

Supplemental Data

Supplemental Experimental Procedures

Construction of a mock vector. The pTL26 plasmid was constructed by replacing the murine, erythroid ALAS active site loop-encoding sequence with a non-ALAS-encoding sequence. pGF42, a ferrochelatase expression plasmid (1), was digested with *Xba* I and *Bam* HI and the generated fragment was ligated into the ALAS expression plasmid, pGF23 (2), previously digested with the same two enzymes.

Construction of the ALAS synthetic shuffled library. The construction of the synthetic shuffled library depended on the design of five overlapping partially overlapping oligonucleotides, which contain DNA degeneracies to allow multiple mutations to be introduced in 10 of the 18 positions of the ALAS active site loop (Table S1). The primers used in this study were the following (positions with base changes are indicated in *bold*):

Fwd Primer MALAS216: 5'-CTG CTC TCC AAG CAC AGC ATC TAT **VTK** CAG **VSS** ATC
AAC-3'

Rev Primer RALAS217: 5'-GGG GGC CAA GCG **MAR** **BHK** CWS **VBB** WYY **NHB** **VBB**
CAC AGT TGG **RHD** GTT GAT **SSB** CTG-3'

Fwd Primer MALAS218: 5'-CGC TTG GCC CCC TCC CCC CAC CAC AGC CCT CAG ATG
ATG GAA AAC TTT-3'

Fwd Primer MALAS219: 5'-ATC TGT GAT CTT CTG CTC TCC AAG CAC AGC AT-3' **Rev**

Primer RALAS220: 5'-CTT CTC CAC AAA GTT TTC CAT CAT CTG A-3'

where R = 50%A + 50%G; Y = 50%C + 50%T; M = 50%A + 50%C; K = 50%G + 50%T; S = 50%C + 50%G; W = 50%A + 50%T; H = A + 25%C,T; B = 33%C + 33%G + 33%T; V = 33%A + 33%C + 33%G; D = 33%A + 33%G + 33%T; N = 25%A + 25% C + 25% G + 25%T.

An annealing reaction (20 μ L) containing 100 pmol of each of the five oligonucleotides was incubated at 65 °C for 2 min and then slowly cooled to room temperature. The annealed oligonucleotides were extended with 2 units of thermostable DNA polymerase and dNTPs, at final concentration of 55 μ M each, in a 20min reaction at room temperature.

The generated double-stranded DNA was used as template in a PCR reaction using the outermost forward and reverse primers (MALAS219 and RALAS220) and following the cycling parameters: a total of 30 cycles of 94°C for 1 minute, 55°C for 1 minute, and 72°C for 1 minute was followed by an extension of 5 minutes at 72°C. The shuffled DNA product was further amplified by PCR and using 2 primers (MALAS166 and RpBREvo2 – *sequence below*), which annealed to the 5' and 3' ends of the shuffled DNA product. The annealing sites for MALAS166 and RpBREvo2 were upstream and downstream of the sequences for the restriction endonucleases, *Xba* I and *Bam* HI, respectively. PCRs were performed using a MJ Research Minicycler and following the cycling parameters: a total of 30 cycles of 94°C for 1 minute, 55°C for 1 minute, and 72°C for 1 minute was followed by an extension of 5 minutes at 72°C. The amplified, shuffled DNA product was digested with *Xba* I and *Bam* HI, purified and ligated into pTL26, the mock ALAS expression vector, previously digested with the same two enzymes, as described in (3).

Fwd Primer MALAS166: 5'-TGC AGG CCA TAG AGG AGA CC-3'

Rev Primer RpBREvo2: 5'-TAC GAG TTG CAT GAT AA-3'

Table S1. Designed mutations for incorporation at indicated positions within the ALAS active site loop¹

<i>Position</i>	<i>WT</i>	<i>Mut1</i>	<i>Mut2</i>	<i>Mut3</i>	<i>Mut4</i>	<i>Codon</i>
423	V	I	L			VTK
425	A	S	P			VSS
428	Y	F	H	S		HDY
432	P	A	S	D	R	VVB
433	R	I	V	K	S	VDN
434	G	K				RRW
435	E	Q	T	S	D	VVB
436	E	L				SWG
437	L	R	K	M		MDV
438	L		F			YTK

¹WT denotes amino acid found in mALAS-2 active site loop. Amino acids indicated in the columns labeled *Mut1-Mut4* reflect all *naturally* occurring amino acids found in the ALAS active site loop. Codon indicates the nucleotide codon used to obtain the indicated mixture of amino acid residues. DNA degeneracies are represented in the IUB code: Y, C/T; M, A/C; K, G/T; R, G/A; S, C/G; W, A/T; V, A/C/G; H, A/C/T; D, A/G/T; B, C/G/T; N, A/C/G/T.

