SUPPLEMENTARY FIGURE LEGENDS

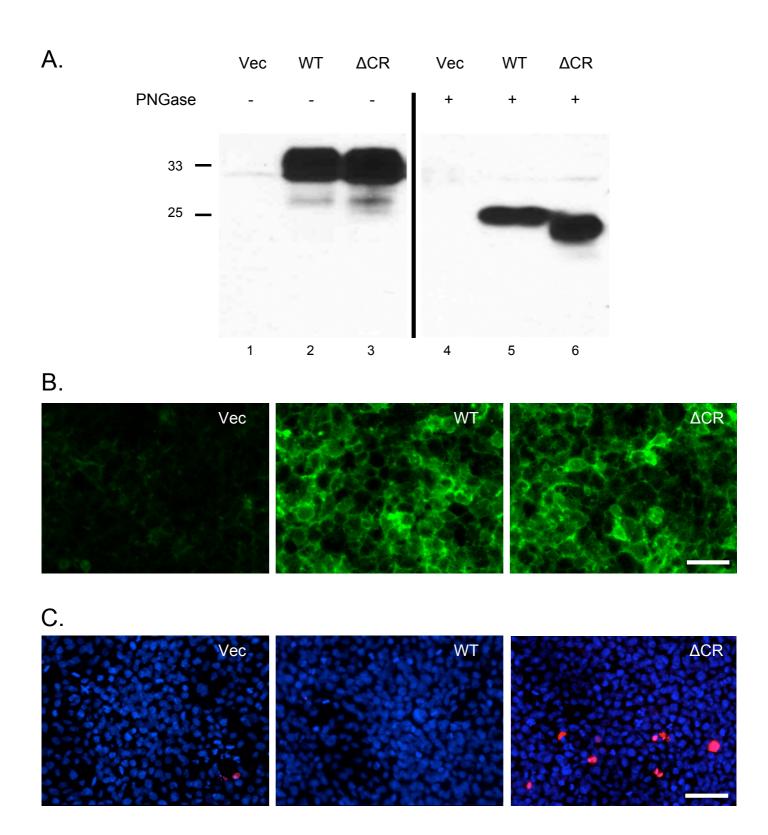
SUPPL. FIGURE S1: Both WT and ΔCR PrP are glycosylated and expressed on the surface of HEK cells; ΔCR PrP causes a small reduction in cell viability. (A) PrP expression was analyzed by Western blotting lysates of HEK cells stably transfected with empty vector (Vec), or with vector encoding WT or ΔCR PrP. Samples in lanes 4-6 were subjected to enzymatic deglycosylation with PNGase F prior to analysis. PrP was detected using anti-PrP antibody SA65. (B) Live cells of the indicated lines were stained for surface PrP with SA65 antibody. (C) Cells of the indicated lines were stained by TUNEL (red) to reveal fragmented DNA and with DAPI (blue) to reveal nuclei. Scale bars in B and C = 50 μ m.

<u>SUPPL. FIGURE S2</u>: *Drug structures*. Structures of G418, hygromycin B, Zeocin, and bleomycin are taken from the Pubchem database (http://pubchem.ncbi.nlm.nih.gov).

SUPPL. FIGURE S3: HEK cells expressing $\triangle CR$ PrP are hypersensitive to G418 and Zeocin, as measured by TUNEL. (A) Cells expressing empty vector, WT PrP, or $\triangle CR$ PrP were treated with the indicated concentrations of drug for 3 days, and were then stained by TUNEL (red) to reveal fragmented DNA and with DAPI (blue) to reveal nuclei. (B, C) Cells of the indicated lines were treated for 3 days with different concentrations of G418 (B) or Zeocin (C). The number of TUNEL-positive cells, expressed as a percentage of the number of DAPI-stained cells, was determined in 5 fields for each sample group. Bars shown means \pm SEM (n = 3 independent experiments).

