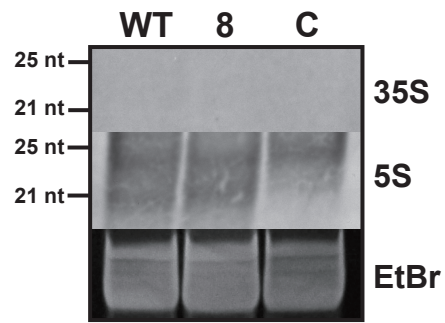
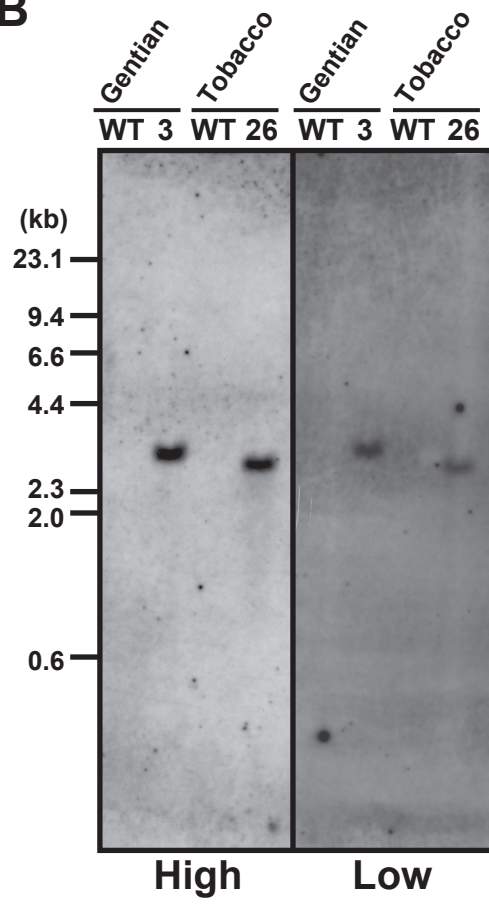


A**B**

Experimental procedures for supplementary data

Small RNA and genomic DNA analyses for 35S enhancer sequence detection

Low molecular weight RNAs were isolated from each 0.4 g F.W. of young leaf tissues of wild-type and 35S-*sGFP* gentians and tobacco plants using a High Pure miRNA Isolation Kit (Roche). The low molecular-weight RNA samples were separated by 15% PAGE at 180V, and the gel was transferred to a Biotodyne A (PALL) nylon membrane with a semidry blotter. After transfer, 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (EDC)-mediated cross-linking (Pall and Hamilton, 2008) was performed for 2h at 60 °C. Hybridization and non-radioactive detection of small RNAs were performed as described by Goto et al. (2003). DIG-labeled 35S enhancer (278-bp; -355 to -78) and 5S rRNA gene (114-bp) probes were prepared by PCR using primers as follows: for 35S enhancer, 5'-CCAAAGGGCAATTGAGACTTTT-3' and 5'-CGTCAGTGGAGATATCACATCAA-3'; for 5S, 5'-TGCGATCATAACCAGCACTAA-3' and 5'-GRTGCAACACRAGGACTTCCC-3'.

For Southern analysis, genomic DNAs of gentian and tobacco plants were isolated as described above and each 5 µg was used. Hybridization, membrane washing and detection procedures were performed using DIG-labeling system (Roche) following the supplier's instructions. Different hybridization conditions (at 42 °C and 37 °C) were tested using the DIG-labeled 35S enhancer probe as described above.

References

- Goto K, Kanazawa A, Kusaba M, Masuta C (2003) A simple and rapid method to detect plant siRNAs using nonradioactive probes. *Plant Mol Biol Rep* 21: 51-58.
- Pall GS, Hamilton AJ (2008) Improved northern blot method for enhanced detection of small RNA. *Nat Protocol* 3: 1077-1084.