Table S2. Thermodynamic activation parameters of wild-type ALAS and the SS2 variant

Enzyme	Slope/(K)	E_a (kcal/mol)	ΔH^\ddagger (30°C) (kcal/mol)	ΔG^\ddagger (30°C) (kcal/mol)	ΔS^\ddagger (30°C) (cal/mol·K)
SS2	-9200	18	18	19	-4.4
WTALAS	-24000	48	48	19	-95

Figure S1

Active Site Loop Y422.....R439

1	ALAS_DELLEU	486	LLLSKHGIYVQAINY	PTVPRG-EELLRLAP	SPHH-SPQMMEFVE	KLLAAWTEVGLPLQD	-VSIACNFCRRPVH	FELMSEWERSYFGNM	573
2	ALAS_DLLLEU	486	LLLSKHGIYVQAINY	PTVPRG-EELLRLAP	SPHH-SPQMMEFVE	KLLAAWTEVGLPLQD	-VSIACNFCRRPVH	FELMSEWERSYFGNM	573
3	ALAS_HOMSAP	491	LLLSKHGIYVQAINY	PTVPRG-EELLRLAP	SPHH-SPQMMEFVE	KLLLAWTAVGLPLQD	-VSVACNFCRRPVH	FELMSEWERSYFGNM	578
4	ALAS_RATNOR	491	LLLAKHSIYVQAINY	PTVPRG-EELLRLAP	SPHH-SPQMMEFVE	KLLLAWTEVGLPLQD	-VSVACNFCRRPVH	FELMSEWERSYFGNM	578
5	ALAS_RATRAT	491	LLLAKHSIYVQAINY	PTVPRG-EELLRLAP	SPHH-SPQMMEFVE	KLLLAWTEVGLPLQD	-VSVACNFCRRPVH	FELMSEWERSYFGNM	578
6	ALAS_MUSMUS	491	LLLSKHSIYVQAINY	PTVPRG-EELLRLAP	SPHH-SPQMMEFVE	KLLLAWTEVGLPLQD	-VSVACNFCRRPVH	FELMSEWERSYFGNM	578
7	ALAS_DANRER	486	ILLEKHNIYVQAINY	PTVPRG-EELLRLAP	SPFH-NPIMMNYFAE	KLLDVWQEVGLPLNG	-PAQASCTFCDRPLH	FDLMSEWEKSYFGNM	573
8	ALAS_DANROS	486	ILLEKHNIYVQAINY	PTVPRG-EELLRLAP	SPFH-NPIMMNYFAE	KLLDVWQEVGLPLNG	-PAQASCTFCDRPLH	FDLMSEWEKSYFGNM	573
9	ALAS_OPSTAU	486	SLLEKHNIYVQAINY	PTVPRG-QELLRLAP	SPHH-HPAMMEYFVD	KLVEVWQEVGLPLNG	-PATVSCCTFCDRPLH	FDLMSEWEKSYFGNM	573
10	ALAS_HOMSPN	545	ELMSRHNIYVQAINY	PTVPRG-EELLRIAP	TPHH-TPQMNNYFLE	NLLVTWKQVGLLEKLP	-HSSAECNFCRRPLH	FEVMSEREKSYFSGM	632
11	ALAS_DELDEL	545	ELMSRHNIYVQAINY	PTVPRG-EELLRIAP	TPHH-TPQMNNYFVE	NLLATWKRVGLELKP	-HSSAECNFCRRPLH	FEVMSEREKSYFSGM	632
12	ALAS_MOUDEM	546	ELMTRHNIYVQAINY	PTVPRG-EELLRIAP	TPHH-TPQMNNYFVE	KLLVTWKRVGLELKP	-HSSAECNFCRRPLH	FEVMSEREKSYFSGM	633
13	ALAS_GALVAR	540	KLMSQHSIYVQAINY	PTVPRG-EELLRIAP	TPHH-TPQMMSYFLE	KLLATWKDVGLELKP	-HSSAECNFCRRPLH	FEVMSERERSYFSGM	627
14	ALAS_XENLAE	308	KLMDYSIYVQAINY	PTVPRG-EELLRIAP	TPHH-NPQ-----	-----	-----	-----	344
15	ALAS_OPSBET	532	LMMSHHNIYVQAINY	PTVARG-DELLRIAP	TPHH-TPEMMKYFVD	RLVQTWKEVGLLEKLP	-HSSAECTFCQQPLH	FEVMNEREKSYFSGM	619
16	ALAS_OPSPAR	532	LMMSHHNIYVQAINY	PTVARG-DELLRIAP	TPHH-TPEMMKYFVD	RLVQTWKEVGLLEKLP	-HSSAECTFCQQPLH	FEVMNEREKSYFSGM	619
17	ALAS_DANDAN	518	IMMSRYNIYVQAINY	PTVARG-EELLRIAP	TPHH-TPQMNNYFVE	KLTQTWTEVGLPLKP	-HSSAECNFCRRPLH	FEIMSEREKSYFSGM	605
18	ALAS_MYXGLU	565	ELMSRHNIYVQAINY	PTVPRG-EEMLRVVV	TPHH-TPQMNNYFVE	HLTNSWKDIGLNLRP	-HASAECNYCKMPIH	FELMSEHDQVYFDGM	652
19	ALAS_STRDRO	512	SLLEEHNIVQAINY	PTVPSG-EEKLRVAA	SPXH-TPDMMDFRVA	SLSEVWAKSGLRFNT	PICPRECEFCCKNPEK	FEELSSRERSFAEES	578
20	ALAS_DROMEL	463	VLIEQFGHVLQSIY	PTVARG-QEKLRLAP	TPFH-TFEMMNALVT	DLKKVWEMVDLSTNV	PLSPNACMFCNSESC	WHQDTSPLDLECGIPN	529
21	ALAS_LIMPOL	494	ELISMHGHIYVQAINY	PTVPRG-EEKLRVAA	TPFH-TRPMMEAFVR	DLVSVWRGLKPLPLRD	GICEKCEFCCKPLY	FEHLESRVLPCCGMR	560