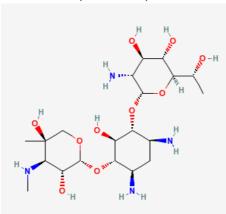
SUPPL. FIGURE S4: CHO cells expressing $\triangle CR$ PrP are hypersensitive to G418 and Zeocin, as measured by TUNEL. (A) CHO cells transiently transfected with empty vector (Vec), or with vector encoding WT or $\triangle CR$ PrP, were treated with the indicated concentrations of drug for 3 days, and were then stained by TUNEL (red) to reveal fragmented DNA and with DAPI (blue) to reveal nuclei. (B, C) Cells transfected with the indicated plasmids were treated for 3 days with the indicated concentrations of G418 (B) or Zeocin (C). The number of TUNEL-positive cells, expressed as a percentage of the number of DAPI-stained cells, was determined in 5 fields for each sample group. Bars show means \pm SEM (n = 3 independent experiments).

SUPPL. FIGURE S5: Co-expression of WT PrP suppresses Zeocin-induced DNA damage in cells expressing ΔCR PrP. (A) HEK cells expressing WT or ΔCR PrP were mock-transduced, or transduced with a recombinant lentiviruses encoding WT PrP or EGFP. Forty-eight hrs later, cells were incubated with or without Zeocin (500 µg/ml) for 60 min, after which cells were lysed, and γ -H2AX and PrP detected by Western blotting. The relative amount of PrP is given below each set of lanes. (B) Levels of γ -H2AX induced by Zeocin were determined by Odyssey analysis of blots like those shown in panel A, normalized to actin levels (not shown), and expressed as a percentage of γ -H2AX levels in the absence of drug. Bars represent means \pm SEM from 3 independent experiments. Asterisks indicate values that are significantly different from mock-transduced ΔCR or EGFP-transduced ΔCR (p<0.01).

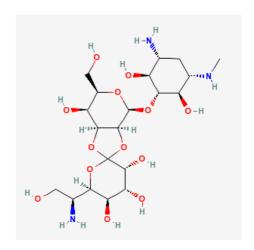


SUPPL. FIGURE S1

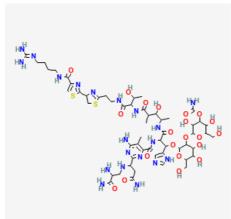
G418 (MW=496) (Geneticin)



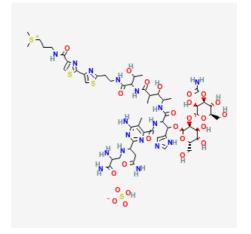
Hygromycin B (MW=528)

