Pharmacological inhibition of TRPV2 phagosomal recruitment. Wild-type macrophages are shown after 5 min phagocytosis of IgG-coated beads (arrowheads), without drugs or in the presence of PLC inhibitor U73122 (10  $\mu$ M), Syk kinase inhibitor piceatannol (20  $\mu$ M), PKC  $\zeta$  pseudosubstrate (40  $\mu$ M), general PKC inhibitor calphostin C (500 nM), Akt inhibitor X (10  $\mu$ M), PI3 kinase inhibitor wortmannin (100 nM), or Src kinase inhibitor PP2 (1  $\mu$ M). Top, TRPV2 immunofluorescence. Bottom, brightfield images. Scale bar, 8  $\mu$ m.

**Supplementary Figure 5:** Phagocyte receptor expression levels are similar between wildtype and TRPV2 deficient macrophages. Representative histograms from flow cytometric data of wild-type macrophages (F4/80 positive, CD11b positive, CD3 negative, B220 negative) in acutely isolated peritoneal lavage. Histograms compare wild-type (blue traces) and TRPV2KO (red traces) macrophages and isotype controls (green traces), with respect to intensity of surface binding by antibodies against Fc $\gamma$



receptors (anti-CD16/32, anti-CD64) (6) and complement C3 receptor (anti-CD11b, anti-CD18) (7).

**Supplementary Figure 6: IC-induced actin depolymerization.** a. Representative photomicrographs of wild-type and TRPV2KO BMMs, stained with rhodamine-phalloidin, which are untreated or following a 2 min stimulation with ICs at 37°C. b. Representative photomicrographs of TRPV2KO BMMs, stained with rhodamine-phalloidin, which are untreated, treated with KCl (50 mM), KCl + wortmannin (100 nM, PI(3)K inhibitor), KCl + adenosine (300 µM, PI4 kinase inhibitor), or KCl + 3m3FBS (10 µM, PLC activator). Scale bar, 10 µm.