22	ALAS_SEPOFF	540	DLLIKHNIVQAINY	PTVARG-EEKLRVAA	TPHH-TKEMMDHFVD	CVVVKWLEHGLTLNP	-DSTRPAEFNVKFKK	FSI-----	603
23	ALAS_GLYDIB	516	ELMEEHGIYVQAINY	PTVPRG-QELLRVAP	TPHH-TKEMMDSFVN	ATLSVFLNNNIELKS	-TCGINCLYCHQPMK	CEAFTNRERAPCDGV	586
24	ALAS_GALGAL	420	ALLEEHGLIVQAINH	PTVPRG-QELLRIAP	TPHH-SPMLENLAD	KLSECWGAUGLPLRD	-PPGPSCSSCHRPLH	LSLLSPLERDQFQVGR	508
25	ALAS_SINMEL	346	LLLDNFGIYVQAINY	PTVPRG-TERLRITP	TPMH-SDADIDLHVS	ALHSLWSRCALARAV	A-----	-----	405
26	ALAS_SMRMEL	275	LLLDNFGIYVQAINY	PTVPRG-TERLRITP	TPMH-SDADIDLHVS	ALHSLWSRCALARAV	A-----	-----	334
27	ALAS_AGRTUM	364	ILLDNHGIYVQAINY	PTVPRG-TERLRITP	TPMH-SDADIEHLVQ	ALHQLWSHCALARAV	A-----	-----	423
28	ALAS_AGRRAD	346	ILLDSHGVIYVQAINY	PTVPRG-TERLRITP	TPMH-SDADIEHLVQ	ALHQLWSHCALARAV	A-----	-----	405
29	ALAS_RHIRAD	276	ILLDSHGVIYVQAINY	PTVPRG-TERLRITP	TPMH-SDADIEHLVQ	ALHQLWSHCALARAV	A-----	-----	335
30	ALAS_RHOPSE	344	ELINRYGIYVQAINY	PTVPRG-TERLRITP	SPQH-TDADIEHLVQ	ALSEIWARVGLAKAA	-----	-----	403
31	ALAS_RHOPSE	346	ALLARHAIYVQAINY	PTVPRG-QERFRITP	TPFH-TTSHMEALVE	ALLAVGRDLGWAMSR	RAA-----	-----	407
32	ALAS_EUGGRA	343	LLLKRFGIYVQAINY	PTVDVG-TERLRITP	SPVH-TNEHMATLIT	ALLQVWEEELGLPLRP	PVFDTEGYPEVEAAE	WLPNASAMWRDASPHH	431
33	ALAS_BRAJAP	348	LLLEEHLIYVQAINY	PTVAKG-SERLRITP	SPYH-DDGLIDQLAE	ALLQVWDRGLGLPLKQ	KSLAAE-----	-----	409
34	ALAS_BRAJAP	275	LLLEEHLIYVQAINY	PTVAKG-SERLRITP	SPYH-DDGLIDQLAE	ALLQVWDRGLGLPLKQ	KSLAAE-----	-----	339
35	ALAS_BRUMEL	345	RLLEEVHGIYVQAINY	PTVPRG-TERLRITP	SPLH-DKLDIDGLKD	ALLEVWNEELGIPFAE	PSAPQAANSDRIIPL	MVSKAGG-----	425
36	ALAS_ZYMMOB	287	ILLNEYGAIYVQAINF	PTVPRG-TERLRITP	GPTH-NEAMLRELTD	SLVAIWHRLDMRFAA	-----	-----	345
37	ALAS_RHOCAP	348	MLLSDYGVYVQAINF	PTVPRG-TERLRITP	SPVH-DLKQIDGLVH	AMDLLWARCA-----	-----	-----	401
38	ALAS_RHOCAP	278	MLLSDYGVYVQAINF	PTVPRG-TERLRITP	SPVH-DLKQIDGLVH	AMDLLWARCA-----	-----	-----	331
39	ALAS_PARDEN	292	MLLADFSIYVQAINF	PTVPRG-TERLRITP	SPVH-DPKQIDHLVK	AMDSLWSQCKLNRST	SAA-----	-----	339
40	ALAS_PARZEA	360	MLLADFSIYVQAINF	PTVPRG-TERLRITP	SPVH-DPKQIDHLVK	AMDSLWSQCKLNRST	SAA-----	-----	409
41	ALAS_RHOPSE	364	MLLIHFGIYVQAINF	PTVPRG-TERLRITP	SPVH-DSGMIDHLVK	AMDVLWQHICALNRAE	VVA-----	-----	407
42	ALAS_EMENID	510	KLLEEHLIYVQAINY	PTVPRG-EERLRITP	TPGH-TQELRDHLVE	AVNTVWVDLGIKRAS	DWKAMGGFVGVGVEA	AELENQPIWTDACLN	598
43	ALAS_EMENID	440	KLLEEHLIYVQAINY	PTVPRG-EERLRITP	TPGH-TQELRDHLVE	AVNTVWVDLGIKRAS	DWKAMGGFVGVGVEA	AELENQPIWTDACLN	528
44	ALAS_ASPNID	440	KLLEEHLIYVQAINY	PTVPRG-EERLRITP	TPGH-TQELRDHLVE	AVNTVWVDLGIKRAS	DWKAMGGFVGVGVEA	AELENQPIWTDACLN	528
45	ALAS_ASPORI	508	KLLEEHLIYVQAINY	PTVPRG-EERLRITP	TPGH-IKEHRDHLVQ	AVQTVWNEELGIKRST	DWEAQGGFVGVGVDG	AAEENQPIWVDVQLG	596
46	ALAS_NEUCRA	334	KLLNDHQIYVQAINY	PTVPRG-QERLRITP	TPGH-TKQFRDHLVA	ALDSIWTELGIKRST	DWAAEAGGFVGVGVEA	A-EPVAPLWTDQQLG	421