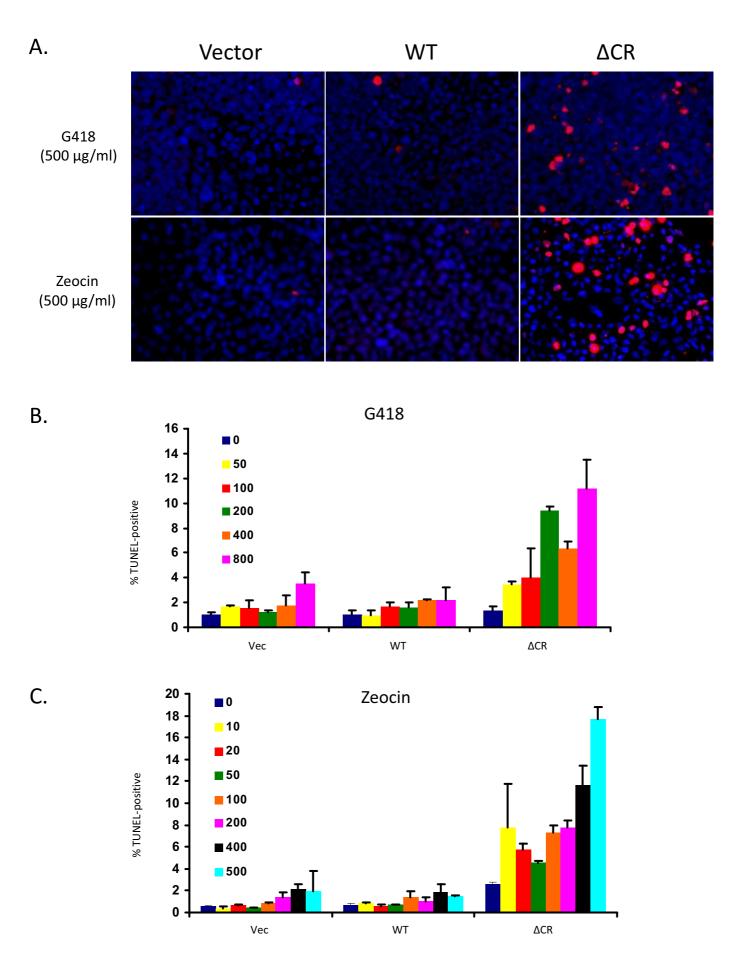


Zeocin (MW=1,428) (Phleomycin D1)

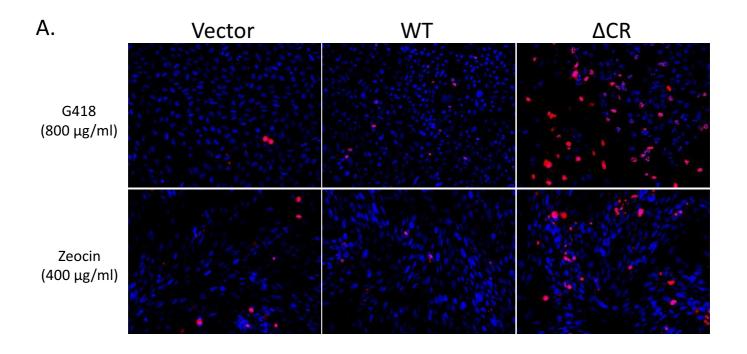


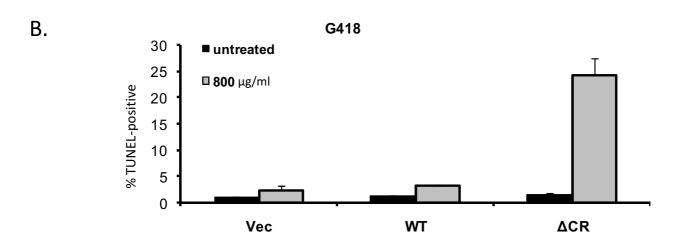
Bleomycin (MW=1,416)

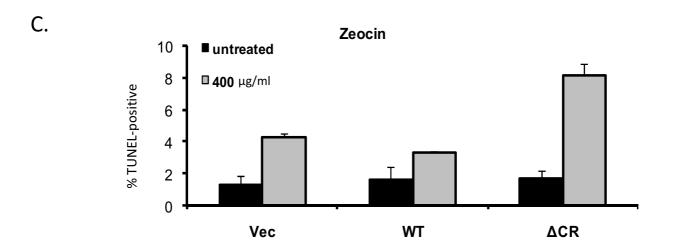




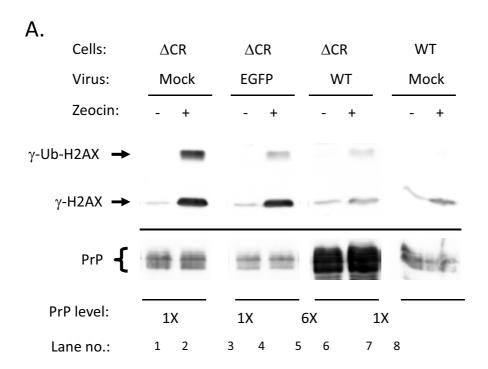
SUPPL. FIGURE S3

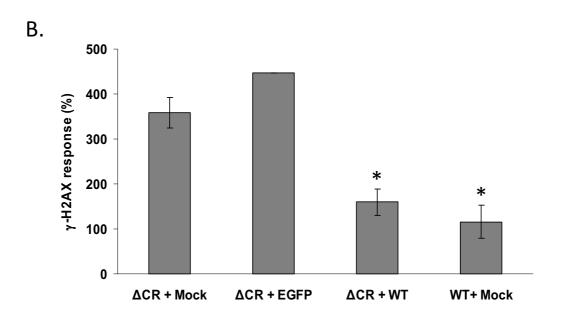






SUPPL. FIGURE S4





SUPPL. FIGURE S5