Glycine-rich RNA-binding proteins are functionally conserved in *Arabidopsis thaliana* and *Oryza sativa* during cold adaptation process

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Gene	Primer (5' to 3')
For real-time RT-PCR	
OsGRP1	Forward; GTTCAATGCTGCTCGTCTGATGTCC
	Reverse; ACCCGAGCTTCAGTCACATCCC
OsGRP2	Forward; GAGATGCCCAACCACGTCTTCC
	Reverse; TGTGGTATCTGTGGTATCGAGGTCG
OsGRP3	Forward; GACTCCAAGATCATCAACGACAGG
	Reverse; GTCGAGCTCCTTGCCGTTCATG
OsGRP4	Forward; GAAGCCGCCAATAAAGCAAAGG
	Reverse; TGTCGCTCTTGGACTGGAATCTG
OsGRP5	Forward; GGTGGGACAACGACTACAATGGC
	Reverse; TTGGAGAAGGAGTCCTTGAGGGAG
OsGRP6	Forward; AACATCAGGGTTAACACTGCCAATG
	Reverse; ACCACTGTAGCCACCACCACCATAG
pBC121	Forward; TTGACTGGCTTTCCAACAACATTGC
	Reverse; TTTGTGAGAGGCAACCTATGTGTGG
DREB1A	Forward; GGGATCAAGCAGGAGATGAGCG
	Reverse; TGCCTCGTCTCCCTGAACTTGG
OsActin	Forward; TGCGATAATGGAACTGGTATGG
	Reverse; AAGACAGCCCTGGGCGCATCA
For RT-PCR	
<u>101 KI 1 CK</u>	
OsGRP1	Forward; CGGTAGCTTCCTCAGGCACAG
	Reverse; CCTCTTGACCTTCCAGATTCTCTG
OsGRP4	Forward; CAAATTCACAAAACATGGC
	Reverse; TTCAAACACAGCAGGTAGG
OsGRP6	Forward; ATGGGAATAGCCAATAAG
	Reverse; TACACTATCGGCGCATAC
AtActin	Forward; CAGAGCGGGAAATTGTAAGAG
	Reverse; CCTTTCAGGTGGTGCAACGAC

Supplemental Table S1. Gene-specific primer pairs used in the RT-PCR experiments

Supplemental Figure Legend

Supplemental Figure S1. Alignment of the amino acid sequences of OsGRPs. The alignment was made using the ClustalW program. Gaps in the sequences are indicated by dashes. The positions of ribonucleoprotein (RNP) 1 and 2 regions are indicated by boxes. The arbitrary gene number for each OsGRP is indicated in parenthesis next to the corresponding accession numbers.

Supplemental Figure S2. Growth of BX04 *E. coli* mutant cells expressing OsGRPs at 37° C. The diluted cultures (10^{-1} to 10^{-5} dilution) of the BX04 cells harboring each OsGRP, CspA (positive control), or pINIII vector (negative control) were spotted on LB-agar plates and incubated at 37° C. The pictures were taken 1 day after incubation.

Supplemental Figure S3. Purification of recombinant proteins and nucleic acid-melting activities of OsGRPs. (A) The OsGRPs were cloned into pGEX-4T-3 vector, and *E. coli* BL21 cells harboring each construct were cultured in the presence of 0.4 mM IPTG. The GST-OsGRP fusion proteins as well as GST (negative control) and CspA (positive control) were purified and analyzed via SDS-PAGE. (B) For DNA-melting assay, the fluorescence of a molecular beacon was monitored as the recombinant GST fusion proteins (10 to 30 μ g) were added.

Supplemental Figure S4. Confirmation of transgenic *Arabidopsis* plants expressing OsGRP. Expression of (A) OsGRP1, (B) OsGRP4, and (C) OsGRP6 in *grp7* mutant was verified in the transgenic lines (C1-5, C1-7 and C1-9 for OsGRP1; C4-2, C4-7 and C4-11 for OsGRP4; C6-4 and C6-5 for OsGRP6) by RT-PCR analysis. Actin was used as a reference to show that

equal amounts of RNA were used in the analysis.

Supplemental Figure S5. Cellular localization of OsGRP1 and OsGRP4. Confocal image of the leaf and root showing the localization of OsGRP1-GFP and OsGRP4-GFP fusion proteins in the chloroplast and in the nucleus, respectively. Bar = $50 \mu m$.



Supplemental Fig. S1



Supplemental Fig. S2







Supplemental Fig. S4



Supplemental Fig. S5