47	ALAS_GIBFUJ	393	MLLN	YGI	YVQAIN	Y	PTV	FVG	QERLR	RVTP	TPGH	IKEYR	DQLVE	AIDEI	WTRLD	DIKRTS	DWAAE	GGF	IGV	GEQD	--NV	QEPL	WTDK	QLN	479	
48	ALAS_YARLIP	404	LLL	TKHQI	YVQAIN	F	PTV	FIG	QERLR	RVTP	TPGH	HEGLC	DDELVA	ALEDV	VQEL	DLKRVE	DWTAE	GGC	CGV	GEV	--E	VEPL	WSEE	QLS	489	
49	ALAS_CANALB	451	LLL	NKHD	YVQAIN	F	PTV	FIG	EERLR	RVTP	TPGH	GP	ELSKQ	LVE	AVDSV	FTEL	NLNRIN	DWKKL	GGLV	GVG	VEG	-AAK	VEHI	WTEE	QLA	538
50	ALAS_CANALB	381	LLL	NKHD	YVQAIN	F	PTV	FIG	EERLR	RVTP	TPGH	GP	ELSKQ	LVE	AVDSV	FTEL	NLNRIN	DWKKL	GGLV	GVG	VEG	-AAK	VEHI	WTEE	QLA	468
51	ALAS_DEBHAN	392	LLL	DKYNI	YVQAIN	F	PTV	FIG	QERLR	RVTP	TPGH	GP	ELSNQ	LIG	ALDSV	FNEL	SLSRIG	DWEGK	GGLC	GVG	EPD	-IEPI	EHI	WTEE	QLA	479
52	ALAS_SACCR	435	IL	INKHQI	YVQAIN	F	PTV	ARG	TERLR	RVTP	TPGH	TNDL	SDILIN	AVDDV	FNEL	QLPRVR	DWESQ	GGLL	GVG	ESG	-FVE	ESNL	WTSS	QLS	522	
53	ALAS_SACCAS	365	IL	INKHQI	YVQAIN	F	PTV	ARG	TERLR	RVTP	TPGH	TNDL	SDILIN	AVDDV	FNEL	QLPRVR	DWESQ	GGLL	GVG	ESG	-FVE	ESNL	WTSS	QLS	452	
54	ALAS_CANGLA	347	IL	MEKHRI	YVQAIN	F	PTV	SRG	TERLR	RVTP	TPGH	TNDL	SDILIA	AVDDV	FNEL	QLPRIR	DWEMQ	GGLL	GVG	DKN	-FVPE	PNL	WTEE	QLS	434	
55	ALAS_KLULAC	387	IL	MDKHRI	YVQAIN	F	PTV	ARG	TERLR	RVTP	TPGH	TNDL	SDILMD	ALEDV	VSTL	QLPRVR	DWEAQ	GGLL	GVG	DPN	-HVP	QPNL	WTKD	QLT	474	
56	ALAS_EREGOS	373	IL	MEKHRI	YVQAIN	F	PTV	PRG	TERLR	RVTP	TPGH	TNDL	SDVLLD	AMDDV	VKTL	QLPRVS	DWAAH	GGLL	GVG	EPD	-YVPE	ANL	WTEE	EQMS	460	
57	ALAS_AGABIS	460	KLL	SEHDI	YVQAIN	Y	PTV	ARG	EERLR	ITV	TPRH	TMEQ	MEGLIR	SLNQV	F	EELNINR	LS	DWKL	AGGR	AGV	GIPG	AADDV	QPI	WTDE	QIG	548
58	ALAS_AGADIV	390	KLL	SEHDI	YVQAIN	Y	PTV	ARG	EERLR	ITV	TPRH	TMEQ	MEGLIR	SLNQV	F	EELNINR	LS	DWKL	AGGR	AGV	GIPG	AADDV	QPI	WTDE	QIG	478
59	ALAS_SCHPOM	475	SLL	HDHNI	YVQAIN	F	PTV	SVG	TERLR	RVTP	TPAHN	TEHY	VQSLTN	AMNDV	VSKF	NINRID	GWEKR	GIDV	GR	LCKF	PVLP	PFTT	TH	-----	558	
60	ALAS_SCHMIK	405	SLL	HDHNI	YVQAIN	F	PTV	SVG	TERLR	RVTP	TPAHN	TEHY	VQSLTN	AMNDV	VSKF	NINRID	GWEKR	GIDV	GR	LCKF	PVLP	PFTT	TH	-----	488	
