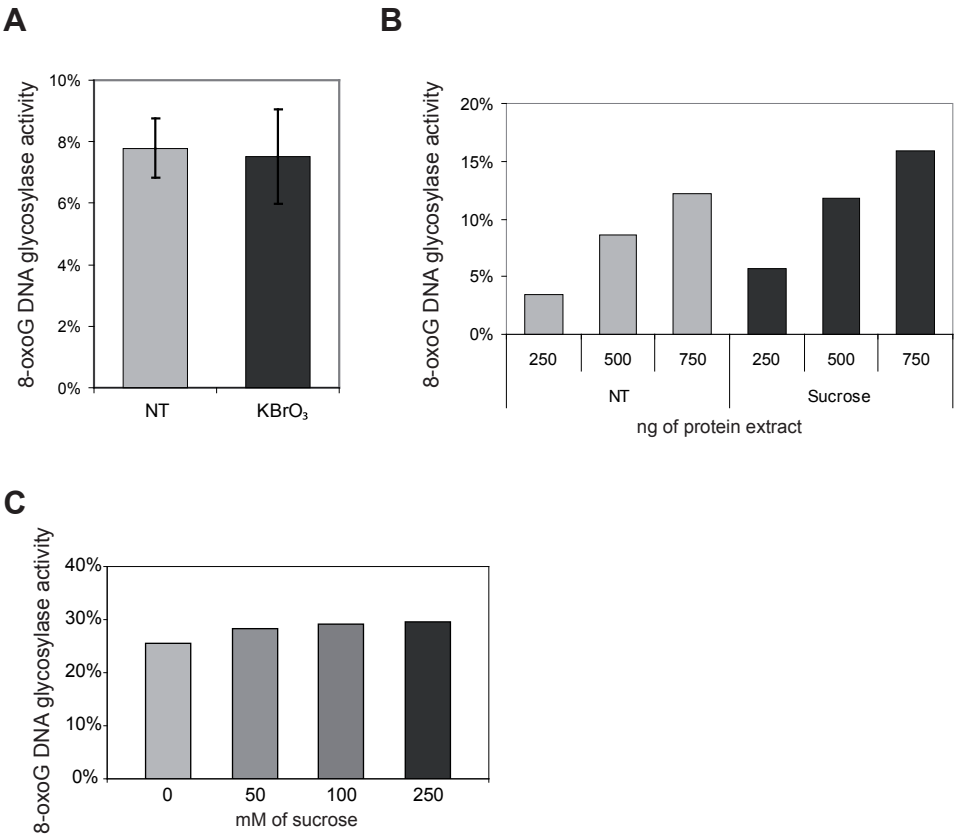
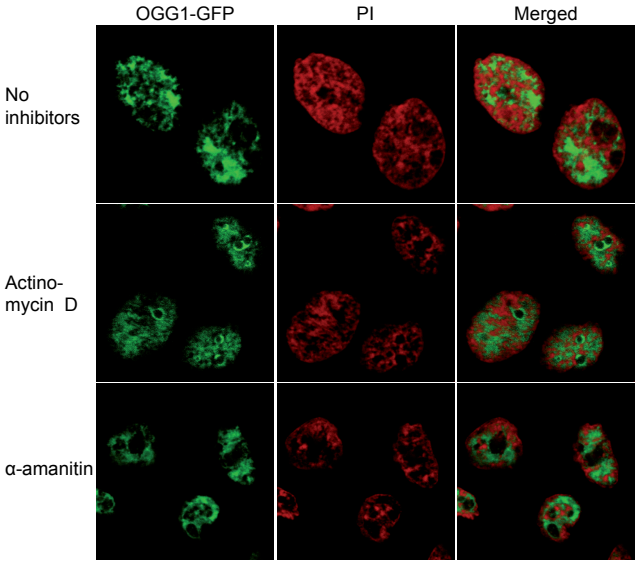


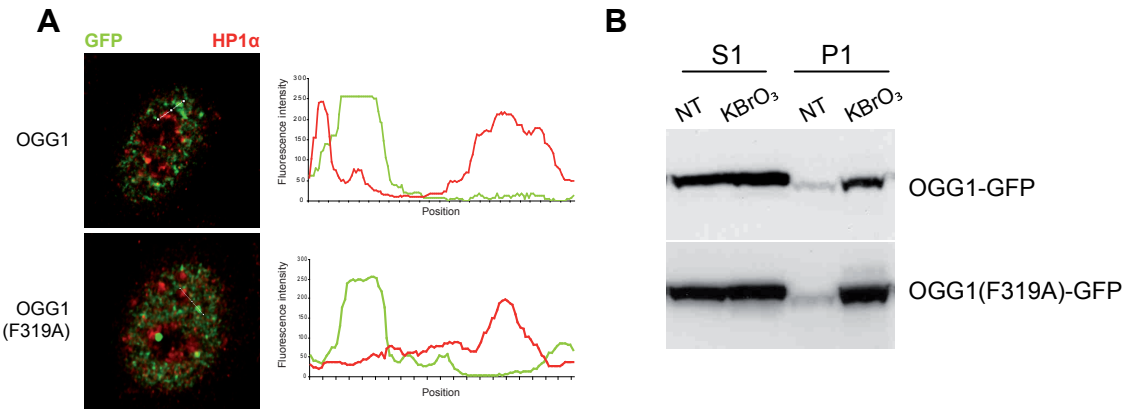
Supplementary figure 1.

Potassium bromate or sucrose incubation do not inhibit OGG1 activity. (A) HeLa cells were treated with $KBrO_3$ and allowed to recover for 3h in fresh medium. 8-oxoG DNA glycosylase activity was quantified in 5 µg of total cell extracts and compared to extracts from control HeLa cells. (B) OGG1-GFP cells were pre-incubated for 3h with 250 mM sucrose prior to extraction. The indicated amounts of cell extract were tested for 8-oxoG DNA glycosylase activity and compared to that of non-treated cells. (C) 500 ng of total protein extract from non-treated cells were analysed for its 8-oxoG DNA glycosylase activity in buffer containing 0, 50, 150 or 250 mM sucrose. Values represent an average of two experiments.



Supplementary figure 2.

Active transcription is not required for the recruitment of OGG1-GFP to the chromatin fraction. After potassium bromate treatment, transcription was blocked by either α-amanitin (100 μg/ml) or actinomycin D (2 μg/ml) during the 3 hours recovery period. After that time, cells were pre-extracted with CSK buffer and DNA was stained with Propidium Iodide-RNase (red).



Supplementary figure 3.

The OGG1 mutant F319A, unable to recognise 8-oxoG, is also recruited to the chromatin fraction. (A) Cells stably expressing OGG1-GFP or the mutant protein OGG1(F319A)-GFP were treated with $KBrO_3$ and allowed to recover in fresh medium for 3 hours before CSK washing and fixation. An antibody against HP1α was used to visualize heterochromatin (red). Positions of the line scans used for the plot profile are indicated in the images. The plot profile indicates the exclusion between the green and the red signals (B) Western blot with an anti-GFP antibody showing the OGG1-GFP and OGG1(F319A)-GFP accumulation in the chromatin fraction (P1) after $KBrO_3$ treatment compared to untreated cells (NT). S1 corresponds to the soluble fraction.