61	ALAS_RICPRO	342	MLL	NEYGI	YVQHIN	F	PTV	PRG	TERLR	RVTP	TPAH	TDK	MINDLST	ALVHI	F	DEL	DI	ELSS	AKELN	KEV	R	LHLIA	-----	-----	414	
62	ALAS_RICTYP	342	MLL	NEYGI	YVQHIN	F	PTV	PRG	TERLR	RVTP	TPAH	TDK	MINDLST	ALVHI	F	DEL	DI	ELSS	TKELN	KEV	R	LHLIA	-----	-----	414	
63	ALAS_RICPRO	272	MLL	NEYGI	YVQHIN	F	PTV	PRG	TERLR	RVTP	TPAH	TDK	MINDLST	ALVHI	F	DEL	DI	ELSS	AKELN	KEV	R	LHLIA	-----	-----	344	
64	ALAS_RICCON	354	MLL	NEYGI	YVQHIN	F	PTV	PRG	TERLR	RVTP	TPAH	TDK	MINDLSV	ALVHI	F	DEL	DI	ELSS	AKELN	EEV	R	LNLIA	-----	-----	426	
65	ALAS_CHRVIO	354	RL	LEFDI	YVQPIN	Y	PSV	PRG	GERFR	LTV	GPRR	SHEE	IQR	FVA	ALKH	CLA	-----	-----	-----	-----	-----	-----	-----	-----	403	
	A8	491	LLL	SKHSI	YVQAIN	Y	PTV	PRG	EEL	LRLAP	SPHH	SPQ	MMENFVE	KLLL	AWTE	VGL	PLQD	-VSVA	ACNF	CHRP	VH	FEL	MSEW	ERSY	FGNM	578
	D8	491	LLL	SKHSI	YVQAIN	Y	PTV	PRG	EE	QLRLAP	SPHH	SPQ	MMENFVE	KLLL	AWTE	VGL	PLQD	-VSVA	ACNF	CHRP	VH	FEL	MSEW	ERSY	FGNM	578
	G7	491	LLL	SKHSI	YVQAIN	Y	PTV	PRG	EEL	LRLAP	SPHH	SPQ	MMENFVE	KLLL	AWTE	VGL	PLQD	-VSVA	ACNF	CHRP	VH	FEL	MSEW	ERSY	FGNM	578
	F1	491	LLL	SKHSI	YVQAIN	Y	PTV	PKK	QE	QLRLAP	SPHH	SPQ	MMENFVE	KLLL	AWTE	VGL	PLQD	-VSVA	ACNF	CHRP	VH	FEL	MSEW	ERSY	FGNM	578
	F10	491	LLL	SKHSI	YVQAIN	Y	PTV	Q	EN	LRLAP	SPHH	SPQ	MMENFVE	KLLL	AWTE	VGL	PLQD	-VSVA	ACNF	CHRP	VH	FEL	MSEW	ERSY	FGNM	578
	A4	491	LLL	SKHSI	YVQAIN	Y	PTV	RK	NEL	LRLAP	SPHH	SPQ	MMENFVE	KLLL	AWTE	VGL	PLQD	-VSVA	ACNF	CHRP	VH	FEL	MSEW	ERSY	FGNM	578
	F3	491	LLL	SKHSI	YVQAIN	Y	PTV	PHN	KEK	LRLAP	SPHH	SPQ	MMENFVE	KLLL	AWTE	VGL	PLQD	-VSVA	ACNF	CHRP	VH	FEL	MSEW	ERSY	FGNM	578
	H1	491	LLL	SKHSI	YVQAIN	Y	PTV	NIE	KEK	LRLAP	SPHH	SPQ	MMENFVE	KLLL	AWTE	VGL	PLQD	-VSVA	ACNF	CHRP	VH	FEL	MSEW	ERSY	FGNM	578
	SS2	491	LLL	SKHSI	YVQAIN	Y	PTV	EIN	QE	KLRLAP	SPHH	SPQ	MMENFVE	KLLL	AWTE	VGL	PLQD	-VSVA	ACNF	CHRP	VH	FEL	MSEW	ERSY	FGNM	578

Fig S1. Alignment of sixty-five different species specific ALAS active site loop sequences with the hyperactive variants isolated in this study. The naturally occurring amino acid sequences were obtained from public databases (NCBI) using a BLAST search and aligned using CLUSTAL W (4). The 10 positions within the 18-amino acid sequence targeted for mutagenesis are high-lighted in cyan, and changes observed in hyperactive variants are high-lighted in magenta. The amino acid numbering in red refers to that of murine erythroid ALAS (mALAS-2). Represented proteins are: ALAS_DELLEU: *Delphinapterus leucas* ALAS (20138447); ALAS_DLLLEU: *Delphinapterus leucas* cook inlet subspecies 2 (5281116); ALAS_HOMSAP, *Homo sapiens* ALAS erythroid (4557299); ALAS_RATNOR, *Rattus norvegicus* erythroid ALAS (51980582); ALAS_RATRAT, *Rattus rattus* erythroid ALAS (6978485); ALAS2_MUSMUS, *Mus musculus* erythroid ALAS (33859502); ALAS_DANRER, *Danio rerio* ALAS (18858263); ALAS_DANROS, *Danio roseus* ALAS (20138448); ALAS_OPSTAU, *Opsanus tau* ALAS (1170202); ALAS_ORYLAT, *Oryzias latipes* ALAS (49022596); ALAS_HOMSPN, *Homo sapiens* erythroid ALAS (4502025); ALAS_DELDEL, *Delphinus delphis* erythroid ALAS (20138445); ALAS_MOUDOM, *Mus musculus domesticus* erythroid ALAS (23956102); ALAS_GALVAR, *Gallus varius* erythroid ALAS (122821); ALAS_XENLAE, *Xenopus laevis* erythroid ALAS (44968228); ALAS_OPSEBET, *Opsanus beta* ALAS (1170206); ALAS_OPSPAR, *Opsanus pardus* ALAS (532630); ALAS_DANDAN, *Danio danglia* ALAS (32451642); ALAS2_MYXGLU, *Myxine glutinosa* erythroid ALAS (4433550); ALAS1_BRALAN, *Branchiostoma lanceolatum* ALAS 1 (28630217); ALAS1_STRDRO, *Strongylocentrotus droebachiensis* ALAS 1 (4433548); ALAS1_DROMEL, *Drosophila melanogaster* ALAS 1 (2330591); LAS1_LIMPOL, *Limulus polyphemus* ALAS 1 (4433540); ALAS1_SEPOFF, *Sepia officinalis* ALAS 1 (4433546); ALAS2_GLYDIB, *Glycera dibranchiate* ALAS 2 (4433544); ALAS2_GALGAL, *Gallus gallus gallus* ALAS 2 (1170201); ALAS_SINMEL, *Sinorhizobium meliloti* 1021 ALAS (15966742); ALAS_SMRMEL, *Sinorhizobium meliloti* ALAS (18266808); ALAS_AGRUM, *Agrobacterium tumefaciens* ALAS (889869); ALAS_AGRAD, *Agrobacterium radiobacter* ALAS (95069); ALAS_AGRUM, *Agrobacterium tumefaciens* ALAS (122818); ALAS_RHOPAL,

Rhodopseudomonas palustris ALAS (4001678); ALAS_RHOSPO, *Rhodobacter sporagenes* ALAS (541302); ALAS_EUGGRA, *Euglena gracilis* ALAS (12620813); ALAS_BRAELK, *Bradyrhizobium elkanii* ALAS (66534); ALAS_BRAJAP, *Bradyrhizobium japonicum* ALAS (30179569); ALAS_BRUMEL, *Brucella melitensis* ALAS (25286547); ALAS_ZYMMOB, *Zymomonas mobilis* ALAS (4511998); ALAS_RHOGLU, *Rhodobacter gluconicum* ALAS (97435); ALAS_RHOCAP, *Rhodobacter capsulatus* ALAS (122828); ALAS_PARDEN, *Paracoccus denitrificans* ALAS (1170207); ALAS_PARZEA, *Paracoccus zeaxanthinifaciens* ALAS (537435); ALAS_RHOSPH, *Rhodobacter sphaeroides* ALAS (541301); ALAS_EMENID, *Emericella nidulans* ALAS (418756); ALAS_EMCNID, *Emericella nidulans* ALAS (585244); ALAS_ASPNID, *Aspergillus nidulans* ALAS (40745239); ALAS_ASPORY, *Aspergillus oryzae* ALAS (5051989); ALAS_NEUCRA, *Neurospora crassa* ALAS (52782908); ALAS_GIBFUJ, *Gibberella fujikuroi* ALAS (15721883); ALAS_YARLIP, *Yarrowia lipolytica* ALAS (52782857); ALAS_CANBER, *Candida berate* ALAS (7493758); ALAS_CANALB, *Candida albicans* ALAS (10720014); ALAS_DEBHAN, *Debaryomyces hansenii* ALAS (52782855); ALAS_SACCER, *Saccharomyces cerevisiae* ALAS (6320438); ALAS_SACCAS, *Saccharomyces castellii* ALAS (122831); ALAS_CANGLA, *Candida glabrata* ALAS (52782865); ALAS_KLULAC, *Kluyveromyces lactis* ALAS (52788271); ALAS_EREGOS, *Eremothecium gossypii* ALAS (52782894); ALAS_ASPBIS, *Agaricus bisporus* ALAS (1679599); ALAS_AGADIV, *Agaricus divoniensus* ALAS (2492846); ALAS_SCHPOM, *Schizosaccharomyces pombe* ALAS (7492336); ALAS_SCHMIK, *Schizosaccharomyces mikatae* ALAS (52782853); ALAS_RICCON, *Rickettsia conorii* ALAS (7433712); ALAS_RICTYP, *Rickettsia typhi* ALAS (51474008); ALAS_RICPRO, *Rickettsia prowazekii* ALAS (6225494); ALAS_RICRIC, *Rickettsia rickettsia* ALAS (2528635); ALAS_CHRVIO, *Chromobacterium violaceum* ALAS (34102112).

Figure S2

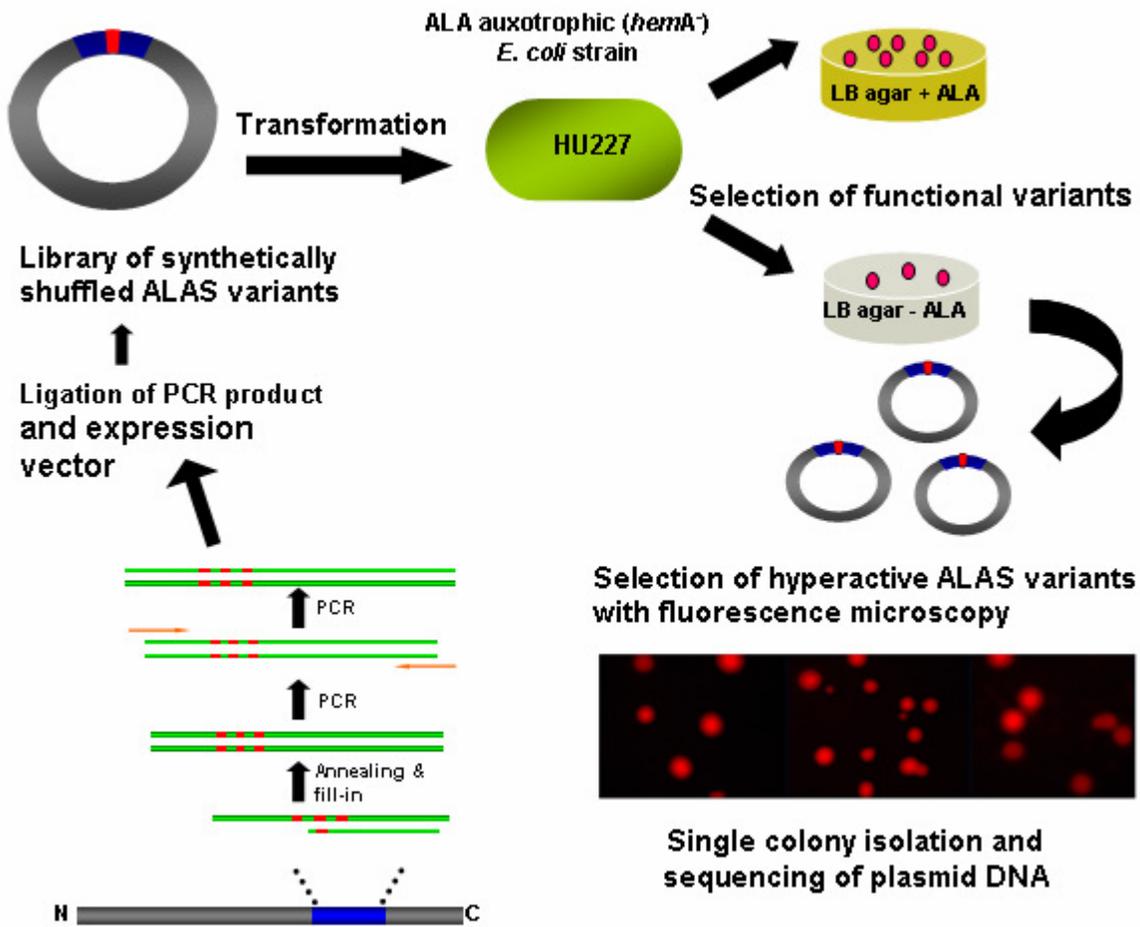
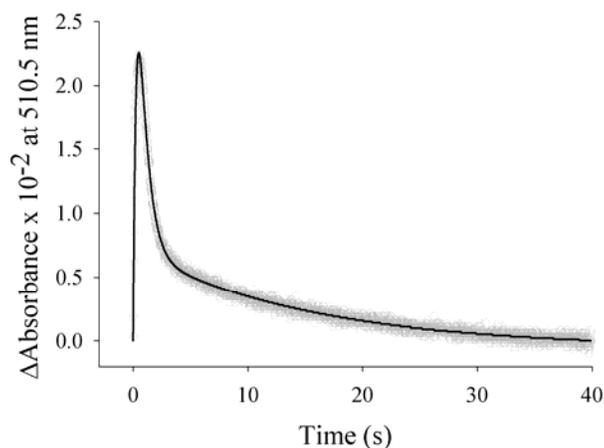


FIG S2. The generation and screening of the library.

A library of over 100,000,000 possible ALAS variants was constructed with PCR using a series of degenerate mixed base oligonucleotides. The PCR product was ligated into an expression vector. The resulting plasmids were transformed into *Escherichia coli* strain HU227 and plated on LB + ampicillin agar with and without ALA. The colonies that grew in the absence of ALA were identified as functional variants. Functional variants were screened for porphyrin overproduction by fluorescence microscopy.

Figure S3



Time course for the reaction of wild-type ALAS-glycine complex with succinyl-CoA under single turnover conditions. The reaction was monitored by following the absorbance changes at 510 nm, which was previously assigned to the quinonoid intermediate.

REFERENCES

1. Ferreira, G. C. (1994) *J. Biol. Chem.* **269**, 4396-4400
2. Ferreira, G. C., and Dailey, H. A. (1993) *J. Biol. Chem.* **268**(1), 584-590
3. Gong, J., and Ferreira, G. C. (1995) *Biochemistry* **34**(5), 1678-1685
4. Thompson, J. D., Higgins, D. G., and Gibson, T. J. (1994) *Nucleic Acids Res.* **22**, 4673-4680